ARCTIC DATA COMPILATION AND APPRAISAL

VOLUME 9

Beaufort Sea: Biological Oceanography – Bacteria, Plankton, Epontic Community 1914 to 1985

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Canadian Data Report Of Hydrography and Ocean Sciences

These reports provide a medium for the documentation and dissemination of data in a form directly useable by the scientific and engineering communities.

Generally, the reports will contain raw and/or analyzed data but will not contain interpretations of the data. Such compilations will commonly have been prepared in support of work related to the programs and interests of the Ocean Science and Surveys (OSS) sector of the Department of Fisheries and Oceans.

Data Reports are produced regionally but are numbered and indexed nationally. Requests for individual reports will be fulfilled by the issuing establishment listed on the front cover and title page. Out of stock reports will be supplied for a fee by commercial agents.

Regional and headquarters establishments of Ocean Science and Surveys ceased publication of their various report series as of December 1981. A complete listing of these publications and the last number issued under each title are published in the *Canadian Journal of Fisheries and Aquatic Sciences,* Volume 38: Index to Publications 1981. The current series began with Report Number 1 in January 1982.

Rapport statistique canadien sur l'hydrographie et les sciences océaniques

Ces rapports servent de véhicule pour la compilation et la diffusion des données sous une forme directement utilisable par les scientifiques et les techniciens.

En général, les rapports contiennent des données brutes ou analysées mais ne fournissent pas d'interprétations des données. Ces compilations sont préparées le plus souvent à l'appui de travaux reliés aux programmes et intérêts du service des Sciences et Levés océaniques (SLO) du ministère des Pêches et des Océans.

Les rapports statistiques sont produits à l'échelon régional mais sont numérotés et placés dans l'index à l'échelon national. Les demandes de rapports seront satisfaites par l'établissement auteur dont le nom figure sur la couverture et la page de titre. Les rapports épuisés seront fournis contre rétribution par des agents commerciaux.

Les établissements des Sciences et Levés océaniques dans les régions et à l'administration centrale ont cessé de publier leurs diverses séries de rapports depuis décembre 1981. Vous trouverez dans l'index des publications du volume 38 du *Journal canadien des sciences halieutiques et aquatiques,* la liste de ces publications ainsi que le dernier numéro paru dans chaque catégorie. La nouvelle série a commencé avec la publication du Rapport n° 1 en janvier 1982.

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PREFACE

These catalogues are cooperatively produced by the Scientific Information Division at the Institute of Ocean Sciences and the Environmental Secretariat at the Freshwater Institute. Joint government and industry contract projects have catalogued marine data sets, their focus being mainly on oceanography and fisheries. Data quality appraisals are included to assist in establishing the usefulness of given data for particular analyses or purposes. The ratings also determine the confidence that can be placed on interpretations incorporating those data.

The appraisals will assist in establishing priorities for incorporating the most useful data in the national Marine Environmental Data Service (MEDS) archives. Additional uses of the catalogue include provision of the best available resume of marine data sources for research planning, environmental assessments, land use planning, regulatory approvals and operational procedures.

In the past, the pace of offshore development activity has emphasized the need to review the sufficiency and suitability of available scientific information for design, regulatory and planning purposes. The review is a three-stage process: (1) compilation and appraisal of the existing data sets; (2) analysis of the suitability of existing data sets for contributing answers to questions of concern; and (3) analysis and interpretation of data and estimation of scientific confidence in the answer to the particular question. This catalogue of the bacteria, plankton and epontic community of the southeastern Beaufort Sea represents part of the results of the first stage.

Brian Smiley and Larry de March
Scientific Editors
Arctic Data Compilation and Appraisal Series

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ABSTRACT

Woods, S.M. and B.D. Smiley. 1987. Arctic Data Compilation and Appraisal. Volume 9 . Beaufort Sea: Biological Oceanography - Bacteria, Plankton, Epontic Community. Can. Data Rep. Hydrogr. Ocean Sci. 5:394 p.

This volume is one of a group of catalogues designed to compile and appraise marine data sets for the Canadian Arctic. For ease of reference, the group is organized with its subject matter divided into three disciplines: physics, chemistry, and biology. The Arctic is arbitrarily divided into seven geographical areas to include, where possible, major oceanographic regions. The format is structured to facilitate comparison between subjects and regions. With such a large undertaking it is not possible to provide all reports at once. Therefore, catalogues which are presently available in the series are indicated on the inside back cover of each volume.

Data collection is a continuing process and further updates of the catalogues are planned. Readers are invited to submit corrections and additions by writing to the issuing establishment. These corrections will be incorporated in on-line computerized data set listings; they will be continuously available upon request.

RESUME

Woods, S.M. and B.D. Smiley. 1987. Arctic Data Compilation and Appraisal. Volume 9 . Beaufort Sea: Biological Oceanography - Bacteria, Plankton, Epontic Community. Can. Data Rep. Hydrogr. Ocean Sci. 5:394 p.

Le présent volume fait partie d'un groupe de catalogues destinés à compiler et à évaluer les séries de données marines sur l'Arctique canadien. Pour plus de commodité, la question traitée est structurée en trois grandes disciplines: physique, chimie et biologie. L'Arctique a été divisé arbitrairement en sept régions géographiques qui englobent autant que possible les grandes régions océanographiques. Les catalogues sont presentés de façon à faciliter la comparaison entre les sujets et les régions. Le domaine est si vaste qu'il est impossible de fournier tous les catalogues en une seule fois. Les catalogues de la série actuellement disponibles sont indiqués à la fin de chaque volume à l'intérieur de la couverture.

La collecte des données est un processus permanent et il est prévu de mettre à jour les catalogues par la suite. Les lecteurs sont invités a soumettre par écrit les corrections et les additions à l'éstablissement auteur. Ces corrections seront traitées en direct sur ordinateur et incorporées aux listes qui pourront être obtenus sur demande.

ACKNOWLEDGEMENTS

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We would like to acknowledge the contributions of the individuals who assisted with the production of this catalogue. Many useful discussions were held with Lois Harwood and Peter Wainwright, who provided practical suggestions and insight. Colleen Bryden helped to assemble the data reports, prepare the indices and patiently and painstakingly proofed and photocopied the manuscript. The maps in Section 10 were digitized by Peter Brouwer. Word processing was accurately and efficiently carried out by Jim Lepard. Special thanks are deserved by Ted Grainger and Jim Bunch, of the Arctic Biological Station, and Ken Denman, Ocean Ecology Division, Institute of Ocean Sciences, for their review of the manuscript.

ARCTIC DATA COMPILATION AND APPRAISAL

VOLUME 9

BEAUFORT SEA: BIOLOGICAL OCEANOGRAPHY - BACTERIA, PLANKTON, EPONTIC COMMUNITY

1914 to 1985

Volume 9 : Beaufort Sea: Biological Oceanography - Bacteria, Plankton, Epontic Community

VOLUME ABSTRACT

This inventory contains descriptions of 82 data collections of bacteria, plankton and epontic community from the southeastern Beaufort Sea and Amundsen Gulf region for the period 1914 to 1985. Sampling effort and subsequent analyses and data compilation are summarized in three primary information tables followed by supporting maps and indices. Included in the tabulations are: collecting agency, sampling times and locations; taxonomic groups; specific measurements; concurrent biological, chemical and physical parameters; details on study methods; quality rating of study methods according to a five-point scale; and details on sampling effort. Laboratory experimental data and chemical analyses are excluded from the detailed review.

Key Words: Amundsen Gulf, Beaufort Sea, data appraisal, data inventory, marine bacteria, marine epontic algae, marine epontic invertebrates, marine ichthyoplankton, marine phytoplankton, marine zooplankton

1. INTRODUCTION

The objective of this inventory was to compile descriptions of bacteria, plankton and epontic community (the biota which inhabit sea ice) data sets from the Canadian Beaufort Sea region into a single data catalogue, and to appraise the documented methodologies used to sample and analyse these data. The inventory includes all known studies which were designed and completed specifically to study these biota during and prior to 1985, as well as those in which the data were collected opportunistically or concurrently during other research programmes conducted in the southeastern Beaufort Sea and Amundsen Gulf. The data themselves or statistical summaries of these data are not included in this document because this document is intended to be a detailed index to existing data collections rather than a repository.

In total, 82 data sets were inventoried. A "data set" consists of data collected during a single expedition or cruise, usually during one field season, by a single institution or agency, using a uniform manner of sampling and analysis which conforms to a common standard of measurement (i.e., be internally consistent). For easy reference, the presentation of information within each data set is subdivided into the following categories: bacteria, phytoplankton, zooplankton, ichthyoplankton, epontic algae and epontic invertebrates.

Studies which involved biochemical analyses of these biota for toxicological purposes (e.g., to quantify levels of contaminants) were catalogued in Volume 2 (Beaufort Sea: Chemical Oceanography) of this inventory series and, therefore, are only cross-referenced in this catalogue. Other experimental studies conducted are listed in Table 1 but no further details are presented as the inventory was designed to include only direct measurements on the biota.

The location and accessibility of all original data sets for bacteria, plankton and the epontic community were determined by a literature search encompassing scientific journals, books, and government, industry and public reports, and by contacting known principal investigators (see Appendix 2). More than 80% of these data sets also appear in the other Beaufort Sea inventories, which served as the starting point for the present inventory. Wherever possible, information is presented in a format consistent with those used in the companion biological inventories for the Canadian Beaufort Sea region.

Compilation of the inventory is ongoing. When new data and previously inaccessible data become available, they will be added when the inventory is updated. The inventory will be entered onto a computerized database maintained at the Institute of Ocean Sciences, Sidney, B.C. Information about new data sets, older data sets which do not appear in this inventory, or errors in this inventory should be submitted in writing to the Scientific Information Division at the Institute of Ocean Sciences.

2. SCOPE

All known studies conducted in the Canadian Beaufort Sea for bacteria were reviewed; these included studies of bacterioplankton (those present in the water column) as well as bacteria which inhabit the sediment. Similarly, this inventory includes all studies of the epontic community, even though some of these organisms may be considered members of the benthic community in open water conditions.

The planktonic community is comprised of both phytoplankton and zooplankton, which includes the ichthyoplankton (some fish eggs and larvae). However, it is important to note that, depending on the time of day, season and/or stage of the organisms, many species which might commonly be considered benthic are found in the water column. In addition, most common methods of sampling benthos may trap planktonic organisms during the descent through the water column. To resolve this overlap relative to the marine zoobenthos inventory (in preparation) and to clearly define the composition of this inventory, sampling methodology was used as the main criterion to identify the plankton data sets.

For the plankton, therefore, the scope of this compilation is arbitrarily defined as follows:

- Those methodologies which collect or record planktonic species found within the water column are included, e.g., all types of plankton nets, water bottles, photographs and sonar records.
- All biota reported as present in samples collected by the above gear are considered, including planktonic stages of fish and benthic invertebrates.

For the bacteria and epontic community, as well as the plankton, the scope of this compilation is arbitrarily defined as follows:

- Data sets describing biota present in gut contents of marine invertebrates, fish, birds and mammals are excluded.
- With one exception, only direct measurements conducted on the organisms are included; measurements derived under experimental conditions, whether _in situ_ or in the laboratory, are excluded. Due to the nature of bacteria, enumeration and

identification which were carried out on cultures grown from the water and sediment samples collected are included.

3. STUDY AREA

The study area includes that portion of the Beaufort Sea bounded on the north by the 75°N latitude, on the west by the 141°W longitude, on the south by the coastline of the Northwest Territories and on the east by Banks and Victoria Islands (Figure 1). Arbitrary subregions designated within the study area and referred to in Table 1 of this inventory are illustrated in Figure 2. The Mackenzie River delta channels are included in the inventory when data existed for these areas in the same data set as offshore data.

4. SUMMARY OF DATA COVERAGE

4.1 Study Objectives

Of the total 82 data sets, nine (11% of the total) are classified as oceanographic cruises, during which biological samples were collected concurrently with physical and/or chemical data. Seven were carried out by university (e.g., Scripps Institution of Oceanography, University of Alaska and University of Washington) and government (Pacific Biological Station) scientists on board American research ships. The remaining two were undertaken by Canadian government researchers from the Arctic Biological Station and the Department of National Defence.

On one cruise, plankton samples were collected by the Institute of Ocean Sciences during a hydrographic and magnetic survey of Amundsen Gulf.

The general objective for 23 data sets (28%) was to obtain information on the natural history or the marine ecology of the Canadian Beaufort Sea. These include the first data set appearing in this inventory, the Canadian Arctic Expedition of 1913-1918 conducted for the Department of National Defence. Samples collected during three of the other studies

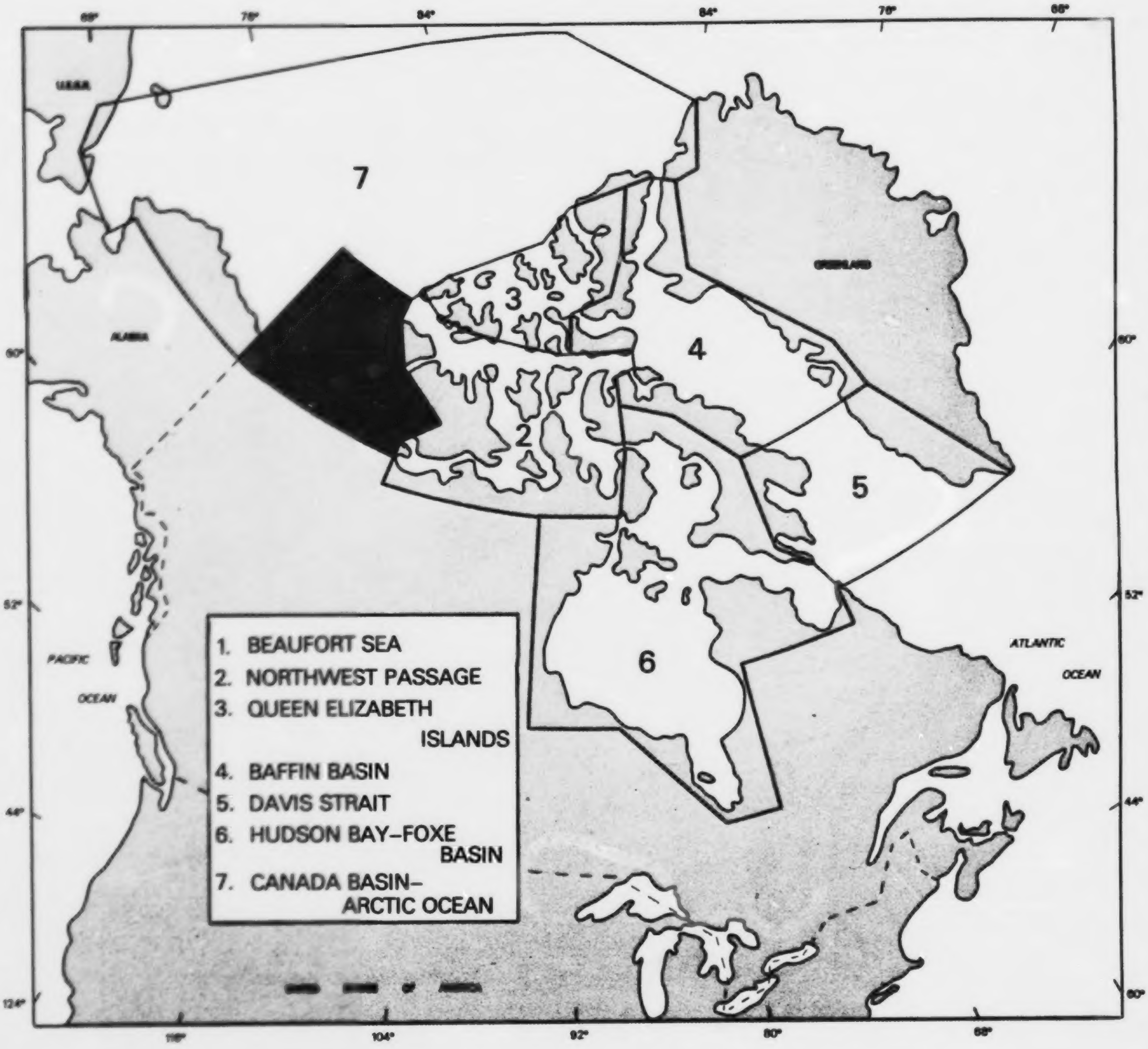

U.S.S.R.
ALASKA
GREENLAND
PACIFIC OCEAN
ATLANTIC OCEAN
7
3
4
2
5
6
1. BEAUFORT SEA
2. NORTHWEST PASSAGE
3. QUEEN ELIZABETH ISLANDS
4. BAFFIN BASIN
5. DAVIS STRAIT
6. HUDSON BAY-FOXE BASIN
7. CANADA BASIN-ARCTIC OCEAN
84°
84°
60°
52°
44°
124°
116°
104°
92°
80°
68°

FIGURE 1. Beaufort Sea study area included in this catalogue.

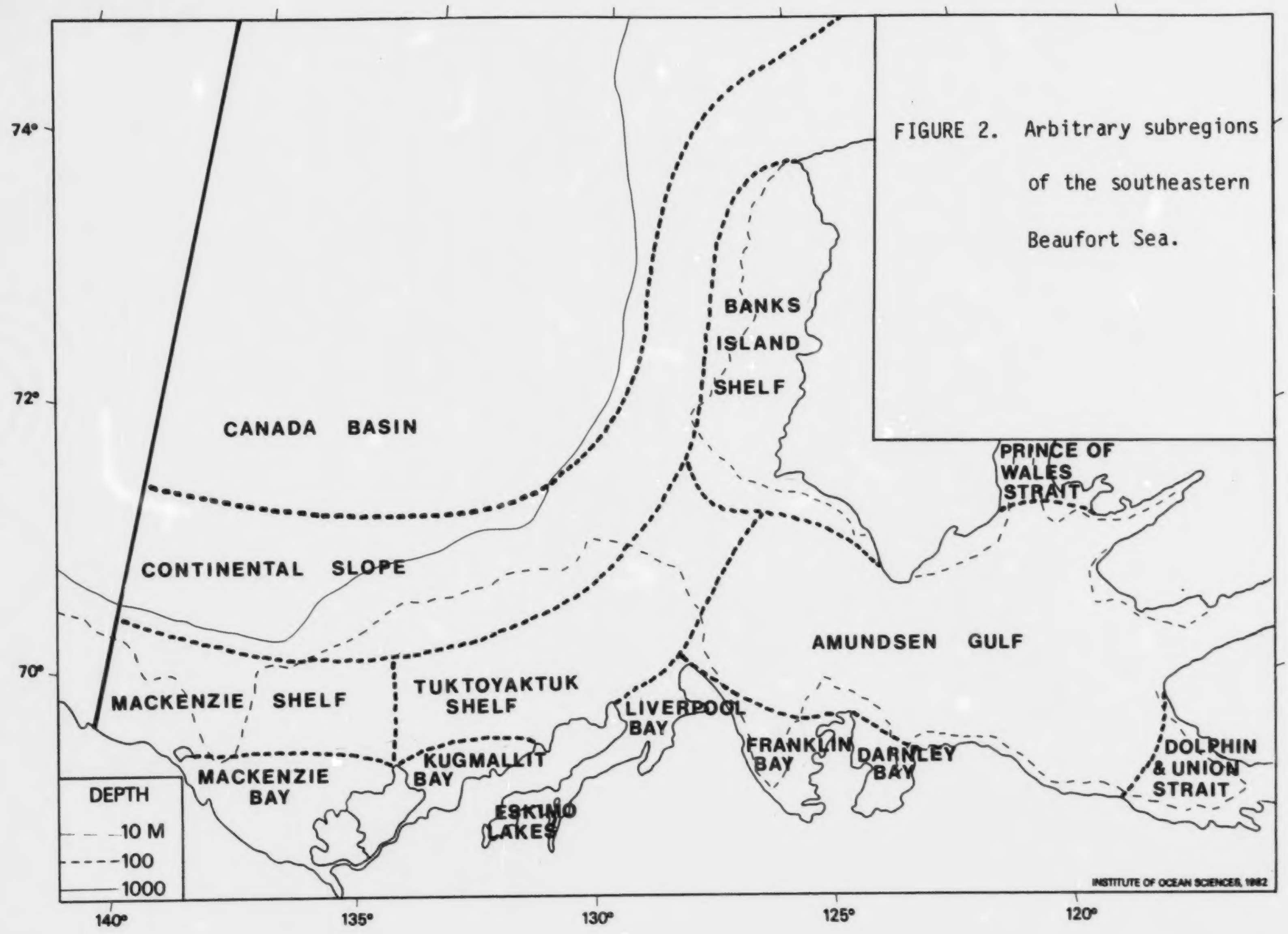

74°
72°
70°
140°
135°
130°
125°
120°
CANADA BASIN
CONTINENTAL SLOPE
MACKENZIE SHELF
MACKENZIE BAY
TUKTOYAKTUK SHELF
KUGMALLIT BAY
ESKIMO LAKES
LIVERPOOL BAY
BANKS ISLAND SHELF
AMUNDSEN GULF
FRANKLIN BAY
DARNLEY BAY
PRINCE OF WALES STRAIT
DOLPHIN & UNION STRAIT
DEPTH
10 M
100
1000
INSTITUTE OF OCEAN SCIENCES, 1982

FIGURE 2. Arbitrary subregions of the southeastern Beaufort Sea.

remain in an unanalysed state. The majority (19 data sets) were carried out by the Arctic Biological Station during 1961-1975.

In 1960, the U.S. nuclear submarine Seadragon collected plankton, in a specially-designed sampler housed in the vessel's superstructure, during a cruise to and from the North Pole.

Two cruises were conducted in 1970 and 1972 by the Atlantic Geoscience Centre in order to survey for the distribution of sediments and sediment-associated and planktonic Foraminifera.

Simon Fraser University was the collecting agency in 1983 and 1985 for two surveys to assess the bacterial community in the southwestern Canadian Beaufort Sea.

Nine fisheries surveys (11%) were carried out between 1960 and 1982 by government researchers from the Arctic Biological Station, National Museum of Natural Sciences, Freshwater Institute and the Department of Fisheries and Oceans (Western Region). Samples from six of these remain unanalysed. Of the nine surveys, three were specifically designed to obtain information on ichthyoplankton.

Plankton samples were also collected during four studies (5%) designed to obtain information on distribution, feeding and behaviour of bowhead whales.

With the advent of hydrocarbon exploration in the Beaufort Sea, numerous studies were carried out to acquire baseline information for impact assessment related to industrial activities. Three surveys (4%) were designed to determine potential effects of dredging operations and, in part, to form a basis for mitigation recommendations and future planning guidelines. The evaluation of effects of artificial island construction and operation was the objective of 15 data sets (18%) compiled in the period between 1972 and 1982. Two data sets were baseline surveys at sites proposed for construction of a petroleum-unloading pier (Mackenzie Bay) and a pipeline (Mackenzie Shelf). Five data sets (6%) provide baseline data for an impact assessment of under-ice oil spill experiments conducted between 1974 and 1980. Plankton were collected by the Ocean Chemistry Division (Institute of Ocean Sciences) during their two chemical oceanographic surveys near a drilling area in 1974 and 1975; only the samples from the 1975 survey were analysed. Monitoring surveys following accidental gas and water flows and immediately following a discharge of drilling fluid were the objectives of two data sets compiled for 1978 and 1979, respectively. In 1976, a supplementary environmental survey was carried out in an area of the Eskimo Lakes proposed for the development of natural gas reserves.

The last data set in the inventory (85-0017) was a plankton survey carried out by the Freshwater Institute under the NOGAP programme; the data from this study are not yet widely available.

4.2 Study Methods

4.2.1 Ice

Nine data sets (11%) carried out between 1972 and 1982 included an examination of the epontic community. The methods by which the samples were collected are as follows:

Method	No. of Data Sets
Sipre ice corer	5
Scrapings taken from bottom of ice	3
Ice corer (type not specified)	2
Block cut from ice by chainsaw	1
Plastic bottle (melt pond)	1
Method not specified	1

4.2.2 Water Column

Water samples were collected by seven different types of containers in 29 data sets (35%) compiled between 1973 and 1985. Samples from one of these data sets (83-0058) have not been analysed. The water samplers were indicated as follows:

Sampler	No. of Data Sets
Van Dorn bottle	14
Niskin micro-biological sampler	9
Niskin bottle	3
Plastic bottle	3
PVC bottle	2
Kemmerer bottle	1
Bucket	1

Thirteen basic types of plankton nets were used to collect the samples reported in 61 data sets (74%) between 1914 and 1985. No sample data were reported for 19 data sets; of these 19, four provided no information regarding sampling methodology used to obtain the planktonic organisms known to have been collected. The types of nets used were as follows:

Net	No. of Different Types (e.g., mesh, diameter combinations)	No. of Data Sets
Birge	1	1
Bongo	2	1
Clarke-Bumpus	1	1
Conical	26	42
Isaacs-Kidd	1	1
Faber	2	2
Miller	5	5
Nansen-type	1	2
Neuston	2	3
Scor	1	2
Stramin	1	1
Subsampler	1	1
Wisconsin	5	6

4.2.3 Sediment

Between 1974 and 1985, sediment samples were collected and reported in four data sets (5%). The gear employed in these studies were:

Gear	No. of Data Sets
Craib corer	1
Phleger corer	1
Petersen grab	2

4.3 Taxonomic Groups Examined

4.3.1 Bacteria

Sediment-associated and planktonic bacteria were reported in 15 data sets (18%) compiled between 1973 and 1985, during the time of hydrocarbon exploration in the Beaufort Sea. Although some enumeration and identification were carried out, the main aim in collecting the bacteria was to carry out experimental studies related to the effects on and of hydrocarbons.

4.3.2 Phytoplankton

Between 1914 and 1984, 30 data sets (37%) reported the presence of phytoplankton species. The groups collected most often include the orders Centrales and Pennales of class Bacillariophyceae (both in 25 data sets - 30%), class Dinophyceae (20 data sets - 24%) and class Chrysophyceae (17 data sets - 21%).

4.3.3 Zooplankton

Zooplankton were present in samples collected for 50 data sets (61%) between 1914 and 1982. In descending order of frequency of occurrence, the major groups were Calanoida (48% of 82 data sets), Cyclopoida (37%), Copepoda (those organisms identified just to subclass) (35%), Amphipoda (34%), Harpacticoida (30%), Anthomedusae (28%), Polychaeta (28%), Trachymedusae (26%), Chaetognatha (24%), Copelata (23%) and Diplostraca (21%). The remaining groups were present in less than 20% of the data sets.

4.3.4 Ichthyoplankton

Only two of the 18 data sets (22%) in which ichthyoplankton were present identified the organisms to family level or lower. The remaining data sets indicated that fish eggs and/or larvae were present. These 18 studies were conducted between 1951 and 1985.

4.3.5 Epontic Algae

Epontic algae were examined in only nine data sets (11%); these were collected between 1972 and 1982 in conjunction with petroleum exploration activities. As occurred with the phytoplankton, members of the class Bacillariophyceae were the predominant organisms, centric diatoms (Centrales) were recorded in seven data sets while pennate diatoms (Pennales) were identified from all nine studies. The next most commonly occurring organisms were the green algae (Chlorophyceae) and the dinoflagellates (Dinophyceae) which were collected during six of the research programmes.

4.3.6 Epontic Invertebrates

These organisms were recorded in the least number of data sets; only four data sets (5%) of the total 82 carried out indicated the presence of epontic invertebrates. The data were collected between 1975 and 1982 and the main taxonomic groups present were copepods, amphipods, polychaetes, medusae and protozoans.

4.4 Trends in Spatial and Temporal Coverage

The most frequently recorded study locations, based on the subregions illustrated in Figure 2, were Tuktoyaktuk Shelf (45%), Kugmallit Bay (29%), Mackenzie Bay (28%), Mackenzie Shelf (27%), Continental Slope (18%) and Eskimo Lakes (16%). The data sets are indexed by geographic location in Section 10.1.

The majority of the studies (79%) have been conducted since 1970. Only one study (13-0001) was carried out prior to 1950; since then the percentage of studies conducted, based on five-year intervals, is as follows:

Time Period	No. of Data Sets
1950-1954	6
1955-1959	2
1960-1964	7
1965-1969	1
1970-1974	22
1975-1979	25
1980-1984	15
1985	3

The year in which most studies were undertaken was 1975 (12 data sets, or 15% of total).

5. GENERAL LAYOUT OF THE CATALOGUE

The data are organized into a chronological series of data sets beginning with the year 1914. Each data set is assigned an identification number of the form yy-nnnn, where yy = last two digits of the year in which the data were collected, and nnnn = order of identification for that particular data set for that year. The data set number is a unique identifier which applies throughout this series of inventories; for example, any data set identified as 72-0001 is the same no matter where the reference to it is made. Similarly, previously assigned data set numbers which include a letter (e.g., 75-0010A, 75-0010B, etc.) are so identified in this catalogue; new data sets which logically belong to one of these "series" are identified by the next letter in the sequence. If the data set contains observations made in several different years, then the last two digits of the year that the study started are used. Since each data set does not necessarily appear in each inventory, gaps may appear in the sequence of data set numbers in this inventory for a particular year.

This inventory comprises three primary information tables followed by supporting maps and indices. Each table is preceded by an explanation of column headings.

Table 1 is a chronological list of data sets by data set number, and gives the collecting agency, sampling period, study location, taxa reported and the specific measurements made. Each measurement is assigned to one of the following eleven measurement categories:

- Number;
- Identification;
- Morphometrics;
- Age;
- Reproduction;
- Food;
- Parasites/Predators;
- Movements;
- Behaviour;
- Physiology; and
- Other.

In addition, concurrent measurements are listed by discipline (biology, chemistry and physics). Concurrent biological measurements are indicated for taxonomic groups by measurement category. Taxonomic groups included are:

- Zoobenthos;
- Phytobenthos;
- Anadromous fish;
- Marine fish;
- Birds;
- Cetaceans;

- Pinnipeds;
- Ice-associated mammals; and
- Terrestrial mammals.

Concurrent chemical measurements are given for:

- Atmosphere;
- Ice;
- Water column;
- Suspended particulates;
- Sediment; and
- Biota.

Concurrent physical measurements are given for:

- Atmosphere;
- Ice;
- Water column; and
- Substrate.

A brief note describing the purpose of the study and any experimental work conducted is also included. The Table 1 format is standardized for all biological inventories.

Table 2 contains detailed information by taxa and measurements on sampling intensity and study methods utilized; these latter include collection, storage and analysis, with comments on precision and accuracy. A quality rating code for each method is presented, accompanied by any notes which apply specifically to that rating.

Table 3 presents a detailed description of the sampling effort; information listed includes station name, location (latitude, longitude), station depth, gear types used, sampling date/hour/interval, number of samples collected and sampling depth. Where sampling co-ordinates were not specified, approximate positions were obtained by measuring plotted locations from maps given in the data report.

Station and/or transect locations are plotted by data set on a series of **maps** arranged in chronological order (Section 9). The main base map is a Lambert Conformal Conic projection with standard parallels of 71°N and 73°N, drawn to a scale of 1:4,680,000. When stations are too concentrated to assess readily, a larger-scale map is included.

Indices are presented in Section 10. The data sets are indexed by geographic area, by taxa, by measurement and by data reference/location (published, unpublished and additional information); the latter cites the information presented for each data set in the tables.

The **Appendix** includes an explanation of the codes representing gear types and a list of contacts and addresses for information sources.

5.1 User's Guide to Catalogue

Step 1: There are three indices - geographic, taxonomic and measurement - which assist in a search for specific types of information in the catalogue (Section 10). They should be consulted when information is desired on a specific geographic area (e.g., you want to know which studies were done in Kugmallit Bay), on a specific taxonomic group (e.g., you want to know which studies contain information on Amphipoda) or on a specific measurement (e.g., you want to know which studies involved a measurement of overall length), respectively. Consult the index which lists the item of interest, and record the indentification numbers of the data sets listed under the appropriate geographic area, taxonomic group or measurement.

Step 2: The data set identification numbers now listed will focus your search through the catalogue tables, maps and list of references. First, consult Table 1 entries for each data set number listed, to obtain general information on collecting and funding agencies, sampling areas and times, taxa reported, types of bacteria, plankton and epontic community measurements made and concurrent measurements. This consultation may allow you to delete some data set identification numbers from your list which no longer meet your search criteria.

Step 3: Consult Table 2 entries for each data set identification number still listed after Step 2, to obtain specific information on which measurements were made on which taxa, the number of data points, units, materials and methods, precision and accuracy, and an appraisal of the method used.

Step 4: Refer to Table 3 entries for each data set number still listed after Step 2, to obtain specific information on sampling times, locations, depths, gear types and number of collections made. If the spatial coverage is of interest, refer to the maps in Section 9 for those data set numbers listed.

Step 5: Consult the Data Reference/Location List Index (Section 10), for citation of reports or publications which report the data sets of interest, and also for information on the form, location and availability of original data.

6. APPRAISAL OF STUDY METHODS

6.1 Data Quality Criteria

The biological data for the Canadian Beaufort Sea which are described in this catalogue were appraised using a rating system based on four aspects of methodology:

- Sample collection;
- Sample storage/preservation;
- Sample analysis; and
- Precision and accuracy.

These four parameters were chosen to quantify the level of confidence in the history of the measurement, from sample collection to final analysis. They thus represent some indication of the ultimate credibility and comparability of results. General concepts and comments which outline how these are used in the data evaluation are discussed below.

It is important to note that, although the reported methodologies were rated based on objective criteria, the selection of factors encompassed by the criteria may be considered somewhat arbitrary. For example, when rating zooplankton enumeration data, we looked for details on the washing down of the net following sample collection. Although it may be considered that the correct procedure is instilled in the plankton researcher and thus does not need to be documented, we believe that biologists should report, and indeed in one or more instances have reported, this level of detail as improper procedures can bias the data. The user of this catalogue may consider this level of detail to be too rigorous and thus the criteria and data rating system are presented to enable the user to conduct an independent rating assessment.

6.1.1 Sample Collection

Numerous factors need to be considered when assessing the completeness of documentation of sample collection. Some factors apply to all the groups considered in this inventory, while others are more specific. General factors include:

a) **Sampling conditions** - High winds, rough seas and/or presence of ice can impede the proper deployment, operation and/or retrieval of the sampling gear.

b) **Station location co-ordinates and instrumentation** - Accurate positioning is essential to permit repeat and comparable sampling in the future.

c) **Date** - A precise date needs to be recorded to permit repeat and comparable sampling due to seasonal changes of life cycle and vertical distribution within the marine biological environment.

d) **Sampling depth** (and how measured) - Since the biological composition within a habitat, whether it is ice, water, or sediment, can change with depth, it is important to know from which stratum(a) the sample was collected.

e) **Sampling gear** (and how used) - Complete descriptions of the samplers and their mode of operation are necessary, not only to permit comparable repeat sampling at a future time but also to be able to assess the appropriateness of the methodology for the measurements recorded. Necessary details include at least the following:

- Ice corer: inside diameter, length, any design adaptations (e.g., to retain bottom "slush");
- Water bottles: capacity;
- Nets: mouth diameter, length to diameter ratio, mesh size, opening/closing design, tow speed, any design adaptations, flow meter;
- Sediment corer: inside diameter, length, depth of penetration, any design adaptations;
- Sediment grab: dimensions, capacity, depth of penetration, any design adaptations (e.g., weights used).

f) **Equipment calibration** - Although data or samples may be collected by a piece of equipment, the validity of the collection depends upon whether and how frequently the equipment was calibrated.

g) **Number of samples collected** - Since analysis of replicate samples provides an indication of the level of precision involved in the methodology, it is important to know whether single or replicate samples were collected.

h) **Sample handling** - The method by which the sample is retrieved from the sampler can alter not only the condition but also the composition of the sample. For example, if only a portion of the contents of a water sampler is drawn off, then the sampler should be gently agitated first to ensure an even distribution of the organisms in the water. If a net is used, it should be washed down from the outside with filtered sea water to ensure that all organisms collected end up in the sample container at the cod end and that no extraneous organisms are added.

Factors which apply to more specific measurements include:

a) **Sampling time** - Since some planktonic organisms undergo diurnal vertical migration, the hour of the day that the samples were collected should be reported, especially if the entire depth of the water column was not sampled.

b) **Sampling interval** - As nets are towed, the mesh may become clogged, especially during plankton blooms, and the efficiency of the net decreased accordingly. The length of tow (in terms of time and/or distance) should be documented.

c) **Station depth** (and how measured) - Since the depth of the water column can change significantly over a short distance, a record of the station depth acts as a check on the station location to assure the comparability of future repeat sampling. Prior knowledge of station depth assists in the proper deployment and operation of sediment samplers. Also, when interpreting plankton data, it is important to know whether all or which portion of the water column was sampled; both station and sampling depths should be reported.

d) **Ice thickness** - Since the abundance and composition of the epontic community are greatest near the under-surface of the ice, the total ice thickness relative to the length of core should be outlined in order to assess whether all or only a portion of the community was sampled.

6.1.2 Sample Storage/Preservation

After collection, the sample must be preserved in such a way as to retain its integrity and that any changes which do occur can be accounted for. Criteria examined include:

a) The potential influence of the manner of sample fixation on the ability to identify the biota (such as use of a preservative strength inadequate to prevent decomposition, freezing of soft-bodied or delicate biota, or use of a formaldehyde solution buffered inadequately to prevent decalcification of planktonic molluscs).

b) The potential influence of the manner of sample fixation on the ability to perform specific analyses (such as measurement of body length on soft-bodied organisms that were not narcotized prior to fixation or dry weight measurements on samples preserved in formaldehyde).

c) The potential influence of duration of time, between sample collection/fixation and analysis, on the data reported.

6.1.3 Sample Analysis

Although specific techniques used may differ between taxonomic groups, between measurements and from data set to data set, there are a number of general factors which need to be considered when assessing the completeness of documentation of analytical methodology. These include:

a) **Sorting** (or the procedure whereby biota are separated either from each other or from debris in the samples collected) - Criteria examined to assess the sorting procedures include the adequacy of the description of the sorting method; whether more than one sorter was involved; evidence of a quality assurance programme (e.g., calibration of equipment used) or quality control procedures (e.g., resorting of samples after the initial sort); and whether samples were split prior to sorting.

b) **Splitting** (or the procedure where the numbers of organisms are reduced, when the samples are too large to practically warrant the measurement of each individual present) - Criteria examined include whether large organisms were separated from smaller organisms prior to splitting (failure to remove larger biota may influence the randomness of the split); the adequacy of the description of the splitting device (use of too small a splitter for the sample volume may result in a non-random split); and whether an assessment of the precision and accuracy of splitting was performed. Of particular importance is the specification of the protocol used to determine which samples were split and whether documentation for the data set clearly identifies which samples were split. Splitting procedures are examined in detail, as splitting is fundamentally a procedure to estimate the composition of the sample. Thus, data sets which include samples that were not split, split once and split more than once must be examined carefully to ensure that the precision and accuracy of the splitting does not influence the internal consistency of the data set.

c) **Taxonomy** - Although identification is listed as a specific measurement in this inventory, taxonomy also applies to the other measurements as they are generally related to a specific group. The taxonomic criteria examined include whether the keys used to perform identifications were specified; the level to which the identifications were performed (identification to species level typically requires more rigorous techniques); whether more than one taxonomist was involved in the identification of a single class or order (systematic differences in identifications are possible); whether the taxonomist performing the identifications was a recognized expert or whether the identifications were verified; and whether detailed descriptions of typical specimens were reported, a reference collection maintained or archived museum specimens identified.

Typical taxonomic keys provide a means of assigning specific identities to specimens on the basis of critical morphological characteristics (diagnostic keys). Keys are normally only valid for a specific geographic region, since (i) the number of characteristics examined is frequently the minimum required to differentiate between the species known to be present in the region, and (ii) morphological characteristics may vary significantly in response to environmental conditions. Where the possibility of previously unreported species exists, and/or where the range of morphological characteristics through development and/or in response to environmental conditions is not well understood, the possibility will always exist that available keys will be proven to be inadequate. In such instances, it is necessary to either record in meticulous detail the morphological features of representative specimens, and maintain a reference collection of the same, or to perform all identifications relative to reference specimens archived elsewhere, in order to ensure the ability to re-examine the adequacy of the keys employed.

d) **Other analyses** - Criteria applicable to each specific analysis are too numerous to describe in detail. General criteria examined include the completeness of the description of the method of analysis (particularly in the case of weight or length measurements, where differences in the manner of handling of samples prior to weighing or the reference points used for length measurements may lead to systematic differences between data sets); detection limits or resolving power of instruments used; whether the specific analysis is of an objective or subjective nature, and if subjective, whether more than one analyst was involved; whether techniques used were applicable to the biota analysed; evidence of a quality assurance programme or quality control procedures; and the precision and accuracy of the analysis.

6.1.4 Precision and Accuracy

In order to compare biological phenomena between areas, seasons, years or before and after environmental perturbations, it is necessary to ensure that the data collected in different areas or at different times are comparable. The only objective way to compare data is to ensure that the precision and accuracy of the measurements are the same.

Precision is the measure of the random variation in results; it can be expressed as a standard deviation (e.g., length of carapace = 175u ± 15u). The more precise the measurements, the closer together are repeated measurements of the same parameter on the same animal or structure.

Accuracy is a measure of how close to the true value a measurement is. It is a measure of systematic variation in the results, for example a weight measurement may always be 1 mg too high if the scale is zeroed incorrectly.

In the physical and chemical sciences, multiple measurements are often easy to make to determine precision. To determine accuracy, standards are available, although relating chemical standards of one chemical composition to samples of a differing structure is often difficult. Physical and chemical laboratories develop sampling and analytical protocols and quality control procedures to ensure obtaining the best results possible. Some biological measurements are simple physical measurements on organisms (e.g., weight, total length). Simple protocols and quality control procedures can ensure the precision and accuracy of such measurements.

Although, in general, explicit information to allow an assessment of precision and/or accuracy was not available, an attempt was made to indicate and assess the relative precision which was documented.

6.2 Definition of the Rating System

The rating system is intended to be a guide to appraisal of study methodologies and not an absolute statement of data quality. Study methodologies were rated according to a five-level scale, defined as follows:

Rating Score	Definition
0	Data are found or judged to be wrong.
1	Data are suspect because of ill-defined doubts; the data are probably not internally consistent, and trends or patterns within the data are likely erroneous.
2	Insufficient information is provided to assess data; data were not or could not be investigated.

3	Data are internally consistent; patterns or trends within data are probably real but comparison with other data sets may be difficult or impossible.
4	Data are internally consistent and are sufficiently standardized or tied to a reference that comparison with other data at this rating score should be possible. Data may not be accurate in an absolute sense.

Because a 2 rating is not better than a 0 or 1 rating, the system is not truly hierarchical. Ignoring the 2 rating, however, the system is hierarchical (0, 1, 3, 4). The system is presented this way to provide continuity with other catalogues in the series.

The operational unit of the catalogue series is the "measurement." This is defined as a single determination of some variable, or if multiple determinations were made, the mean value plus or minus the standard deviation. The measurement reflects what was actually recorded, such as the length and width of a bacterium. It does not encompass derived values, such as biovolume, unless these were the only values documented. A composite list of measurements is provided in Table 6.1 (follows Section 6.4).

Each measurement type in each data set was appraised separately. The investigator's methods were judged by the criteria outlined in Section 6.1 and a rating score from 0 to 4 applied. Because data are only as credible as the weakest "link" in the methodology, the overall score is the lowest of those assigned to any one individual criterion.

It is fundamental to the rating system that the data be repeatable, not only by the original investigator, but also by anyone else wishing to repeat the measurements. For this to be possible, the original work would be performed in a consistent and defined manner, and the investigator would provide a complete and detailed description of all sampling and analytical methods. All terms and units would be defined without any vagueness or imprecison.

Completeness implies that enough data were collected to answer the question or test the hypothesis for which they were collected. For example, determination of the life stage of only ten copepods would not be complete enough to describe the age composition of an area's copepod population, even if all ten measurements were rated 4. Completeness and final use are independent of the quality of the data and have not been taken into account in the rating system. The rating achieved by particular data may, on the other hand, determine the purposes for which they can be used.

6.3 Utility of the Data Ratings

The usefulness of the data presented in this inventory will depend on the question or environmental concern that is being considered. Such uses may include identification of appropriate taxonomic keys that may be required for a study, information for the design of sampling programmes, and data for baseline purposes, impact assessment and monitoring. The data ratings assigned may be seen to separate groups of data. These are distinguished by fundamental differences in information, such as those data sets which include invalid data; those which provide insufficient information to assess the validity or potential utility of the data; and those which allow comparison between data sets.

Level 1: Identification of keys required and methodology problems.

At this least discriminating level, the data can be used to identify some of the taxonomic keys that may be required for a study and to identify methodology problems. Most data, even scoring 0, 1 or 2, may be usable for such purposes. However, data rating 0 may include data sets where the data have been lost or discarded. Such data sets may be of limited value even at this level.

Level 2: Use of the data at the individual organism level.

Certain applications of biological data are, to some extent, independent of the manner in which the sample was collected and the size of the sample, in that the sample unit is the individual organism. Such uses include estimation of the relationship between length and weight, and compilation of life-history information (such as presence of gravid females at a given time). Taxonomic identifications must be adequate to the intended level of organization (e.g., species) to allow such usage. Use of the data for such purposes may be tentatively possible even with 1-rated data. It is incumbent on the user to define acceptable data quality standards for such applications.

Level 3: Comparison of data within the data sets.

At this level, comparison of time series or relative sample properties (such as community composition, population parameters and the presence of a given species) within a given data set could be used to determine whether there are significant differences between areas or between stations. Data with a rating of 3 may tentatively be used for this purpose. It is necessary to verify that the precision of the data is sufficient to resolve differences within the range of the reported data and, also, that sample collection did not introduce large differences between stations with respect to what the samples represent.

Level 4: Comparison of data between data sets.

This is the minimum rating level required for long-term studies and for studies describing spatial patterns. Studies involving subtle changes in communities or populations in response to perturbations (such as chemical contaminants or physical impacts) or in response to physical factors (such as depth or salinity) require data with a high level of confidence and the ability to standardize the data to remove biases between stations. Only data with a rating of 4 could be considered for such applications, but will be inadequate when the samples are not representative of the community, population or area sampled. Full interpretation of the biological data is impossible without consideration of how the method of sampling, and physical and seasonal factors influence what the samples or observations represent. Thus, when information such as the presence of ice, upwelling zones and other factors is not available to support the biological data, even 4-rated data will be of limited value. In such cases, conclusions will be tentative, being based primarily on inference and conjecture.

In this catalogue, more than 2,200 measurements were rated; approximately 99% of these were assigned a 2 rating as we consider that insufficient documentation was available to assess the credibility and comparability of the research data. As discussed above, the usefulness of the 2-rated data sets to the user of this catalogue will depend on the question or environmental concern being considered.

6.4 Data Rating Flow Chart

The 109 measurements identified in this catalogue are listed in Table 6.1, which is followed by a description of the gear codes used in determining Number. To assign a rating to each measurement, the documented methods were scrutinized according to the specific criteria described in Section 6.1. These criteria were developed on the basis of experience and discussion with investigators. For ease of use, the criteria are summarized in chart form to allow users of this catalogue to review an assigned rating, or to conduct an independent appraisal.

The rating chart is organized into a sequence of questions, and the ratings depend on the answers to them. "Sampling parameters" refer to the set of experimental parameters (e.g., type of equipment used, time of day work conducted, station location, etc.) which define the scope of the collecting programme.

While some criteria are fundamental to the making of some measurements, the same criteria may be less important or irrelevant to others. For this reason, a "bypass" system has been used with the flow chart. Questions 1 and 2 (Q.1 and Q.2), which deal with sample collection, primarily apply to numerical measurements (e.g, Number; Morphometrics - biomass, weight; Reproduction - spermatophores, egg number) which could be influenced if improper methods were used. For example, rough handling could dislodge eggs and spermatophores. Question 5 (Q.5), which deals with calibration of analytical equipment, does not apply to those measurements which are purely descriptive (e.g., Morphometrics - shape, appearance, coiling, colour). Question 7 (Q.7) applies only to the above descriptive morphometrics.

All but the last question on the chart are weighted equally, since the criteria represented by each question are considered fundamental to the making of the particular measurement. A response of "NOT ENOUGH INFORMATION" to any but the last question leads to a rating of 2. The final rating assigned to a measurement is the lowest value assigned to any of the questions.

The answer to the last question on the chart, common to each measurement, determines whether a 3 or a 4 is assigned. It is worded:

> "Were the methods used and data recording sufficiently standardized and/or tied to a reference to enable comparison with another data set?"

A "YES" response to this question leads to a rating of 4, while a "NO" or "NOT ENOUGH INFORMATION" leads to a 3. This last question is the most subjective of those on the chart, relying to a certain extent on the judgement of the assessor.

In order for the user to identify the question(s) for which the documentation failed to meet the criteria (ratings 1 and 2), the ratings are presented in Table 2 (Section 8.2) accompanied by the question (Q.) number(s). For example, a rating of "2:1,5" indicates that not enough information was provided to determine whether the sampling and analytical equipment were calibrated and whether attempts were made to ensure optimum performance of the equipment.

TABLE 6.1. LIST OF MEASUREMENTS

NUMBER: (Bypass Q.7)

#/NS
#/Craib core
#/Phleger core
#/Petersen grab
#/KB
#/NB
#/NBS
#/PB
#/PVC
#/VD
#/bucket
#/IKT
#/NS CBH
#/NS N
#/NS PN
#/NS PNH
#/NS PNV
#/NS SNV
#/Subsampler
#/C-23 PNH
#/C-23 PNV
#/F-23 PNH
#/NS-12 WNV and.or 392-60 FNH
#/NS-100 STNH
#/S-30 PNV
#/12-20 PNH
#/50 NNV
#/60-25 and 240-50 PNH and PNV
#/60-25 PNV and.or 240-50 PNV
#/72-50 PNH
#/73?-NS MNH
#/73?-12 WNV
#/73-30 PNH
#/73-30 PNO
#/73-30 PNV
#/75-C PNV
#/75-NS MNH and.or 243-NS MNH
#/79-30 PNH
#/79-30 PNV
#/80-12.7 BRNH
#/80-12.7 BRNV
#/100-30 WNV
#/150-50 PNV
#/156-10 MNH
#/156-50 WNH
#/156-50 WNV
#/158-22.5 PNO
#/158-22.5 PNV
#/202-C PNV
#/202-S PNH
#/202-S PNV
tests/202-S PNV
#/202-10 MNH or 726-66 PN
#/233-30 PNV
#/233-50 PNH
#/233-50 PNV
#/282-30 PNH
#/282-30 PNV
#/392-60 FNH
#/505-60 FNH
#/569-50 PNO
#/571-50 PNH
#/571-50 PNV
#/571-100 PNH
#/750-50 WNH
#/1000-100 PNH
#/NS core
#/S core
#/scraping
#/block

IDENTIFICATION: [Bypass Q.1, 2 (except for Centrales and Pennales in 73-0002) and 7]

MORPHOMETRICS:

biomass, total dry (Bypass Q.7)
biomass, total wet (Bypass Q.7)
biovolume, (Bypass Q.1, 2 and 7)
size, overall (Bypass Q.1, 2 and 7)
weight, dry (Bypass Q.7)
weight, wet (Bypass (Q.7)
length, overall (Bypass Q.1, 2 and 7)
length, foot (Bypass Q.1, 2 and 7)
length, toes (Bypass Q.1, 2 and 7)
length, gonads (Bypass Q.1, 2 and 7)
width, overall (Bypass Q.1, 2 and 7)
width, maximum (Byass Q.1, 2 and 7)
width, minimum (Bypass Q.1, 2 and 7)
width, external pores (Bypass Q.1, 2 and 7)
width, internal pores (Bypass Q.1, 2 and 7)
width, proloculus (Bypass Q.1, 2 and 7)
height, overall (Bypass Q.1, 2 and 7)
number, external pores (Bypass Q.1, 2 and 7)
number, internal pores (Bypass Q.1, 2 and 7)
shape, overall (Bypass Q.1, 2 and 5)
shape, head (Bypass Q.1, 2 and 5)
shape, dorsal antennae (Bypass Q.1, 2 and 5)
shape, lateral antennae (Bypass Q.1, 2 and 5)
shape, foot (Bypass Q.1, 2 and 5)
shape, A1 (Bypass Q.1, 2 and 5)
shape, A2 (Bypass Q.1, 2 and 5)
shape, Mxp2 (Bypass Q.1, 2 and 5)
shape, P1 (Bypass Q.1, 2 and 5)
shape, P2 (Bypass Q.1, 2 and 5)
shape, P4 (Bypass Q.1, 2 and 5)
shape, P5 (Bypass Q.1, 2 and 5)
appearance, surface (Bypass Q.1, 2 and 5)
coiling (Bypass Q.1, 2 and 5)
colour (Bypass Q.1, 2 and 5)

AGE: (Bypass Q.1, 2 and 7)

REPRODUCTION:

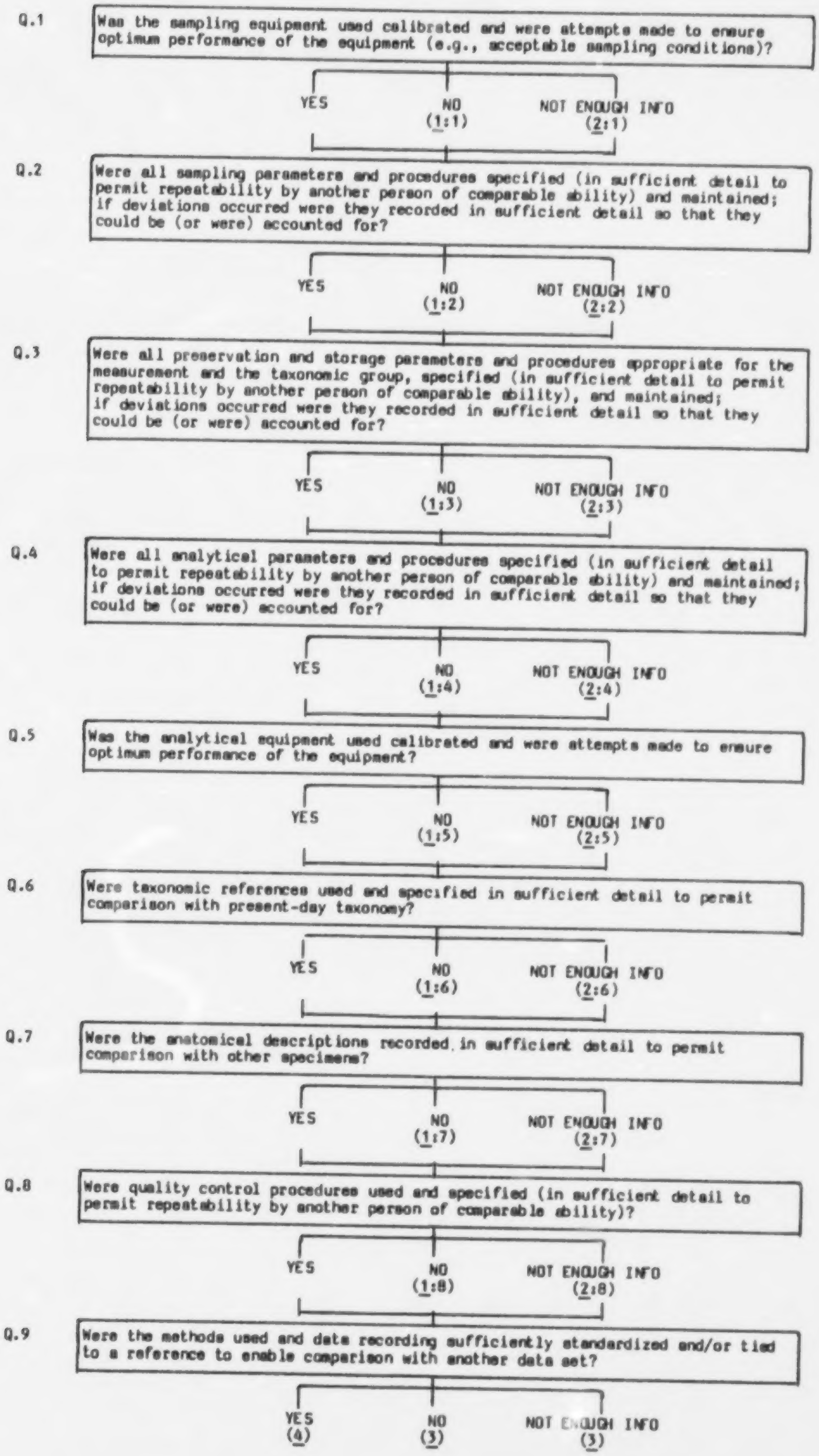
sex (Bypass Q.1, 2, 5 and 7)
spermatophores (Bypass Q.5 and 7)
egg number (Bypass Q.5 and 7)

Description of Gear Type Codes

BN - Bongo Net
BRN - BiRge closing Net
C-23 PN - Coarse-silk conical Plankton Net
CB - Clarke Bumpus net
F-23PN - Finer silk conical Plankton Net
FN - Faber Net
IKT - Isaacs-Kidd midwater Trawl
KB - Kemmerer Bottle
MN - Miller Net
N - Neuston net
NB - Niskin Bottle
NBS - Niskin Bag-Sampler
NN - Nansen-type Net
NS - Not Specified
PB - Plastic Bottle
PN - conical Plankton Net
PP - PhytoPlankton net
PVC - PVC sampler
S corer - Sipre ice corer
S-30 PN - Silk conical Plankton Net
SN - Scor Net
STN - STramin Net
Subsampler - nuclear submarine sampler
VD - Van Dorn sampler
WN - Wisconsin Net

Note: In general, numerical prefixes to the codes indicate mesh size (u) and mouth diameter (cm) of the plankton net. The suffixes H, O and V indicate Horizontal, Oblique and Vertical tows, respectively. Further details are provided in Appendix I.

FLOW CHART



Q.1 Was the sampling equipment used calibrated and were attempts made to ensure optimum performance of the equipment (e.g., acceptable sampling conditions)?

YES — NO (1:1) — NOT ENOUGH INFO (2:1)

Q.2 Were all sampling parameters and procedures specified (in sufficient detail to permit repeatability by another person of comparable ability) and maintained; if deviations occurred were they recorded in sufficient detail so that they could be (or were) accounted for?

YES — NO (1:2) — NOT ENOUGH INFO (2:2)

Q.3 Were all preservation and storage parameters and procedures appropriate for the measurement and the taxonomic group, specified (in sufficient detail to permit repeatability by another person of comparable ability), and maintained; if deviations occurred were they recorded in sufficient detail so that they could be (or were) accounted for?

YES — NO (1:3) — NOT ENOUGH INFO (2:3)

Q.4 Were all analytical parameters and procedures specified (in sufficient detail to permit repeatability by another person of comparable ability) and maintained; if deviations occurred were they recorded in sufficient detail so that they could be (or were) accounted for?

YES — NO (1:4) — NOT ENOUGH INFO (2:4)

Q.5 Was the analytical equipment used calibrated and were attempts made to ensure optimum performance of the equipment?

YES — NO (1:5) — NOT ENOUGH INFO (2:5)

Q.6 Were taxonomic references used and specified in sufficient detail to permit comparison with present-day taxonomy?

YES — NO (1:6) — NOT ENOUGH INFO (2:6)

Q.7 Were the anatomical descriptions recorded in sufficient detail to permit comparison with other specimens?

YES — NO (1:7) — NOT ENOUGH INFO (2:7)

Q.8 Were quality control procedures used and specified (in sufficient detail to permit repeatability by another person of comparable ability)?

YES — NO (1:8) — NOT ENOUGH INFO (2:8)

Q.9 Were the methods used and data recording sufficiently standardized and/or tied to a reference to enable comparison with another data set?

YES (4) — NO (3) — NOT ENOUGH INFO (3)

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8. DATA CATALOGUE TABLES

There are three different primary information tables in this inventory. Each table is preceded by explanations of column headings; however, to use the tables, the rationales for dividing plankton enumeration measurements according to gear type and for reporting the taxonomic levels must be understood.

We discovered that, when describing the methodologies, the use of "plankton net" alone to describe the collection of the biota is too all-encompassing to permit maximum use of the inventory. As discussed by Grainger (1965), due to the variety of net types used in research projects, the data can only be assessed qualitatively unless more quantitative aspects are detailed about the plankton nets. Therefore, to provide sufficient documentation to facilitate both the assessment of measurements and comparison of data sets, the quantitative measurements are described here according to collection method employed in the study. For example, #/233-50 PNH indicates that a number of specimens (#) were collected in a 233 u mesh Plankton Net with a 50 cm mouth diameter during a Horizontal tow. The gear type codes used in this inventory are summarized prior to each of the three tables and detailed in Appendix 1.

The taxa reported are listed under the following groups:

- Bacteria;
- Phytoplankton;
- Zooplankton;
- Ichthyoplankton;
- Epontic algae; and
- Epontic invertebrates.

The lowest taxonomic level chosen to record each of these groups was decided by two factors: i) the provision of maximum information and usefulness while minimizing the bulk and complexity of the inventory; and ii) the most common usage in the literature. Bacteria generally appear under the group name, due to the infrequency of specific identification. Phytoplankton and epontic algae are recorded as Class level, except for Bacillariophyceae which is divided into the Orders Centrales and Pennales. Usually, zooplankton and epontic invertebrates are recorded as Order. To facilitate comparison with the fish inventory and where the data permit, ichthyoplankton are recorded to Species using standard codes. Taxonomic classification of all these biota and codes used are presented in Section 8.2.

8.1 Table 1: Summary Listing of Data Sets

Table 1 presents a chronological summary listing of all the data sets included in this inventory. The information on each data set is described below, by table column. "INC" indicates that the information is INComplete and not available for inclusion in this catalogue.

Data Set I.D.

Each identification (I.D.) number is unique to a data set; all subsequent references to data sets are by I.D. numbers. The first two digits of the I.D. refer to the last two digits of the year during which the data were gathered; the last four digits of the I.D. are assigned to the data set.

Collecting Agency

The collecting agency was responsible for obtaining the data. If the funding agency was different from the collecting agency, the funding agency is given in parentheses. Original titles are used. Known changes in agency names include:

- Bureau of Land Management to Minerals Management Service;
- Dep. of Naval Service to Dep. of National Defence; and
- Imperial Oil Ltd. to Esso Resources Canada Ltd.

When documented, the name or type of collecting platform is also indicated.

Collecting Period

The collecting period is the time actually spent within the limits of the region, as defined in Figure 1. If known, precise dates are given. Where sampling dates were not clearly reported and were estimated from comments within the report or from concurrent studies, the dates (month and/or day) are denoted by "?".

Data sets are usually limited to information gathered in one year. The sole exception is 13-0001; this was an extended cruise which started in 1913 and surveyed the region, apparently with the same crew and the same objectives, until 1918.

Area

Subregions in which the measurements were made are listed, and shown in Figure 2.

Taxa Reported

The taxa to which the measurements pertain are indicated in this column. Where appropriate, bacteria are classified according to group, phytoplankton and epontic algae to Class (except for Bacillariophyceae which is to Order), zooplankton and epontic invertebrates to Order, and ichthyoplankton to Species. Species listings and codes are described in Section 8.2.

Biological Categories and Measurements

This column contains the specific types of measurements recorded. The information is listed according to eleven categories:

- Number;
- Identification;
- Morphometrics;
- Age;
- Reproduction;
- Food;
- Parasites/Predators;
- Movements;
- Behaviour;
- Physiology; and
- Other.

Within each category, the specific measurements or observations (parameters) are given. An explanation of the gear codes used (e.g., #/C-23 PNH) is presented following Remarks section.

Concurrent Measurements

Other measurements obtained during the study are indicated here, and are listed according to discipline: biology, chemistry and physics. Concurrent biological measurements are indicated by taxonomic group. Level of detail is to category, and the categories are the same as those used to report the plankton measurements.

Remarks

The purpose of the study and any experimental work conducted are indicated in this column. Where appropriate to the table contents, short explanatory notes may also appear.

Description of Gear Type Codes

BN - Bongo Net
BRN - BiRge closing Net
C-23 PN - Coarse-silk conical Plankton Net
CB - Clarke Bumpus net
F-23PN - Finer silk conical Plankton Net
FN - Faber Net
IKT - Isaacs-Kidd midwater Trawl
KB - Kemmerer Bottle
MN - Miller Net
N - Neuston net
NB - Niskin Bottle
NBS - Niskin Bag-Sampler
NN - Nansen-type Net
NS - Not Specified
PB - Plastic Bottle
PN - conical Plankton Net
PP - PhytoPlankton net
PVC - PVC sampler
S corer - Sipre ice corer
S-30 PN - Silk conical Plankton Net
SN - Scor Net
STN - STramin Net
Subsampler - nuclear submarine sampler
VD - Van Dorn sampler
WN - Wisconsin Net

Note: In general, numerical prefixes to the codes indicate mesh size (u) and mouth diameter (cm) of the plankton net. The suffixes H, O and V indicate Horizontal, Oblique and Vertical tows, respectively. Further details are provided in Appendix I.

TABLE 1: SUMMARY LISTING OF DATA SETS

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
13-0001	Canadian Arctic Expedition of 1913-1918 (for Dept. of Naval Service)	1914: Aug. 02-23; 1915: Sep. 14; 1916: July 17	Mackenzie Shelf, Darnley Bay, Dolphin and Union Strait	PHYTOPLANKTON Centrales Pennales Dinophyceae	Number #/C-23 PNH #/F-23 PNH #/S-30 PNV Identification Morphometrics length, overall	Zoobenthos Number Identification Fish - Anadromous Number Identification Food Birds Number Identification Food Behaviour Cetaceans Number Identification Movements Behaviour Pinnipeds Number Identification Food Behaviour Ice-associated Mammals Number Identification Morphometrics Age Reproduction Food Movements Behaviour Terrestrial Mammals Number Identification Morphometrics Age Reproduction Food Movements Behaviour		Atmosphere Wind speed Wind direction Precipitation Atm. conditions Ice Thickness Coverage Type Water Column Temperature Salinity Current Speed Current direction Depth Transparency Water level Wave climate	Natural history cruise
				ZOOPLANKTON Foraminiferida Trachymedusae Ploima Calanoida Harpacticoida Cyclopoida Copelata	Number #/C-23 PNH #/C-23 PNV #/F-23 PNH #/S-30 PNV Identification Morphometrics length, overall length, foot length, toes width, overall height, overall shape, overall shape, head shape, dorsal antennae shape, lateral antennae shape, foot shape, A1 shape, A2 shape, Mxp2 shape, P1 shape, P2 shape, P4 shape, P5 colour Age Reproduction sex spermatophores				
50-0001	Scripps Institution of Oceanography; USS Burton Island	Aug. 19-24	Canada Basin, Continental Slope, Banks Island Shelf	ZOOPLANKTON Anthomedusae Trachymedusae Narcomedusae Thecosomata Pelecypoda Annelida	Number #/50 NNV Identification Morphometrics length, overall length, gonads width, overall		Water Column Dissolved oxygen Sediment	Atmosphere Atm. conditions Cloud cover Ice Coverage Break-up Water Column	Oceanographic cruise Samples are located in Scripps Institution of Oceanography marine invertebrate collections

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
50-0001 (cont'd)				Ostracoda Copepoda Calanoida Cyclopoida Cirripedia Amphipoda Ophiuroidea Copelata	height, overall shape, overall Age Reproduction sex			Temperature Salinity Current speed Current direction Depth Turbidity Transparency Secchi depth	
51-0001	Scripps Institution of Oceanography; USS Burton Island	Sep. 14-21	Canada Basin, Continental Slope, Banks Island Shelf	ZOOPLANKTON Trachymedusae Narcomedusae Thecosomata Pelecypoda Annelida Ostracoda Copepoda Calanoida Cyclopoida Cirripedia Amphipoda Euphausiacea Echinoidea Ophiuroidea Copelata	Number #/50 NNV Identification Morphometrics length, gonads width, overall height, overall shape, overall Age	Zoobenthos Number Fish	Water Column Dissolved oxygen Sediment	Atmosphere Atm. conditions Water Column Temperature Salinity Current speed Current direction Depth Turbidity Transparency Secchi depth	Oceanographic cruise Samples are located in Scripps Institution of Oceanography marine invertebrate collections
51-0002	Defence Research Board and Arctic Biological Station; CCMV Cancolim II	Aug. 21-Sep. 15	Mackenzie Shelf, Tuktoyaktuk Shelf, Banks Island Shelf, Amundsen Gulf	ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae Narcomedusae Siphonophora Cydippida Beroida Thecosomata Gymnosomata Polychaeta Diplostraca Myodocopa Copepoda Calanoida Harpacticoida Cyclopoida Thoracica Isopoda Amphipoda Decapoda Insecta	Number #/NS CBH #/NS PNH #/NS PNV #/79-30 PNH #/79-30 PNV #/202-30 PNH #/202-30 PNV Identification Age Reproduction sex	Fish Number Identification Birds Identification Morphometrics Cetaceans Number Identification Pinnipeds Number Identification Ice-Associated Mammals Number Identification		Atmosphere Wind speed Wind direction Atm. conditions Ice Thickness Coverage Water Column Temperature Salinity Depth	Oceanographic cruise

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
51-0002 (cont'd)				Chaetognatha Copelata ICHTHYOPLANKTON Other	 Number #/NS PNH #/202-30 PNV Age				
52-0002	University of Washington; USS Burton Island	Sep. 02-10	Continental Slope, Tuktoyaktuk Shelf, Banks Island Shelf, Amundsen Gulf	INC	INC		Water Column Nutrients Dissolved oxygen Sediment	Atmosphere Wind speed Atm. conditions Ice Coverage Movement Water Column Temperature Salinity Current speed Current direction Depth Water level Sound velocity	Oceanographic cruise Inquiries at University of Washington failed to reveal location of data and samples
53-0001	USS Burton Island	Aug. 08-25	Amundsen Gulf, Prince of Wales Strait	INC	INC		Water Column Nutrients Dissolved oxygen Sediment	Atmosphere Wind speed Wind direction Water Column Temperature Salinity Current speed Current direction Depth Water level	Oceanographic cruise Samples are located in Scripps Institution of Oceanography marine invertebrate collections
54-0003	Pacific Biological Station; USCGC Northwind	Aug. 09 - Sep. 10	Tuktoyaktuk Shelf, Banks Island Shelf, Amundsen Gulf, Prince of Wales Strait	INC	INC	Zoobenthos Number Identification Phytobenthos Number Identification Fish - Anadromous Number Identification Fish - Marine Number Identification Birds Number	Water Column Nutrients Dissolved oxygen	Atmosphere Wind speed Wind direction Precipitation Atm. conditions Temperature Ice Thickness Coverage Type Water Column Temperature Salinity Current speed	Oceanographic cruise Samples unanalyzed; inquiries with Mr. R.H. Herlinveaux and Pacific Biological Station failed to reveal location of field data and samples

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
54-0003 (cont'd)						Identification Cetaceans Number Identification Pinnipeds Number Identification		Current direction Depth Sound velocity	
55-0009	Arctic Biological Station	Sep. 01-13	Eskimo Lakes	INC	INC	Zoobenthos Number Identification Morphometrics Fish Number Identification		Water Column Temperature Salinity Depth Substrate Particle size	Marine ecology survey Inquiries at Arctic Biological Station failed to reveal location of data and samples
58-0001	Scripps Institution of Oceanography; USS Burton Island	Sep. 02-04	Mackenzie Shelf, Tuktoyaktuk Shelf	INC	INC			Atmosphere Wind speed Wind direction Atm. conditions Water Column Temperature Salinity Depth Current speed Current direction Wave climate Substrate Particle size	Oceanographic survey Inquiries at Scripps Institution of Oceanography failed to reveal location of data and samples
60-0003	Arctic Biological Station; CGS Salvelinus	Aug. 06-25	Mackenzie Shelf, Mackenzie Bay	ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae Thecosomata Calanoida Mysidacea Amphipoda Euphausiacea Decapoda Chaetognatha	Number #/NS PN #/NS-100 STNH #/571-50 PNH Identification Morphometrics width, overall height, overall Age Reproduction sex	Zoobenthos Number Identification Morphometrics Reproduction Food Movements Fish Number Identification Morphometrics Age Reproduction Food Parasites	Water Column Dissolved oxygen	Water Column Temperature Salinity	Fisheries survey

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
6U-0067	Submarine Seadragon	Aug. - early Sep.	Canada Basin, Continental Slope	ZOOPLANKTON Copepoda Calanoida	Number #/Subsampler Identification Age Reproduction sex				Nuclear submarine cruise to and from North Pole Sampling location specifics designated as classified information
61-0001	Arctic Biological Station; CGS Salvelinus	July 10 - Aug. 23	Tuktoyaktuk Shelf, Liverpool Bay, Eskimo Lakes	ZOOPLANKTON Trachymedusae Siphonophora Thecosomata Polychaeta Acarina Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Thoracica Decapoda Chaetognatha Copelata	Number #/73-30 PNH #/73-30 PNV Identification Age Reproduction sex	Zoobenthos Number Identification Morphometrics Reproduction Food Movements Fish Number Identification Morphometrics Age Reproduction Food Parasites Movements	Water Column Dissolved oxygen	Water Column Temperature Salinity Depth	Marine ecology survey
61-0079	Arctic Biological Station	July 06 - Aug. 10	Mackenzie Bay	INC	INC Age	Fish			Fisheries survey Inquiries at Arctic Biological Station failed to reveal location of data and samples
62-0001	Arctic Biological Station; CGS Salvelinus	July 21 - Aug. 22	Tuktoyaktuk Shelf, Liverpool Bay, Eskimo Lakes, Franklin Bay	ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae Siphonophora Polychaeta Acarina Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia Thoracica	Number #/73-30 PNV #/233-50 PNH #/233-50 PNV Identification Morphometrics size, overall width, overall height, overall Age Reproduction sex egg number	Zoobenthos Number Identification Fish Number Identification Morphometrics Age Reproduction Food Movements	Water Column Dissolved oxygen	Water Column Temperature Salinity Depth	Marine ecology survey

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
62-0001 (cont'd)				Amphipoda Decapoda Insecta Chaetognatha Copelata ICHTHYOPLANKTON Other	Number #/233-50 PNH Age				
63-0001	Arctic Biological Station; CGS *Salvelinus*	July 05 - Aug. 24	Amundsen Gulf, Franklin Bay	INC	INC	Zoobenthos Number Identification Morphometrics Fish Identification Morphometrics Reproduction	Water Column Dissolved oxygen	Water Column Temperature Salinity	Fisheries survey Inquiries at Arctic Biological Station failed to reveal location of data and samples
64-0001	Arctic Biological Station; CGS *Salvelinus*	July 05	Franklin Bay	INC	INC			Water Column Temperature Salinity	Oceanographic cruise Inquiries at Arctic Biological Station failed to reveal location of data and samples
69-0066	Institute of Marine Science, University of Alaska; USCGC *Staten Island*	Aug.	Canada Basin, Continental Slope, Mackenzie Shelf	PHYTOPLANKTON Centrales Pennales Chrysophyceae Dinophyceae ZOOPLANKTON Foraminiferida Radiolaria Tintinnida Rotifera Polychaeta Copepoda	Number #/NS PNH? #/NS PNV Identification Number #/NS PNH? #/NS PNV Identification Age		Water Column Metals Nutrients DOC POC Suspended Particulates Sediment	Water Column Depth Substrate Particle size	Oceanographic cruise - part of continuing Arctic Ocean programme Only preliminary analyses available
70-0002	Atlantic Geoscience Centre; CSS *Hudson*	Aug. 27 - Sep. 24	Canada Basin, Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	ZOOPLANKTON Foraminiferida	Number #/202-S PNH #/202-S PNV # tests/202-S PNV Identification Morphometrics biomass, total dry	Zoobenthos Number Identification Morphometrics	Sediment Metals	Ice Coverage Type Water Column Temperature Salinity Depth	Survey for distribution of sediments and foraminiferida

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
70-0002 (cont'd)					width, maximum width, minimum width, external pores width, internal pores width, proloculus number, external pores number, internal pores appearance, surface coiling Age			Substrate Particle size	
70-0067	National Museum of Natural Sciences	July 10-29	Kugmallit Bay	ZOOPLANKTON Anthomedusae Rotifera Polychaeta Diplostraca Calanoida Harpacticoida Cyclopoida Mysidacea Isopoda Amphipoda Chaetognatha	Number #/202-10 MNH or 726-66 PN Identification Age				Larval fish survey Oxycephalus samples analysed; other ichthyoplankton samples lost
				ICHTHYOPLANKTON	INC				
71-0001	Arctic Biological Station; CGS Salvelinus	July 19	Tuktoyaktuk Shelf, Kugmallit Bay	ZOOPLANKTON Anthomedusae Trachymedusae Narcomedusae Gastropoda Polychaeta Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia Mysidacea Cumacea Amphipoda Chaetognatha Copelata	Number #/73-30 PNV #/233-30 PNV Identification Age Reproduction sex	Zoobenthos Number Identification Morphometrics	Water Column Nutrients Chlorophyll Dissolved oxygen DOC POC	Atmosphere Atm. conditions Ice Coverage Type Movement Water Column Temperature Salinity Depth Secchi depth	Marine ecology survey
				ICHTHYOPLANKTON Other	Number #/233-30 PNV Age				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
71-0004	Arctic Biological Station	Aug. 26 - Dec. 15	Eskimo Lakes	ZOOPLANKTON Anthomedusae Polychaeta Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia	Number #/73-30 PNV #/233-30 PNV Identification Age Reproduction sex	Zoobenthos Number Identification Morphometrics	Water Column Nutrients Chlorophyll Dissolved oxygen pH POC	Ice Thickness Snow cover Water Column Temperature Salinity Depth Secchi depth	Marine ecology survey
72-0001	Atlantic Geoscience Centre	Mar. 21 - Apr. 09	Canada Basin, Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay	ZOOPLANKTON Foraminiferida	Number #/75-C PNV #/202-C PNV #/202-S PNV Identification Morphometrics biomass, total dry width, maximum coiling Age			Ice Coverage Type Water Column Temperature Salinity Depth Substrate	Survey for distribution of sediments and foraminiferida
72-0004	Arctic Biological Station	Mar. 17 - Nov. 25	Eskimo Lakes	ZOOPLANKTON Anthomedusae Rotifera Nematoda Mollusca Gastropoda Pelecypoda Polychaeta Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia Thoracica Cumacea Amphipoda Other	Number #/73-30 PNO #/73-30 PNV Identification Morphometrics weight, wet Age Reproduction sex		Ice Nutrients Chlorophyll POC Water Column Nutrients Chlorophyll Dissolved oxygen pH POC	Ice Salinity Thickness Snow cover Water Column Temperature Salinity	Marine ecology survey
				ICHTHYOPLANKTON Other	Number #/73-30 PNV Age				
				EPONTIC ALGAE Centrales Pennales Chlorophyceae	Number #/S core Identification				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
72-0004 (cont'd)				Euglenophyceae Dinophyceae					
72-0007	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.); MV Arcticus	Aug. 15-17?	Mackenzie Bay	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Dinophyceae	Number #/12-20 PNH Identification	Zoobenthos Number Identification Fish Number Identification Morphometrics Reproduction Food Birds Number Identification Movements Behaviour Cetaceans Number Identification Reproduction Food Movements Pinnipeds Number	Water Column Dissolved oxygen Major elements Carbon dioxide pH Alkalinity Suspended Particulates	Atmosphere Wind speed Wind direction Precipitation Atm. conditions Ice Thickness Coverage Freeze-up Break-up Water Column Temperature Salinity Conductivity Current speed Current direction Depth Water level Wave climate Secchi depth Substrate Particle size	Study to assess the impact of artificial island construction Report states mid-August collecting period; specific dates estimated from concurrent studies
				ZOOPLANKTON Ploima Diplostraca Copepoda	Number #/12-20 PNH Identification Age				
73-0001	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.)	Mar. 20?-21?	Mackenzie Bay	PHYTOPLANKTON Pennales Chlorophyceae	Number #/73?-12 WNV Identification	Zoobenthos Number Identification	Water Column Metals Nutrients Dissolved oxygen Major elements pH Alkalinity	Ice Thickness Water Column Temperature Salinity Conductivity Current speed Current direction Turbidity	Winter baseline data programme Report states March collecting period; specific dates estimated from concurrent studies
				ZOOPLANKTON Rotifera Copepoda Other	Number #/73?-12 WNV Identification Age				
73-0002	Arctic Biological Station; MV North Star of Herschel Island	July 20-27	Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	BACTERIA	Number #/NBS Identification	Zoobenthos Number Identification Morphometrics Fish Number Identification Morphometrics Age Reproduction Food	Water Column Nutrients Chlorophyll Dissolved oxygen POC Sediment Metals	Atmosphere Atm. conditions Water Column Temperature Salinity Depth Secchi depth Substrate Particle size	Marine ecology survey
				PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Cyanophyceae Euglenophyceae Dinophyceae	Number #/VD Identification				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
73-0002 (cont'd)				Other					
				ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae Narcomedusae Siphonophora Gastropoda Thecosomata Polychaeta Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Thoracica Mysidacea Cumacea Amphipoda Euphausiacea Decapoda Insecta Chaetognatha Chordata Copelata	Number #/73-30 PNV #/571-50 PNH #/571-50 PNV Identification Age Reproduction sex				
				ICHTHYOPLANKTON Other	Number #/73-30 PNV #/571-50 PNH Age				
73-0003A	Arctic Biological Station	Feb. 23 - Oct. 01	Eskimo Lakes	BACTERIA	Number #/NBS Identification	Zoobenthos Number Identification Morphometrics Fish Identification Food	Ice Nutrients Chlorophyll POC Water Column Nutrients Chlorophyll Dissolved oxygen pH POC Sediment Metals	Ice Salinity Thickness Snow cover Water Column Temperature Salinity Secchi depth Substrate Particle size	Marine ecology survey Experimental studies included: Bacteria - isolation and characterization of oil-degrading bacteria, determination of capability of mixed and pure cultures to degrade petroleum at different temperatures and determination of limiting factors
				ZOOPLANKTON Cnidaria Anthomedusae Rotifera Nematoda Gastropoda Pelecypoda Polychaeta Diplostraca Copepoda Calanoida Harpacticoida	Number #/73-30 PNV Identification Age Reproduction sex				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
73-0003A (cont'd)				Cyclopoida Thoracica Mysidacea Cumacea Insecta Other					
				ICHTHYOPLANKTON Other	Number #/73-30 PNV Age				
				EPONTIC ALGAE Centrales Pennales Chlorophyceae Euglenophyceae Dinophyceae	Number #/S core Identification				
73-0003B	Arctic Biological Station	Aug. 05-17?	Liverpool Bay, Eskimo Lakes	ZOOPLANKTON Tintinnida Anthomedusae Leptomedusae Trachymedusae Rotifera Polychaeta Crustacea Diplostraca Calanoida Harpacticoida Cyclopoida Cirripedia Other	Number #/73-30 PNV Identification Age		Water Column Nutrients Chlorophyll Dissolved oxygen	Water Column Temperature Salinity Depth Secchi depth	Marine ecology survey Report states two weeks in August as collection period; specific dates estimated from concurrent studies Biological data apply to upper 10 m of water column only
74-0001	F.F. Slaney & Co. Ltd., Vancouver (for Imperial Oil Ltd.)	Mar. 11?-Apr. 18?	Mackenzie Bay	PHYTOPLANKTON Centrales Pennales Chlorophyceae	Number #/73?-12 WNV Identification	Zoobenthos Number Identification Morphometrics	Water Column Metals Nutrients Dissolved oxygen Major elements pH Hardness TC Suspended Particulates Sediment TC TKN	Ice Thickness Snow cover Water Column Temperature Salinity Current speed Current direction Depth Turbidity Substrate Particle size	Winter baseline data programme Report states March-April collecting period; specific dates estimated from concurrent studies
				ZOOPLANKTON Rotifera Copepoda Other	Number #/73?-12 WNV Identification Age				
				EPONTIC ALGAE Pennales	Number #/scraping Identification				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
74-0001 (cont'd)				EPONTIC INVERTEBRATES Crustacea	Number #/scraping Identification Age				
74-0003	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd. & Sun Oil Co. Ltd.); MV *Arcticus*	Aug. 03 - Sep. 10	Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	PHYTOPLANKTON Centrales Cyanophyceae	Number #/PVC #/72-50 PNH Identification	Zoobenthos Number Identification Fish - Anadromous Number Identification Morphometrics Age Reproduction Food Fish - Marine Number Identification Morphometrics Age Reproduction Food	Water Column Metals Nutrients Chlorophyll Dissolved oxygen pH Alkalinity Hardness Suspended Particulates	Atmosphere Wind speed Wind direction Atm. conditions Ice Thickness Coverage Type Break-up Movement Water Column Temperature Salinity Conductivity Current speed Current direction Turbidity Water level	Oceanographic survey to assess the environmental effects of artificial island construction
				ZOOPLANKTON Trematoda Rotifera Pelecypoda Diplostraca Ostracoda Calanoida Harpacticoida Cyclopoida Mysidacea Amphipoda Diptera Ephemeroptera Other	Number #/72-50 PNH Identification Age				
				ICHTHYOPLANKTON Other	Number #/72-50 PNH Identification Age				
74-0007A	Arctic Biological Station; MV *Theta*	Aug. 27 - Sep. 03	Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	BACTERIA	Number #/NBS Identification	Zoobenthos Number Identification Morphometrics	Water Column Nutrients Chlorophyll Dissolved oxygen pH POC	Water Column Temperature Salinity Depth Secchi depth	Marine ecology survey Experimental studies included: Bacteria - effects of shipping conditions, effects of incubation temperature and presence or absence of oxygen, and effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification, carbon metabolism and respiration; Phytoplankton - ef-
				PHYTOPLANKTON Centrales Pennales Chrysophyceae Euglenophyceae Dinophyceae Other	Number #/VD Identification				
				ZOOPLANKTON Cnidaria Anthomedusae Trachymedusae Narcomedusae Siphonophora	Number #/73-30 PNV #/569-20 PNO Identification Age Reproduction				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
74-0007A (cont'd)				Anthozoa Beroida Nematoda Gastropoda Thecosomata Polychaeta Myodocopa Copepoda Calanoida Harpacticoida Cyclopoida Monstrilloida Cirripedia Thoracica Mysidacea Isopoda Amphipoda Euphausiacea Decapoda Chaetognatha Copelata Other	sex				fects of crude oils on growth
74-0007C	Arctic Biological Station; Helicopter	July 14 - Aug. 16	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf	BACTERIA	Number #/NBS Identification		Water Column Nutrients Chlorophyll	Water Column Temperature Salinity	Marine ecology survey Experimental studies included: Bacteria - effects of shipping conditions, effects of incubation temperature and presence or absence of oxygen, and effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification, carbon metabolism and respiration; Phytoplankton - effects of crude oils on growth; Zooplankton - effects of crude oils on behaviour.
				PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Cyanophyceae Euglenophyceae Dinophyceae Other	Number #/VD Identification				
				ZOOPLANKTON Foraminiferida Radiolaria Cnidaria Trachymedusae Narcomedusae Siphonophora Nematoda Gastropoda Thecosomata Polychaeta	Number #/73-30 PNV #/233-30 PNV Identification Age Reproduction sex				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
74-0007C (cont'd)				Myodocopa Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia Thoracica Cumacea Isopoda Amphipoda Euphausiacea Decapoda Chaetognatha Copelata Other					
74-0008	Institute of Ocean Sciences - Ocean Chemistry Division; MV Theta	Aug. 18-24	Mackenzie Shelf, Mackenzie Bay	INC	INC		Water Column Hydrocarbons Nutrients Dissolved oxygen Alkalinity Suspended Particulates Hydrocarbons Sediment Hydrocarbons Biota Hydrocarbons	Water Column Temperature Salinity Conductivity Depth	Chemical oceanographic survey near drilling area Samples preserved frozen and unanalysed; have been discarded
74-0010A	Arctic Biological Station	Feb. 27 - May 26	Eskimo Lakes	BACTERIA	(experimental studies only)		Ice Nutrients Chlorophyll POC Water Column Nutrients Chlorophyll Dissolved oxygen pH PIC	Ice Salinity Thickness Snow cover Water Column Temperature Salinity	Marine ecology survey Experimental studies included: Bacteria - effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification and carbon metabolism
				ZOOPLANKTON Anthomedusae Rotifera Nematoda Gastropoda Polychaeta Copepoda Calanoida Harpacticoida Cyclopoida Thoracica	Number #/73-30 PNV Identification Age Reproduction mm				
				EPONTIC ALGAE Centrales Pennales	Number #/S core Identification				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
74-0010A (cont'd)				Chlorophyceae Euglenophyceae Dinophyceae					
74-0010C	Arctic Biological Station	July 13 - Aug. 21	Eskimo Lakes	BACTERIA PHYTOPLANKTON	Number #/Phleger core Identification (experimental studies only)		Water Column Nutrients Chlorophyll Dissolved oxygen pH POC	Water Column Temperature Salinity Secchi depth	Marine ecology survey Experimental studies included: Bacteria - effects of shipping conditions, effects of incubation temperature and presence or absence of oxygen and effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification and carbon metabolism; Phytoplankton - in situ gross primary production
74-0011	Aquatic Environments Ltd. (for Canadian Arctic Gas Study Ltd. & Alaskan Arctic Gas Study Co.)	July 08 - Sep. 09	Mackenzie Shelf	ZOOPLANKTON Cnidaria Ctenophora Nematoda Polychaeta Oligochaeta Ostracoda Copepoda Mysidacea Isopoda Amphipoda Diptera Chaetognatha	Number #/505-60 FNM Identification	Zoobenthos Number Identification Fish - Anadromous Number Identification Morphometrics Age Reproduction Food Fish - Marine Number Identification Morphometrics Age Reproduction Food	Water Column Metals Nutrients Dissolved oxygen Major elements pH Alkalinity	Water Column Temperature Salinity Conductivity Depth	Baseline survey of nearshore and estuarine habitat prior to pipeline construction
74-0019	F.F. Slaney & Co. Ltd. (for Sunoco E & P Ltd.)	Mar. 05-07	Mackenzie Bay	PHYTOPLANKTON Pennales Chlorophyceae ZOOPLANKTON Rotifera	Number #/100-30 WNV Identification Number #/100-30 WNV	Zoobenthos Number Identification	Water Column Metals Dissolved oxygen Carbon dioxide pH Alkalinity	Ice Thickness Water Column Temperature Salinity Conductivity	Preliminary assessment of possible environmental effect of winter artificial island construction

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
74-0019 (cont'd)					Identification		Hardness Suspended Particulates	Current speed Current direction Depth Turbidity	
74-0027	NORCOR Engineering and Research Ltd.	Aug. 13	Franklin Bay	PHYTOPLANKTON Centrales Pennales Chrysophyceae Dinophyceae	Number #/VD Identification		Water Column Hydrocarbons pH CO_2	Water Column Temperature Salinity	Baseline survey prior to under-ice oil spill experiment
				ZOOPLANKTON Cnidaria Anthomedusae Stauromedusae Cydippida Rotifera Copepoda Calanoida Cyclopoida	Number #/60-25 and 240-50 PNH and PNV Identification				Experimental studies included: Phytoplankton - in situ primary production, effects of oil on growth and photosynthesis
74-0139	Arctic Biological Station	Apr. 01 - Sep. 06	Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	INC	INC	Fish			Fisheries survey Inquiries at Arctic Biological Station failed to reveal location of data and samples
75-0004	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.)	July 10 - Aug. 18	Mackenzie Bay, Kugmallit Bay	PHYTOPLANKTON Centrales Pennales Dinophyceae Other	Number #/PVC Identification	Zoobenthos Number Identification Fish - Anadromous Number Identification Morphometrics Age Reproduction Food Fish - Marine Number Identification Morphometrics Age Reproduction Food Cetaceans Number	Water Column Metals Nutrients Chlorophyll Dissolved oxygen Major elements pH Alkalinity Hardness TIC TOC Suspended Particulates	Atmosphere Wind speed Wind direction Atm. conditions Ice Thickness Type Freeze-up Break-up Movement Water Column Temperature Salinity Conductivity Current speed Current direction Depth Turbidity Substrate	Monitoring effects of summer construction of offshore artificial islands
				ZOOPLANKTON Rotifera Pelecypoda Copepoda Calanoida Harpacticoida Cyclopoida Mysidacea Amphipoda	Number #/72-50 PNH Identification				
				ICHTHYOPLANKTON Other	Number #/72-50 PNH				

TABLE 9 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
75-0004 (cont'd)						Identification			
75-0006	Institute of Ocean Sciences - Ocean Chemistry Division; MV Pandora II	Aug. 05-23	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	PHYTOPLANKTON Centrales Pennales ZOOPLANKTON Thecosomata Calanoida Amphipoda	Number #/737 MMH Identification Number #/737 MMH Identification		Water Column Hydrocarbons Nutrients Dissolved oxygen Alkalinity Suspended Particulates Hydrocarbons Sediment Hydrocarbons Biota Hydrocarbons	Water Column Temperature Salinity Conductivity Depth	Chemical oceanographic survey near drilling area
75-0009	Arctic Biological Station; MV Theta	Aug. 04-29	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay	ZOOPLANKTON Thecosomata Gymnosomata Myodocopa Calanoida Mysidacea Amphipoda	Number #/IKT Identification Morphometrics size, overall Age Reproduction sex	Zoobenthos Number	Suspended Particulates	Water Column Temperature Salinity	Marine ecology survey
75-0010A	Arctic Biological Station	Feb. 27	Eskimo Lakes	ZOOPLANKTON Polychaeta Copepoda Calanoida Harpacticoida Cyclopoida Other	Number #/73-30 PNV Identification Age Reproduction sex		Water Column Nutrients Chlorophyll Dissolved oxygen POC	Water Column Temperature Salinity Secchi depth	Marine ecology survey
75-0010B	Arctic Biological Station	June 15 - Aug. 16	Eskimo Lakes	BACTERIA PHYTOPLANKTON Centrales Pennales Chrysophyceae Euglenophyceae Dinophyceae Other	Number #/Petersen grab Identification Number #/VD Identification		Water Column Nutrients Chlorophyll Dissolved oxygen pH POC	Water Column Temperature Salinity Secchi depth	Marine ecology survey Experimental studies included: Bacteria-effects of shipping conditions, effects of incubation temperature and presence or absence of oxygen and effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification and

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
75-0010B (cont'd)									carbon metabolism; Phytoplankton - effects of oils, Corexit and oil - Corexit mixtures on primary production and growth, and toxicity of oil, Corexit and oil-Corexit mixtures.
75-0012A	Arctic Biological Station; Helicopter	June 17 - July 25	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf	BACTERIA	Number #/NBS #/Petersen grab Identification	Zoobenthos Number Identification Morphometrics	Water Column Nutrients Chlorophyll Dissolved oxygen	Water Column Temperature Salinity Sediment Particle size	Marine ecology survey Experimental studies included: Bacteria- effects of shipping conditions, effects of incubation temperature and presence or absence of oxygen and effects of oil and hydrocarbon fractions on nitrogen fixation, denitrification and carbon metabolism; Phytoplankton - effects of oils, Corexit and oil - Corexit mixtures on primary production and growth, and toxicity of oil, Corexit and oil-Corexit mixtures.
				PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Euglenophyceae Dinophyceae Other	Number #/VD Identification				
				ZOOPLANKTON Porifera Trachymedusae Narcomedusae Siphonophora Anthozoa Gastropoda Thecosomata Polychaeta Myodocopa Copepoda Calanoida Harpacticoida Cyclopoida Thoracica Cumacea Isopoda Amphipoda Euphausiacea Chaetognatha Ophiuroidea Copelata	Number #/73-30 PNV Identification Age Reproduction sex				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
75-0012A (cont'd)				Other					
75-0012B	Arctic Biological Station; CGS *Salvelinus*	July 11-17	Kugmallit Bay	ZOOPLANKTON Anthomedusae Rotifera Nematoda Pelecypoda Polychaeta Tardigrada Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Mysidacea Amphipoda Insecta Copelata Other	Number #/73-30 PNH #/73-30 PNV #/1000-100 PNH Identification Age Reproduction sex	Zoobenthos Number Identification Morphometrics	Water Column Nutrients Chlorophyll Dissolved oxygen	Water Column Temperature Salinity	Marine ecology survey
				ICHTHYOPLANKTON SC	Number #/1000-100 PNH Identification Age				
75-0012D	Arctic Biological Station; Snowmobile	May 09	Kugmallit Bay	PHYTOPLANKTON Centrales Pennales Euglenophyceae Other	Number #/VD Identification		Water Column Nutrients Chlorophyll Dissolved oxygen	Water Column Temperature Salinity	Marine ecology survey
75-0012E	Arctic Biological Station; MV *Pandora II*	Aug. 04-09	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay	BACTERIA	Number #/NBS Identification		Water Column Nutrients Chlorophyll Dissolved oxygen	Water Column Temperature Salinity	Marine ecology survey Experimental studies included: *Bacteria* - effects of petroleum on respiration; *Phytoplankton* - effects of oils, Corexit and oil-Corexit mixtures on primary production and growth, and toxicity of oil, Corexit and oil-Corexit mixtures
				PHYTOPLANKTON Centrales Pennales Chrysophyceae Euglenophyceae Dinophyceae Other	Number #/VD Identification				
				ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae	Number #/1000-100 PNH Identification Age				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
75-0012E (cont'd)				Thecosomata Gymnosomata Amphipoda Decapoda					
				ICHTHYOPLANKTON Other	Number #/1000-100 PNH Identification Morphometrics length, overall				
75-0026	NORCOR Engineering and Research Ltd.	Feb. 07 - July 20	Franklin Bay	PHYTOPLANKTON Pennales Chrysophyceae Dinophyceae Cryptophyceae	Number #/VD Identification	Zoobenthos Number Identification Phytobenthos Number Identification Birds Number Identification Behaviour Ice-Associated Mammals Number Identification Terrestrial Mammals Number Identification	Ice Hydrocarbons Water Column Hydrocarbons Chlorophyll pH CO_2 Biota Hydrocarbons Chlorophyll	Atmosphere Wind speed Wind direction Atm. conditions Solar radiation Ice Temperature Salinity Thickness Snow cover Water Column Temperature Salinity	Under-ice oil spill experiment Experimental studies included: Phytoplankton - primary production
				ZOOPLANKTON Tintinnida Cnidaria Copepoda Echinoidea	Number #/60-25 PNV and.or 240-50 PNV Identification				
75-0028	Inland Waters Directorate, Glaciology Division	May 13 - June 25	Franklin Bay	PHYTOPLANKTON Other	Number #/PB (comb. with zooplankton) #/VD (comb. with zooplankton)		Ice Chlorophyll Water Column Chlorophyll Dissolved oxygen Major elements pH Alkalinity	Atmosphere Atm. conditions Solar radiation Ice Salinity Conductivity Snow cover Under-ice irradiance Absorption coefficient Water Column Temperature Salinity Conductivity Underwater irradiance Attenuation coefficient	Impact assessment of experimental under-ice oil spill Plankton numbers represent the entire community, i.e., phytoplankton and zooplankton combined
				ZOOPLANKTON Other	Number #/PB (comb. with phytoplankton #/VD (comb. with phytoplankton				
				EPONTIC ALGAE Centrales Pennales Chlorophyceae Phaeophyceae Cyanophyceae Dinophyceae	Number #/NS #/PB Identification				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
75-0028 (cont'd)				EPONTIC INVERTEBRATES Mastigophora Ciliophora Heterotrichida Crustacea	Number #/PB Identification Age				
75-0051	Arctic Biological Station	July 07 - Aug. 17	Kugmallit Bay	INC	INC	Fish			Fisheries survey Inquiries at Arctic Biological Station failed to reveal location of data and samples
76-0003	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.)	Aug. 14 - Sep. 07	Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	PHYTOPLANKTON Centrales Pennales Chrysophyceae Dinophyceae Other	Number #/PB Identification	Zoobenthos Number Identification Morphometrics Fish - Anadromous Number Identification Morphometrics Fish - Marine Number Identification Morphometrics	Water Column Metals Nutrients Dissolved oxygen pH Alkalinity Hardness TC TIC TOC Suspended Particulates Sediment Metals	Atmosphere Atm. conditions Water Column Temperature Conductivity Depth Turbidity Substrate Particle size	Bio-physical baseline survey of proposed and existing offshore artificial island sites
				ZOOPLANKTON Foraminiferida Tintinnida Cnidaria Scyphozoa Rotifera Gastropoda Pelecypoda Polychaeta Diplostraca? Calanoida Harpacticoida Cyclopoida Cirripedia Mysidacea Amphipoda Euphausiacea Chaetognatha Copelata	Number #/PB #/156-50 WNH #/156-50 WNV Identification Morphometrics weight, dry				
76-0004	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.)	July 17?-Sep. 01?	Kugmallit Bay	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Dinophyceae	Number #/PB Identification	Zoobenthos Number Identification Morphometrics Fish - Anadromous Number	Water Column Metals Nutrients Chlorophyll Dissolved oxygen pH	Atmosphere Atm. conditions Water Column Temperature Salinity Conductivity	Environmental overview to determine potential effects of dredging operations on the marine environment

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
76-0004 (cont'd)				ZOOPLANKTON Tintinnida Calanoida Cyclopoida	Number #/PB #/156-10 MNH Identification	Identification Morphometrics Fish - Marine Number Identification Morphometrics	Alkalinity Hardness Suspended Particulates Sediment	Current speed Current direction Turbidity Transparency Wave climate Substrate Particle size	Experimental studies included: Phytoplankton - primary production; Zooplankton - grazing rate
76-0005	F.F. Slaney & Co. Ltd. (for Imperial Oil Ltd.); MV Arcticus	July 31 - Sep. 06	Kugmallit Bay	PHYTOPLANKTON Centrales Pennales Chrysophyceae Dinophyceae Other ZOOPLANKTON Foraminiferida Tintinnida Calanoida	Number #/PB Identification Number #/PB #/156-10 MNH Identification Morphometrics length, overall	Zoobenthos Number Identification Morphometrics Fish - Anadromous Number Identification Morphometrics Reproduction Age Food Fish - Marine Number Identification Morphometrics Reproduction Age Food	Water Column Metals Nutrients Chlorophyll Dissolved oxygen Major elements pH Alkalinity Hardness TC TIC TOC CO_2 Suspended Particulates Sediment Metals TC TOC	Atmosphere Wind speed Wind direction Atm. conditions Water Column Temperature Salinity Conductivity Current speed Current direction Depth Turbidity Transparency Wave climate Substrate Particle size	Survey to determine effect of sacrificial beach island construction on local plankton community Experimental studies included: Phytoplankton - primary production and respiration; Zooplankton - grazing rate
76-0036	F.F. Slaney & Co. Ltd. (for Gulf Oil Canada Ltd.)	June 24 - Sep. 21	Eskimo Lakes	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Cyanophyceae Dinophyceae Cryptophyceae ZOOPLANKTON Rotifera Ploima Crustacea Diplostraca Ostracoda Copepoda Calanoida Harpacticoida Cyclopoida Diptera	Number #/VD Identification Age Number #/80-12.7 BRNH #/80-12.7 BRNV Identification Age	Zoobenthos Number Identification Fish - Anadromous Number Identification Morphometrics Age Reproduction Food Fish - Marine Number Identification Morphometrics Age Reproduction Food	Water Column Nutrients Chlorophyll Dissolved oxygen pH Alkalinity Hardness TOC Suspended Particulates	Ice Thickness Water Column Temperature Salinity Conductivity Depth Turbidity Secchi depth Colour	Supplementary environmental survey of area proposed for development of natural gas reserves

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS BIOLOGICAL	CONCURRENT MEASUREMENTS CHEMICAL	CONCURRENT MEASUREMENTS PHYSICAL	REMARKS
76-0036 (cont'd)				Other					
77-0002	Aquatic Environments Ltd. (for Imperial Oil Ltd.)	July 16?-Sep. 02?	Tuktoyaktuk Shelf	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Euglenophyceae Cryptophyceae	Number #/KB Identification	Zoobenthos Number Identification Fish - Anadromous Number Identification Morphometrics Age Reproduction Food Fish - Marine Number Identification Morphometrics Age Reproduction Food	Water Column Dissolved oxygen pH Suspended Particulates	Water Column Temperature Salinity Conductivity Turbidity	Baseline fisheries survey to assess effects of borrow operations and to form basis for mitigation recommendations and future planning guidelines
				ZOOPLANKTON Anthomedusae Trachymedusae Beroida Calanoida	Number #/NS-12 WWV and.or 392-60 FNH #/392-60 FNH Identification				Biological data derived from text of report; report tables not available for review
77-0003	Institute of Ocean Sciences - Ocean Chemistry Division; MV Pandora II	Aug. 09 - Sep. 10	Amundsen Gulf	ZOOPLANKTON Cnidaria Trachymedusae Narcomedusae Siphonophora Thecosomata Gymnosomata Calanoida Cyclopoida Amphipoda Euphausiacea Decapoda Chaetognatha Asteroidea Copelata	Number #/NS N #/75-NS MNH and.or 243-NS MNH Identification Morphometrics size, overall Age		Water Column Hydrocarbons Metals Nutrients Dissolved oxygen Sediment Metals Biota Hydrocarbons Metals	Water Column Temperature Salinity	Hydrographic and magnetic survey
				ICHTHYOPLANKTON Other	Number #/NS N #/75-NS MNH and.or 243-NS MNH Age				
77-0009	Envirocon Ltd. (for Imperial Oil Ltd.); MV Imperial Immerk	July - Aug.	Tuktoyaktuk Shelf	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae	Number #/VD Identification	Zoobenthos Number Identification Morphometrics Fish - Anadromous	Water Column Nutrients Chlorophyll Dissolved oxygen Alkalinity	Atmosphere Wind speed Wind direction Atm. conditions Water Column	Baseline and monitoring survey during artificial island construction

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
77-0009 (cont'd)				Cyanophyceae Dinophyceae ZOOPLANKTON Protozoa Mastigophora Rhizopodea Amoebida Radiolaria Ciliata Tintinnida Anthomedusae Leptomedusae Limnomedusae Trachymedusae Narcomedusae Ctenophora Sipuncula Thecosomata Gymnosomata Pelecypoda Polychaeta Diplostraca Copepoda Calanoida Harpacticoida Cyclopoida Cirripedia Amphipoda Euphausiacea Decapoda Insecta Bryozoa Chaetognatha Asteroidea Ophiuroidea Copelata ICHTHYOPLANKTON Other	 Number #/VO #/150-50 PNV #/571-100 PNH Identification Morphometrics biomass, total wet Number #/150-50 PNV #/571-100 PNH Identification Age	Number Identification Age Food Fish - Marine Number Identification Age Food	Suspended Partic- ulates Sediment TC TOC	Temperature Salinity Conductivity Current speed Current direction Depth Secchi depth Wave climate Irradiance Substrate Particle size	
77-0020	Department of Fisheries and Oceans, Pacific Region	July 27 - Aug. 01	Mackenzie Bay	INC	INC	Zoobenthos Number Identification Fish - Anadromous		Water Column Temperature Salinity	Baseline ecological survey at site proposed for construction of petroleum-

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
77-0020 (cont'd)						Number Identification Morphometrics Age Food Fish - Marine Number Identification Morphometrics Age Food			unloading pier Inquiries at Fisheries and Oceans (Pacific Region) and Freshwater Institute failed to reveal location of data and samples
78-0002	Seakem Oceanography Ltd. (for Canadian Marine Drilling Ltd.); MV Canmar Supplier V	July 17-18	Tuktoyaktuk Shelf	ZOOPLANKTON Hydrozoa Leptomedusae Gastropoda Trachymedusae Thecosomata Gymnosomata Calanoida Mysidacea Amphipoda Decapoda Chaetognatha Copelata	Number #/NS SNV Identification Morphometrics biomass, total dry Age	Zoobenthos Number Identification Morphometrics	Water Column Metals Dissolved oxygen Major elements pH Sediment Metals Biota Metals	Water Column Temperature Salinity Conductivity Depth	Monitoring survey following gas and water flows
				ICHTHYOPLANKTON Other	Number #/NS SNV Morphometrics biomass, total dry Age				
78-0018	Seakem Oceanography Ltd. (for Canadian Marine Drilling Ltd.)	July 20	Mackenzie Shelf	ZOOPLANKTON Hydrozoa Anthomedusae Calanoida Amphipoda Chaetognatha	Number #/NS SNV Identification	Zoobenthos Number Identification Morphometrics	Water Column Metals Major elements pH Sediment Metals Biota Metals	Water Column Temperature Salinity Conductivity Depth	Baseline survey
				ICHTHYOPLANKTON Other	Number #/NS SNV Identification Age				
79-0009	Arctic Laboratories Ltd. (for Canadian Marine Drilling Ltd.); MV Canmar Supplier V	Sep. 09	Continental Slope	PHYTOPLANKTON Bacillariophyceae Centrales	Number #/NB Identification		Water Column Metals Nutrients Suspended Partic-	Water Column Temperature Conductivity Depth	Monitoring effects of freshly discharged drilling fluid on phytoplank-

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
79-0009 (cont'd)				Dinophyceae			ulates	Transparency	ton community Experimental studies included: Phyto-plankton - effect of discharged drilling fluid on primary production
79-0026	Dome Petroleum Ltd.	Dec. 13	Tuktoyaktuk Shelf	EPONTIC ALGAE Bacillariophyceae Pennales Cyanophyceae	Number #/NS core Identification	Zoobenthos Number Identification Morphometrics Birds Number Identification Mammals Number Identification Behaviour	Water Column Hydrocarbons Dissolved oxygen Sediment Hydrocarbons	Atmosphere Wind speed Wind direction Atm. conditions Solar radiation Ice Temperature Salinity Thickness Structure Snow cover Water Column Temperature Salinity Current speed Current direction Depth Water level	Under-ice oil spill experiment
79-0037	Freshwater Institute	May - Sep.	Tuktoyaktuk Shelf	INC	INC	Zoobenthos Phytobenthos Fish - Anadromous Number Identification Age Food Fish - Marine Number Identification Age Food	Water Column Chlorophyll Dissolved oxygen	Water Column Temperature Salinity Water level	Fisheries survey Sampling specifics and analytical data not available at time this inventory prepared
80-0016	Arctic Laboratories Ltd. (for Dome Petroleum Ltd.)	Apr. 08 - June 19	Tuktoyaktuk Shelf	EPONTIC ALGAE Bacillariophyceae Centrales Pennales Other	Number #/block #/NS core Identification	Birds Number Identification Pinnipeds Number Identification Behaviour	Water Column Hydrocarbons Dissolved oxygen Sediment Hydrocarbons	Atmosphere Wind speed Wind direction Atm. conditions Solar radiation Ice Temperature	Under-ice oil spill experiment

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
80-0016 (cont'd)				EPONTIC INVERTEBRATES Nematoda Copepoda	Number #/NS core Identification			Salinity Thickness Structure Snow cover Water Column Temperature Salinity Current speed Current direction Depth Water level	
80-0025	LGL Ecological Research Associates Inc. (for U.S. Bureau of Land Management)	Aug. 14 - Sep. 06	Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay	ZOOPLANKTON Anthomedusae Leptomedusae Trachymedusae Narcomedusae Siphonophora Ctenophora Nematoda Thecosomata Gymnosomata Pelecypoda Polychaeta Phyllodocida Oligochaeta Crustacea Myodocopa Calanoida Harpacticoida Cyclopoida Cirripedia Mysidacea Cumacea Isopoda Amphipoda Euphausiacea Decapoda Chaetognatha Ophiuroidea Copelata Other	Identification Morphometrics biomass, total wet Age	Zoobenthos Identification Morphometrics Fish Identification Morphometrics Birds Number Identification Behaviour Cetaceans Number Identification Age Food Movements Behaviour Pinnipeds Number Identification Behaviour		Atmosphere Wind speed Wind direction Precipitation Atm. conditions Ice Coverage Type Water Column Temperature Salinity Depth Wave climate	Bowhead whale behaviour study

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
81-0003	Arctic Laboratories Ltd. (for Esso Resources Canada Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd.); Helicopter	Mar. 07 - Oct. 10	Tuktoyaktuk Shelf	BACTERIA	Number #/NBS	Zoobenthos Number Identification	Ice Nutrients Chlorophyll Phaeopigments Water Column Hydrocarbons Metals Nutrients Chlorophyll Dissolved oxygen Alkalinity DOC POC TOC Suspended Particulates Metals Sediment Hydrocarbons Metals	Atmosphere Solar radiation Ice Salinity Thickness Water Column Temperature Salinity Depth Substrate Particle size	Time-series survey to evaluate effects of artificial island construction and operation
81-0014	Environmental Protection Service, N.W.T.	July 04	Tuktoyaktuk Shelf	INC	INC		Sediment Hydrocarbons		Sample unanalysed; resides in EPS, N.W.T., warehouse
81-0015	Arctic Laboratories Ltd. (for Fisheries and Oceans Canada)	Mar. 07 - Oct. 10	Tuktoyaktuk Shelf	PHYTOPLANKTON Centrales Pennales Chlorophyceae Chrysophyceae Euglenophyceae Dinophyceae Craspedophyceae Cryptophyceae Other	Number #/NB Identification		Ice Nutrients Chlorophyll Phaeopigments Water Column Nutrients Chlorophyll Dissolved oxygen Phaeopigments	Ice Salinity Thickness Snow cover Water Column Temperature Salinity Depth Underwater irradiance	Survey of seasonal changes in the epontic and planktonic communities near an artificial island
				ZOOPLANKTON Hydrozoa Anthomedusae Narcomedusae Gastropoda Polychaeta Copepoda Calanoida Harpacticoida Cyclopoida Mysidacea Amphipoda	Number #/158-22.5 PNO #/158-22.5 PNV Identification Age				

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
81-0015 (cont'd)				Chaetognatha Ophiuroidea					
				ICHTHYOPLANKTON Stichaeidae Other	Number #/158-22.5 PND #/158-22.5 PNV Identification Age				
				EPONTIC ALGAE Centrales Pennales Chlorophyceae Dinophyceae Other	Number #/S core Identification				
				EPONTIC INVERTEBRATES Nematoda Polychaeta	Number #/S core #/scraping Identification				
				Copepoda Calanoida Harpacticoida Amphipoda	Age				
81-0027	LGL Ecological Research Associates Ltd. (for U.S. Bureau of Land Management); MV *Sequel*	Aug. 01 - Sep. 06	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Kugmallit Bay	ZOOPLANKTON Hydrozoa Anthomedusae Leptomedusae Trachymedusae Narcomedusae Siphonophora Ctenophora Beroida Gastropoda Thecosomata Gymnosomata Diplostraca Ostracoda Calanoida Cirripedia Mysidacea Isopoda Amphipoda Euphausiacea Decapoda Chaetognatha Asteroidea	Identification Morphometrics biomass, total wet Age	Zoobenthos Identification Morphometrics Age Fish Identification Morphometrics Age Cetaceans Number Identification Morphometrics Age Food Movements Behaviour Pinnipeds Number Identification Behaviour		Atmosphere Wind speed Wind direction Precipitation Atm. conditions Ice Coverage Type Water Column Temperature Salinity Conductivity Depth Turbidity Wave climate	Bowhead whale behaviour study

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
81-0027 (cont'd)				Copelata					
81-0031	Freshwater Institute	June 28 - Sep. 05	Kugmallit Bay	INC	INC	Fish - Anadromous Number Identification Age Food Fish - Marine Number Identification Age Food	Water Column Metals Nutrients Chlorophyll Dissolved oxygen pH DOC POC Suspended Particulates	Water Column Temperature Salinity Conductivity	Fisheries survey Sampling specifics and analytical data not available at time this inventory prepared
81-0101	Arctic Biological Station (for Esso Resources Canada Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd.)	July 26	Tuktoyaktuk Shelf	BACTERIA	Number #/MBS		Water Column Nutrients Chlorophyll DOC POC TOC	Water Column Temperature Salinity	Survey to determine microheterotrophic activity around an artificial island
82-0012	Department of Fisheries and Oceans, Western Region	July 11 - Aug. 16	Kugmallit Bay	ICHTHYOPLANKTON PCHR RNSM ARCD SFCD LUMP FHSC SWFL	Number #/750-50 MMH Identification Morphometrics length, overall Age			Water Column Temperature Salinity	Fisheries survey to determine species composition of the ichthyoplankton, abundance and distribution of larvae and their growth
82-0109	Arctic Biological Station (for Esso Resources Canada Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd.)	Apr. 15-16	Tuktoyaktuk Shelf	BACTERIA	Number #/MBS		Water Column Nutrients Chlorophyll DOC POC TOC	Water Column Temperature Salinity	Survey to determine microheterotrophic activity around an artificial island
82-0110	Arctic Laboratories Ltd. (for Fisheries and Oceans Canada)	Feb. 19 - June 11	Tuktoyaktuk Shelf	PHYTOPLANKTON Centrales Pennales Chlorophyceae Euglenophyceae Dinophyceae Craspedophyceae Cryptophyceae Other	Number #/ML Identification		Ice Nutrients Chlorophyll Phaeopigments Water Column Nutrients Chlorophyll Dissolved oxygen Phaeopigments	Ice Salinity Thickness Snow cover Water Column Temperature Salinity Depth Underwater irradiance	Survey of seasonal changes in the epontic and planktonic communities near an artificial island

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
82-0110 (cont'd)				ZOOPLANKTON Hydrozoa Anthomedusae Trachymedusae Narcomedusae Gastropoda Polychaeta Copepoda Calanoida Harpacticoida Cyclopoida Isopoda Amphipoda Chaetognatha Ophiuroidea Copelata	Number #/158-22.5 PNV Identification Age				
				ICHTHYOPLANKTON Osteichthyes	Number #/158-22.5 PNV Identification Age				
				EPONTIC ALGAE Centrales Pennales Chlorophyceae Chrysophyceae Euglenophyceae Dinophyceae Craspedophyceae Other	Number #/S core Identification				
				EPONTIC INVERTEBRATES Tintinnida Hydrozoa Turbellaria Nematoda Polychaeta Copepoda Calanoida Harpacticoida Amphipoda Other	Number #/S core #/scraping Identification Age				
83-0054	Environmental Protection Service, N.W.T.; MV Sequel	July 12-15	Tuktoyaktuk Shelf, Kugmallit Bay	INC	INC		Sediment Metals Hydrocarbons	Water Column Temperature Conductivity	Samples unanalysed; reside in EPS, N.W.T., warehouse

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
83-0054 (cont'd)								Transparency Substrate Particle size	
83-0058	G.A. Borstad Ltd. (for Fisheries and Oceans, Indian and Northern Affairs, Dome Petroleum Ltd., and Gulf Canada Resources Inc.)	Aug. 21 - Oct. 09	Tuktoyaktuk Shelf, Kugmallit Bay, Liverpool Bay, Eskimo Lakes	INC	INC		Water Column Chlorophyll Suspended Particulates	Water Column Temperature Salinity Secchi depth Colour	Survey using ground-truthed remote sensing to compare bowhead whale distribution with water column parameters Samples unanalysed; reside at Arctic Laboratories Ltd., Sidney, B.C.
83-0059	Simon Fraser University	Aug. 23-31	Continental Slope, Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	BACTERIA	Number #/VD Morphometrics length, overall width, overall		Water Column Chlorophyll Suspended Particulates	Water Column Temperature Salinity	Survey to assess bacterial community Experimental studies included: bacterioplankton production and glucose heterotrophic activities
84-0033	University of Toronto (for Environmental Protection Service); Helicopter	Aug. 01-04	Kugmallit Bay	BACTERIA PHYTOPLANKTON Centrales	Number #/bucket Identification Morphometrics biovolume		Water Column Metals Chlorophyll		Survey to evaluate effect of dredged sediment disposal on primary producers and bacteria Experimental studies included: Bacteria - glucose uptake, phosphate uptake and effects of cadmium; Phytoplankton - primary production, phosphate uptake and effects of cadmium
85-0007	LGL Ltd. (for Dept. of Indian and Northern Affairs)	Aug. 18-31	Mackenzie Bay	INC	INC	Fish Number Identification Age Cetaceans Number Identification	Water Column Nutrients Chlorophyll Dissolved oxygen	Atmosphere Wind speed Wind direction Precipitation Atm. conditions Ice Coverage	Bowhead whale food study Samples collected and in storage; analysis incomplete at time this inven-

TABLE 1 (cont'd)

DATA SET I.D.	COLLECTING AGENCY	COLLECTING PERIOD	AREA	TAXA REPORTED	BIOLOGICAL CATEGORIES AND MEASUREMENTS	CONCURRENT MEASUREMENTS			REMARKS
						BIOLOGICAL	CHEMICAL	PHYSICAL	
85-0007 (cont'd)						Age Movements Behaviour Pinnipeds Number Identification Behaviour		Type Water Column Temperature Salinity Conductivity Depth Turbidity Wave climate	tory prepared Sampling specifics also not yet available
85-0016	Simon Fraser University	Aug. 26-27	Kugmallit Bay	BACTERIA	Number #/VD #/Craib core Morphometrics length, overall width, overall		Water Column Chlorophyll Suspended Particulates	Water Column Temperature Salinity	Survey to assess bacterial community Experimental studies included: bacterioplankton production and glucose heterotrophic activities
85-0017	Freshwater Institute	July 18 - Sep. 12	Mackenzie Shelf, Tuktoyaktuk Shelf, Mackenzie Bay, Kugmallit Bay	ZOOPLANKTON ICHTHYOPLANKTON	INC INC		Water Column Chlorophyll	Water Column Temperature Salinity Depth Secchi depth	NOGAP B2.1 programme Samples collected and in storage; analysis incomplete at time this inventory prepared

8.2 Table 2: Measurements, Methodology and Quality Rating

Table 2 presents information on each of the specific measurements in each data set. Data sets are listed chronologically according to I.D. number; the earliest data sets are listed first. Information on the measurements and specific abbreviations used are given below, by column. To reduce the overall length of Table 2, rather than present detailed methodologies and comments on precision/accuracy in the columns, they are identified as METHOD # and a footnote is presented.

Data Set I.D.

Each identification (I.D.) number is unique to a data set; all subsequent references to data sets are by I.D. numbers. The first two digits of the I.D. refer to the last two digits of the year during which the data were gathered; the last four digits of the I.D. are assigned to the data set.

Taxa

The taxa which were measured are indicated in this column. Where appropriate, bacteria are classified according to group, phytoplankton and epontic algae to Class (except for Bacillariophyceae which is to Order), zooplankton and epontic invertebrates to Order and ichthyoplankton to Species. In order to maximize the usefulness of the inventory, when the biota were identified to taxonomic levels lower than those listed above, they appear as codes under the taxonomic group. Species listings and codes are explained at the end of the section following Remarks. "un" means unidentified.

Measurements

Specific measurements within each data set are listed by taxa. For each taxon, the measurements obtained are listed by category in the same order as in Table 1. Abbreviations which represent gear type codes are explained at the end of this section following the Species List.

Units

This column indicates the unit(s) used as a standard when obtaining each measurement. All the units used for each measurement are indicated; semi-colons separate different units. Slashes separate different parts of the same unit. Metric units are abbreviated. "Relative" indicates the use of imprecise units; the units used are listed under precision/accuracy. "NA" designates that the unit was Not Available when the documentation was examined. "u" means micron. "s" means sample; "ss" means subsample.

No. Stns.

The number of stations at which the measurement was taken is listed in this column. Where more than one unit was used to report a measurement, the number of stations are recorded for each unit and are separated by a semi-colon. For example, if "#/s; #/ss" appears under Units and "1; 3" appears under No. Stns., then the enumeration of the taxonomic group in a sample (s) was conducted for one station and in a subsample (ss) for three stations. "NA" designates that the number of stations was Not Available in the documentation examined.

No. Samples

This column includes the number of data points (often no. of individual organisms) obtained on each measurement for each taxon in each data set. "Relative" indicates that the information is expressed in imprecise terms; the terms used are listed under precision/accuracy. Not Specified ("NS") indicates that the number of data points was not reported. "NA" designates that the number of samples was Not Available when the documentation was examined. The addition of "+" to a number indicates that more than the number of data points listed were obtained, but the absolute number was not reported. Where more than one unit was used to report a measurement, the number of data points are recorded for each unit and are separated by a semi-colon. For example, if "#/s, #/ss" appears under Units and "2; 6" appears under No. Samples, then two organisms from the taxonomic group were enumerated from the entire sample (s) while six were enumerated from a subsample (ss).

Methodology: Collection

All methods used to collect data on each measurement are listed. Known details of the methods are included. The notations for the collection method are as follows:

Gear Type	Code and Description
ice corer	A; B; C where A = type of corer used; B = depth of ice sampled (tb = top to bottom of ice); C = other sampling details reported.
water samplers	A; B, C; D; E; F where A = type of sampler used (S = Specified, NS = Not Specified at all); B = depth sampled (S or NS; details are presented in Table 3); C = how depth was measured (MW = Meter Wheel only, WA = meter wheel and Wire Angle measure);

D = time of collection (S or NS; details are presented in Table 3);
E = period of time sampler was allowed to stabilize at specific depth;
F = other sampling details reported.

nets — A; B, C; D; E; F; G
where A = type of net used (S or NS);
B = tow depth (S or NS; details are presented in Table 3);
C = how depth was measured (MW or WA);
D = time of tow (S or NS; details are presented in Table 3);
E = tow speed (m/sec);
F = was a flow meter used;
G = other sampling details reported.

sediment corer or grab — A; B; C; D
where A = type of sampler used (S or NS);
B = station depth (S or NS; details are presented in Table 3);
C = time of collection (S or NS; details are presented in Table 3);
D = other sampling details reported.

It may be assumed that all details on sampling date, station location and station depth are available in Table 3 unless specified as follows:

S/NS: details were Specified only for some stations.

NS: details were Not Specified.

rel.: details reported were RELative, not specific (e.g., station location = off Cape Lyon, Darnley Bay, N.W.T.; sampling date = 24-hour period in Aug.-early Sep.).

?: details were estimated either from concurrent studies or, for station locations, from positions plotted on report maps.

Methodology: Storage

Some important details about methods and materials used in sample storage are provided. "F" refers to seawater-Formaldehyde solution.

Methodology: Analysis

Techniques used to analyse the samples are given. Unless ss (subsample) is prefaced by NS (not specified) or a specific number, it may be assumed that the size of subsample analysed, although variable (e.g., some were 10% of the sample, others were 25%, etc.), was reported. This column does not indicate statistical analyses performed on the data. "ABS taxonomic manual used" means that the manual compiled by E.H. Grainger and associates of the Arctic Biological Station was used to identify the zooplankton.

Precision/Accuracy

All approximations or estimations of the level of reliability (precision and/or accuracy) of the data are given, if available from the documentation. For most measurements, however, no specific indication of either precision or accuracy is available.

Where attempts were made in the study to ensure the precision and accuracy of the data even though these parameters were not specifically determined, the details are included in this column. For example, the column may indicate whether taxonomic references or authors of the nomenclature were cited. The inclusion of "ABS taxonomic manual used" in this column indicates that, although this manual is not available in published form, it has been compiled by experts in the field of zooplankton and is recognized here as a reputable taxonomic text. If equipment was calibrated or random checks were carried out on enumerators and identifiers, this information is provided.

In addition, where the other columns indicate that the data were presented in relative (i.e., imprecise) terms, the exact terminology is included in this column. For example, if the documentation indicates that there were "several" organisms of some species in the samples, while other species were also "found", then the details would be indicated in this column as "several, found".

In most cases, an attempt was made to provide some indication of "precision" in general terms. For example, if exact numbers of organisms were reported, "#" appears in this column; if mixed terminology was used to report ages of organisms found [e.g., copepodite #; copepodite; relative (i.e., juvenile, mature, etc.)], this is indicated in this column as "stage #; stage; juvenile, mature, etc."

Rating

A rating is assigned to each measurement in each data set using the criteria outlined in the flow chart in Section 6.4. It is important to note that the ratings apply to the taxonomic group as a collection of genera and species, unless only the taxonomic group was identified in the documentation. For example, if we considered it essential to know the magnification used when identification of a calanoid copepod species was being conducted but this was not reported, then the rating applied to the Calanoida is 2.

Common comments are indicated by the following numbered notes:

Note 1: Raw numbers (e.g., #/s that was not subsampled, #/ss) were not available. Only derived data (#/s that was subsampled, $\#/m^2$, $\#/m^3$) which were extrapolated from the raw data were reported. Insufficient information on collection and/or analytical methodology used was presented to indicate precision and accuracy of the measurements.

Note 2: Insufficient information is presented to determine whether measurement pertains to data set specimens or to others collected outside the study area during the research programme or to the species in general.

Remarks

Additional comments referring to sampling intensity and/or methodology are given in this column, as follows:

Remark 1 - It is unclear whether measurement relates to specimen(s) collected at study area stations or to all specimens collected during research programme.

Remark 2 - It is unclear whether measurement relates to specimen(s) collected at study area stations or to all specimens collected during research programme or to species in general.

Remark 3 - The measurement applies to specimen(s) collected during research programme but it is unclear whether it relates specifically to specimen(s) collected at study area stations.

Remark 4 - It is unclear whether measurement relates to this data set and/or another data set. (The other data set number is given.)

Species List

The underlined names indicate the Taxa Reported in Tables 1 (Section 8.1) and 2 (Section 8.2).

BACTERIA

B	Bacillus sp.
C	Clostridium sp.
D	Desulfovibrio sp.
E	Enterobacter sp.
K	Klebsiella sp.

PHYTOPLANKTON AND EPONTIC ALGAE[1]

Phylum: Bacillariophyta
Class: Bacillariophyceae
Order: Centrales

A	Arachnodiscus sp.
C	Cerataulina sp. H. Peragallo ex Schuett
Ca	Chaetoceros atlanticus Cleve
Cb	Chaetoceros borealis J.W. Bailey
Cbr	Chaetoceros brevis Schuett
Cc	Cyclotella caspia Grunow
Cce	Chaetoceros ceratosporum Ostenfeld
Ccm	Chaetoceros compressus Lauder
Ccn	Chaetoceros concavicornis Mangin
Cco	Chaetoceros constrictus Gran
Cct	Coscinodiscus centralis Ehrenberg
Ccv	Chaetoceros convolutus Castracane
Cd	Chaetoceros decipiens Cleve
Cdb	Chaetoceros debiles Cleve
Cdi	Chaetoceros diadema (Ehrenberg) Gran
Ce	Coscinodiscus excentricus Ehrenberg
Cf	Chaetoceros furcellatus J.W. Bailey
Cfr	Chaetoceros fragilis Meunier
Cg	Chaetoceros gracilis
CH	Chaetoceros sp. Ehrenberg
Ck	Coscinodiscus kryophilus Grunow
Cka	Chaetoceros karianus Grunow in Cleve & Grunow
Ckg	Coscinodiscus kuetzingii var. glacialis Grunow
Cl	Coscinodiscus limbatus (Ehrenberg) A. Schmidt
Clo	Chaetoceros lorenzianus Grunow
Cm	Chaetoceros mitra (J.W. Bailey) Cleve
Cn	Chaetoceros neogracile Van Landingham
CO	Coscinodiscus sp. Ehrenberg
Co	Coscinodiscus oculusiridis Ehrenberg
Cr	Coscinodiscus radiosus Grunow
Cs	Chaetoceros septentrionalis Oestrup
Csi	Chaetoceros simplex Ostenfeld
Cso	Chaetoceros socialis Lauder
Csu	Chaetoceros subsecundus
Ct	Coscinodiscus tuberculatus Greville
Cw	Chaetoceros wighamii Brightwell
CY	Cyclotella sp. Kuetzing
E	Eucampia sp. Ehrenberg
Eg	Eucampia groenlandica Cleve
Ez	Eucampia zodiacus Ehrenberg
M	Melosira sp. Agardh
Ma	Melosira arctica Dickie ex Ralfs in Pritchard
Mg	Melosira granulata (Ehrenberg) Ralfs in Pritchard
Mi	Melosira islandica O. Mueller
P	Porosira sp. Joergensen
Pg	Porosira glacialis (Grunow) Joergensen
R	Rhizosolenia sp. Ehrenberg
Rh	Rhizosolenia hebatata J.W. Bailey
Rhh	Rhizosolenia hebatata var. heimalis
Rs	Rhizosolenia setigera Brightwell
S	Skeletonema sp. Greville
Sa	Stephanodiscus astraea (Ehrenberg) Grunow in Cleve & Grunow
Sc	Skeletonema costatum (Greville) Cleve
Sh	Stephanodiscus hantzschii Grunow
ST	Stephanodiscus sp. Ehrenberg
T	Thalassiosira sp. Cleve
Ta	Trigonium arcticum (Brightwell) Cleve
Tb	Thalassiosira baltica (Grunow in Cleve & Grunow) Ostenfeld
Tbi	Thalassiosira bioculata var. exigua (Grunow) Hustedt
Td	Thalassiosira decipiens (Grunow) E. Joergensen

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[1] Taxonomic arrangement generally in accordance with Hsiao (1983).

Phylum: Bacillariophyta (cont'd)

Tg	Thalassiosira gravida Cleve
TI	Trigonium sp. Cleve
Tla	Thalassiosira lacustris
Tn	Thalassiosira nordenskioeldii Cleve
Tno	Triceratium nobile Witt
TR	Triceratium sp. Ehrenberg

Order: Pennales

A	Achnanthes sp. Bory
AC	Actinella sp.
Ac	Amphora crassa Gregory
Aco	Amphiprora concilians Cleve
Ad	Achnanthes delicatula (Kuetzing) Grunow in Cleve & Grunow
Ae	Amphora exsecta Grunow in Schmidt et al.
Af	Asterionella formosa Hassall
Ag	Amphiprora gigantea Grunow
Ags	Amphiprora gigantea var. septentrionalis (Grunow in Cleve & Grunow) Cleve
AH	Amphora sp. Ehrenberg
Ak	Amphiprora kjellmanii Cleve in Cleve & Grunow
Akk	Amphiprora kjellmanii var. kariana (Grunow in Cleve & Grunow) Cleve
Akr	Amphiprora kryophila Cleve
Aks	Amphiprora kjellmanii var. striolata (Grunow in Cleve & Grunow) Cleve
Al	Achnanthes lanceolata (Brebisson in Kuetzing) Grunow in Cleve & Grunow
Ala	Amphora laevis
Ale	Achnanthes lemmermannii Hustedt
All	Amphora laevis var. laevissima (Gregory) Cleve
Alm	Amphora laevissima var. minuta Cleve
AM	Amphipleura sp. Kuetzing
Am	Achnanthes minutissima Kuetzing
Ao	Amphora ovalis (Kuetzing) Kuetzing
AP	Amphiprora sp. Ehrenberg
Ap	Amphipleura pellucida (Kuetzing) Kuetzing
Aph	Amphiprora paludosa var. hyperborea (Grunow in Cleve & Grunow) Cleve
Apk	Amphora proteus var. kariana Grunow
Apl	Amphiprora paludosa Wm. Smith
Apr	Amphora proteus Gregory
Ar	Amphipleura rutilans (Trentepohl ex Roth) Cleve
AS	Asterionella sp. Ehrenberg
At	Achnanthes taeniata Grunow in Cleve & Grunow
B	Bacillaria sp. Gmelin
Bp	Bacillaria paradoxa Gmelin in Linnaeus
C	Ceratoneis sp. Ehrenberg
CA	Caloneis sp. Cleve
Ca	Ceratoneis arcus (Ehrenberg) Kuetzing
Cc	Cylindrotheca closterium (Ehrenberg) Reimann & Lewin
Ccs	Cocconeis costata Gregory
CE	Centronella sp.
Ck	Caloneis kryophila (Cleve) Cleve
Cl	Caloneis liber (Wm. Smith) Cleve
CM	Cymbella sp. Agardh
CO	Cocconeis sp. Ehrenberg
CY	Cylindrotheca sp. Rabenhorst
D	Diatoma sp. A.P. de Candolle
De	Diatoma elongatum (Lyngbye) Agardh
Dl	Diploneis litoralis (Donkin) Cleve
Dla	Diploneis litoralis var. arctica Cleve
Dlc	Diploneis litoralis var. clathrata (Oestrup) Cleve
Dln	Diploneis lineata (Donkin) Cleve
Do	Diploneis ovalis var. oblongella (Naegeli in Kuetzing) Cleve
DP	Diploneis sp. Ehrenberg
Ds	Diploneis smithii (Brebisson in Wm. Smith) Cleve
Dsr	Diploneis smithii var. rhombica Mereschkowsky
E	Eunotia sp. Ehrenberg

Phylum: Bacillariophyta (cont'd)

EP	Epithemia sp.
Ep	Eunotia polydentula
EU	Eunotia sp.
F	Fragillaria sp. Lyngbye
Fc	Fragillaria construens (Ehrenberg) Grunow
Fcy	Fragilariopsis cylindrus (Grunow) Helmcke & Krieger
Fo	Fragilariopsis oceanica (Cleve) Hasle
Foc	Fragillaria oceanica
Fp	Fragillaria pinnata Ehrenberg
FR	Fragilariopsis sp. Hustedt in Schmidt et al.
G	Gomphonema sp. Agardh
Ga	Grammatophora angulosa Ehrenberg
Ge	Gomphonema exiguum Kuetzing
Gep	Gomphonema exiguum var. pachycladum (Brebisson in Brebisson & Godey) Cleve
Gf	Gyrosigma fasciola (Ehrenberg) Griffith & Henfrey
Gk	Gomphonema kamtschaticum Grunow
Gw	Gyrosigma wansbeckii (Donkin) Cleve
GY	Gyrosigma sp.
L	Licmophora sp. Agardh
Ld	Licmophora dalmatica (Kuetzing) Grunow
Lg	Licmophora gracilis (Ehrenberg) Grunow
Lt	Licmophora tenuis (Kuetzing) Grunow
N	Navicula sp. Bory
Na	Navicula algida Grunow
Nac	Nitzschia acuminata (Wm. Smith) Grunow
Nan	Nitzschia angularis Wm. Smith
Nb	Navicula bahusiensis (Grunow in Van Heurck) Grunow
Nba	Navicula baculus Cleve
Nbb	Nitzschia brebissonii var. borealis Grunow in Cleve & Mueller
Nbi	Neidium bisulcatum (Lagerstedt) Cleve
Nc	Navicula crucigeroides Hustedt
Nca	Navicula cancellata Donkin
Ncc	Nitzschia acicularis (Kuetzing) Wm. Smith
Ncl	Nitzschia closterium (Ehrenberg) Wm. Smith
Ncp	Navicula cluthensis var. pagophila Grunow
Ncr	Navicula crassirostris Grunow in Cleve & Grunow
Ncy	Nitzschia cylindrus (Grunow) Hasle
Nd	Navicula delicatula Cleve
Nda	Nitzschia diaphana Cleve
Ndc	Navicula directa var. cuneata Oestrup
Nde	Navicula decipiens O'Meara
Ndi	Navicula digitoradiata (Gregory) Ralfs in Pritchard
Ndn	Nitzschia distans (Wm. Smith) Ralfs in Pritchard
Ndp	Nitzschia dissipata
Ndr	Navicula directa (Wm. Smith) Ralfs in Pritchard
Nds	Nitzschia delicatissima
Ndt	Nitzschia distans var. erratica Cleve
NE	Neidium sp. Pfitzer
Nf	Navicula forcipata Greville
Nfr	Nitzschia frigida Grunow in Cleve & Grunow
Ng	Navicula gastrum var. jenisseyensis Grunow
Nga	Navicula granii ((E. Joergensen) Gran
Ngd	Nitzschia gelida Cleve & Grunow in Cleve & Mueller
Nge	Navicula gelida Grunow
Ngg	Navicula gregaria
Ngl	Navicula glacialis (Cleve) Grunow
Ngn	Nitzschia grunowii Hasle
Ngr	Nitzschia gruendleri Grunow in Cleve
Ngs	Navicula gastrum
Nh	Navicula hodgeana Patrick & Freese
Nhu	Navicula humerosa Brebisson in litt. ex Wm. Smith
Nhy	Nitzschia hybrida Grunow in Cleve & Grunow
Nk	Navicula kariana Grunow in Cleve & Grunow
Nkd	Navicula kariana var. detersa Grunow in Cleve & Mueller
Nkj	Navicula kjellmanii (Cleve in Cleve & Grunow) Cleve
Nl	Navicula lineola Grunow
Nla	Nitzschia laevissima Grunow in Cleve & Mueller

Phylum: Bacillariophyta (cont'd)

Nle	Nitzschia lecointei Van Heurck
Nlg	Nitzschia longissima (Brebisson in Kuetzing) Grunow
Nli	Nitzschia linearis (Agardh) Wm. Smith
Nlt	Nitzschia linearis var. tenuis (Wm. Smith) Grunow
Nm	Navicula marina Ralfs in Pritchard
Nme	Navicula membranacea Cleve
Nn	Navicula novadecipiens Hustedt
No	Navicula oestrupi Cleve
Np	Navicula pseudocrassirostris Hustedt
Npe	Navicula perlucens Oestrup
Npl	Navicula pelagica Cleve
Npo	Nitzschia polaris Grunow in Cleve & Mueller
Npy	Navicula pygmaea Kuetzing
Nq	Navicula quadratarea A. Schmidt
Nqu	Navicula quadripedis Cleve-Euler
Nr	Navicula rhynchocephala Kuetzing
Nre	Navicula recurvata Gran
Ns	Navicula salinarum Grunow in Cleve & Mueller
Nsa	Navicula superba var. crassa
Nsb	Navicula subinflata Grunow in Cleve & Mueller
Nsc	Navicula scopulorum Brebisson in Kuetzing
Nse	Nitzschia seriata Cleve
Nsi	Nitzschia sigma (Kuetzing) Wm. Smith
Nso	Navicula solitaria Cleve
Nsp	Navicula spicula (Hickie) Cleve
Nsr	Navicula superba Cleve
Nss	Navicula stuxbergii var. subglabra Oestrup
Nsu	Navicula scutelloides Wm. Smith
NT	Nitzschia sp. Hassall
Nt	Navicula trigonocephala Cleve
Ntd	Navicula transitans var. derasa (Grunow in Cleve & Grunow) Cleve
Nte	Navicula transitans var. erosa (Cleve) Cleve
Ntg	Navicula trigonocephala var. depressa Oestrup
Nti	Navicula transitans var. incudiformis (Grunow in Cleve) Cleve
Ntr	Navicula transitans Cleve
Nv	Navicula vanhoeffenii Gran
Nvs	Navicula valida Cleve & Grunow
Nvm	Navicula valida var. minuta Cleve
P	Pinnularia sp. Ehrenberg
Pa	Pinnularia ambigua Cleve
Pag	Pleurosigma angulatum (Quekett) Wm. Smith
Pan	Pleurosigma antarcticum Heiden & Kolbe
Pc	Pleurosigma clevei Grunow in Cleve & Grunow
PE	Peronia sp.
Pe	Pleurosigma elongatum
Pg	Pleurosigma glaciale Cleve
PL	Pleurosigma sp. Wm. Smith
Pmi	Pinnularia quadratarea var. minima (Oestrup) Boyer
Pq	Pinnularia quadratarea (Schmidt) Cleve
Pqb	Pinnularia quadratarea var. bicontracta (Oestrup) Heiden in Schmidt et al.
Pqc	Pinnularia quadratarea var. constricta (Oestrup) Heiden in Schmidt et al.
Pqd	Pinnularia quadratarea var. densestriata Cleve
Pql	Pinnularia quadratarea var. leptostauron (Cleve) Cleve
Pqm	Pinnularia quadratarea var. maxima (Oestrup) Boyer
Pqs	Pinnularia quadratarea var. stuxbergii (Cleve in Cleve & Grunow) Cleve
Pqu	Pinnularia quadratarea var. bicuneata Heiden & Kolbe
Ps	Pleurosigma salinarum (Grunow) Grunow in Cleve & Grunow
Psb	Pinnularia quadratarea var. subcontinua (Cleve) Cleve
Psc	Pinnularia quadratarea var. subconstricta (Oestrup) Heiden in Schmidt et al.
Psd	Pinnularia semiinflata var. decipiens (Cleve) Gran
Pse	Pinnularia semiinflata
Psm	Pleurosigma stuxbergii var. minor Grunow
Pst	Pleurosigma strigosum Wm. Smith

Phylum: Bacillariophyta (cont'd)

Psu	Pleurosigma stuxbergii Cleve & Grunow
R	Rhabdomena sp.
RH	Rhaphoneis sp.
S	Stauroneis sp. Ehrenberg
Sa	Synedra acus var. radians (Kuetzing) Hustedt
Sf	Synedra fasciculata (Agardh) Kuetzing
Sh	Synedra hyperborea Grunow
Si	Stenoneis inconspicua var. baculus (Cleve in Cleve & Mueller) Cleve
Sj	Surirella japonica A. Schmidt in Schmidt et al.
So	Surirella ovata Kuetzing
Soe	Surirella oestrupi Gran
Sq	Stauroneis quadripedis (Cleve-Euler) Hendey
ST	Stenoneis sp. Cleve
St	Synedra tabulata (Agardh) Kuetzing
Sto	Synedra tabulata var. obtusa (Pantocsek) Hustedt
SU	Surirella sp. Turpin
SY	Synedra sp. Ehrenberg
T	Tabellaria sp. Ehrenberg
TA	Thalassiothrix sp.
Tfe	Tabellaria fenestrata (Lyngbye) Kuetzing
Tfl	Tabellaria flocculosa (Roth) Kuetzing
TH	Thalassionema sp. Grunow
Tl	Tetracyclus lacustris
Tm	Tropidoneis maxima (Gregory) Cleve
Tn	Thalassionema nitzschioides (Grunow) Van Heurck
Tnt	Thalassiothrix nitzschioides
TR	Tropidoneis sp. Cleve

Phylum: Chlorophyta

Class: Chlorophyceae

B	Bolbocoleon sp.
Bp	Bolbocoleon piliferum Pringsheim
C	Chlamydomonas sp. Ehrenberg
CA	Carteria sp.
Cc	Chaetomorpha capillaris (Kuetzing) Borq.
CH	Chaetomorpha sp.
CL	Cladophora sp.
Cm	Chaetomorpha melagonium (Weber & Mohr) Kuetzing
Ew	Ectochaete wittrockii (Wille) Kylin
P	Pediastrum sp.
PE	Percursaria sp.
Pe	Percursaria percursa (C. Agardh) Rosenv.
Pr	Protoderma sp.
S	Scenedesmus sp. Meyen
SP	Spirogyra sp.

Phylum: Phaeophyta

Class: Phaeophyceae

F	Fucus sp.
Fd	Fucus distichus Linnaeus
Pl	Pilayella littoralis (L.) Kjellm.
S	Sorapion sp.
Sa	Sphacelaria arctica Harv.
Sk	Sorapion kjellmanii (Willie) Rosenv.
ST	Stictyosiphon sp.
St	Stictyosiphon tortilis (Rupr) Reinke

Phylum: Chrysophyta

Class: Chrysophyceae

C	Chrysococcus sp.
CH	Chrysomonas sp.
Cr	Chrysococcus rufescens
Db	Dinobryon balticum (Schuett) Lemmermann
DN	Dinobryon sp. Ehrenberg
DS	Distephanus sp. Haeckel
Ds	Distephanus speculum (Ehrenberg) Haeckel

Phylum: Chrysophyta (cont'd)
- **Dsp** Dictyocha speculum
- **E** Ebria sp. Borgert
- **Et** Ebria tripartita (Schumann) Lemmermann
- **M** Mallomonas sp.

Phylum: Cyanophyta
Class: Cyanophyceae
- **A** Anabaena sp.
- **C** Coelosphaerium sp. Naegeli
- **CA** Calothrix sp.
- **O** Oscillatoria sp.

Phylum: Euglenophyta
Class: Euglenophyceae
- **E** Euglena sp. Ehrenberg
- **Ep** Euglena proxima Dangeard
- **T** Trachelomonas sp. Ehrenberg

Phylum: Pyrrophyta
Class: Dinophyceae
- **A** Amphidinium sp. Claparede & Lachmann
- **C** Ceratium sp.
- **Ca** Ceratium arcticum (Ehrenberg) Cleve
- **Ct** Ceratium tripos (O.F. Mueller) Nitzsch
- **D** Dinophysis sp. Ehrenberg
- **Da** Dinophysis acuminata Claparede & Lachmann
- **Dar** Dinophysis arctica Mereschkowsky
- **G** Goniaulax sp. Diesing
- **Gc** Goniaulax catenata (Levander) Peters
- **GR** Gyrodinium sp.
- **GY** Gymnodinium sp. Ehrenberg
- **P** Peridinium sp. Ehrenberg
- **Pb** Peridinium brevipes Paulsen
- **Pbe** Peridinium belgium
- **Pc** Peridinium cerasus Paulsen
- **Pd** Protoperidinium depressum
- **Pm** Peridinium minusculum Pavillard
- **Pp** Peridinium pellucidum (Bergh) Schuett

Phylum:
Class: Craspedophyceae

Phylum: Cryptophyta
Class: Cryptophyceae
- **R** Rhodomonas sp.

ZOOPLANKTON AND EPONTIC INVERTEBRATES[2]

Phylum: Protozoa
  Subphylum: Sarcomastigophora
    Superclass: Mastigophora
    Superclass: Sarcodina
      Class: Rhizopodea
        Subclass: Lobosia
          Order: Amoebida
        Subclass: Granuloreticulosia
          Order: Foraminiferida
- **Cf** Cornuspira foliacea (Philippi)
- **Gp** Globorotalia pachyderma (Ehrenberg)
- **Qs** Quinqueloculina seminulum Linne
- **Sb** Spiroplecta biformis (Parker & Jones)
- **Tn** Trochammina nana (H.B. Brady)

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[2] Taxonomic arrangement generally in accordance with Shih et al. (1971) and Barnes (1974).

- Phylum: Protozoa (cont'd)
    - Class: Actinopodea
        - Subclass: Radiolaria
    - Subphylum: Ciliophora
        - Class: Ciliata
            - Subclass: Spirotrichia
                - Order: Heterotrichida
                    - **S** Spirostomum sp.
                - Order: Tintinnida
- Phylum: Porifera
- Phylum: Cnidaria
    - Class: Hydrozoa
        - **Hp** Homoeonema platygonon
        - Order: Anthomedusae
            - **B** Bougainvillea
            - **Cb** Calycopsis birulai
            - **Cf** Corymorpha flammea
            - **Eb** Eumedusa birulai (Linko)
            - **Ef** Euphysa flammea (Linko)
            - **G** Gemmaria sp.
            - **H** Halitholus sp.
            - **Hc** Halitholus cirratus Hartlaub
            - **Hp** Halitholus pauper Hartlaub
            - **HY** Hydractinia sp.
            - **L** Leukartiara sp.
            - **Ln** Leukartiara nobilis Hartlaub
            - **Mh** Margelopsis hartlaubi
            - **Pk** Polyorchis karafutoensis
            - **Py** Perigonimus yoldia-arcticae
            - **Ro** Rathkea octopunctata (M. Sars)
            - **S** Sarsia sp.
            - **Sp** Sarsia princeps (Haeckel)
        - Order: Leptomedusae
            - **M** Mitrocomella sp.
            - **Mo** Melicertum octocostatum (M. Sars)
            - **Mp** Mitrocomella polydiademata (Romanes)
            - **O** Obelia sp.
            - **Pl** Ptychogena lactea A. Agassiz
            - **Pla** Phialidium languidum (A. Agassiz)
            - **Tm** Tiaropsis multicirrata (M. Sars)
        - Order: Limnomedusae
            - **Gv** Gonionemus vertens A. Agassiz
        - Order: Trachymedusae
            - **A** Aglantha sp.
            - **Ac** Aglantha digitale (O.F. Mueller) var. camtschatica (Brandt) comb. nov.
            - **Ad** Aglantha digitale (O.F. Mueller)
        - Order: Narcomedusae
            - **A** Aeginopsis sp.
            - **Ac** Aegina citrea Eschschotz
            - **AE** Aegina sp.
            - **Al** Aeginopsis laurenti Brandt
        - Order: Siphonophora
            - **A** Agalma
            - **C** Chrysaora sp.
            - **Da** Dimophyes arctica (Chun)
            - **Mb** Muggiaea bargmannae Totton
    - Class: Scyphozca
        - Order: Stauromedusae
            - **Cc** Cyanea capillata (Linne)
    - Class: Anthozoa
- Phylum: Ctenophora
    - Class: Tentaculata
        - Order: Cydippida
            - **Mo** Mertensia ovum (Fabricius)
            - **Pp** Pleurobrachia pileus (O.F. Mueller)

Phylum: Ctenophora (cont'd)
Class: Nuda
Order: Beroida
B Beroe sp.
Bc Beroe cucumis Fabricius

Phylum: Platyhelminthes
Class: Turbellaria
Class: Trematoda

Phylum: Rotifera
Class: Monogononta
Order: Ploima
A Asplanchna sp.
B Brachionus sp.
K Kellicottia sp.
Kc Keratella cochlearis
KE Keratella sp.
Kl Kellicottia longspina
N Notholca sp.
Sj Synchaeta johanseni Harring

Phylum: Nematoda

Phylum: Sipuncula
P Prionglossa sp.

Phylum: Mollusca
Class: Gastropoda
Subclass: Opisthobranchia
Order: Thecosomata
S Spiratella sp.
Sh Spiratella helicina (Phipps)
Order: Gymnosomata
C Clione sp.
Cl Clione limacina (Phipps)
Class: Pelecypoda

Phylum: Annelida
Class: Polychaeta
Order: Phyllodocida
Order: Spionida
Class: Oligochaeta

Phylum: Tardigrada

Phylum: Arthropoda
Class: Arachnida
Order: Acarina
Class: Crustacea
Subclass: Branchiopoda
Order: Diplostraca
(Suborder: Conchostraca)
(Suborder: Cladocera)
B Bosmina sp.
Bl Bosmina longirostris (O.F. Mueller)
D Daphnia sp.
Dl Daphnia longiremis Sars
Dm Daphnia middendorffiana
E Evadne sp.
En Evadne nordmanni Loven
Lk Leptodora kindtii (Focke)
Np Nymphon pixellae
P Podon sp.
Pl Podon leuckarti (G.O. Sars)
Subclass: Ostracoda
Order: Myodocopa
C Conchoecia sp.
Cb Conchoecia borealis maxima (Brady & Norman)

Phylum: Arthropoda (cont'd)

Pg Philomedes globosus (Lilljeborg)

Subclass: Copepoda

Order: Calanoida

A Acartia sp.
Ab Acartia bifilosa (Giesbrecht)
Ac Acartia clausi Giesbrecht
Al Acartia longiremis (Lilljeborg)
C Calanus sp.
Cc Calanus cristatus Kroyer
Cf Calanus finmarchicus (Gunnerus)
Cg Calanus glacialis Yaschnov
Ch Calanus hyperboreus Kroyer
Cha Centropages hamatus (Lilljeborg)
Co Chiridius obtusifrons G.O. Sars
Cp Calanus plumchrus Marukawa
Ct Calanus tonsus
D Diaptomus sp.
Da Diaptomus ashlandi Marsh
Db Drepanopus bungei G.O. Sars
Dm Diaptomus minutus Lilljeborg
Do Diaptomus oregonensis Lilljeborg
Dp Diaptomus pribilofensis Juday & Muttkowski
Ds Diaptomus sicilis S.A. Forbes
Dt Derjuginia tolli (Linko)
E Epischura sp.
Ea Eurytemora arctica Wilson & Tash
Eb Eucalanus bungii bungii Johnson
Ec Eurytemora composita Keiser
Eca Eurytemora canadensis Marsh
Eg Euchaeta glacialis Hansen
Egr Eurytemora gracilicauda Akatova
Eh Eurytemora herdmani Thompson & Scott
El Epischura lacustris S.A. Forbes
En Euchaeta norvegica Boeck
Ene Epischura nevadensis Lilljeborg
ER Eurytemora sp.
Er Eurytemora raboti Richard
Eri Eurytemora richingsi Heron & Damkaer
EU Euchaeta sp.
Gb Gaidius brevispinus (G.O. Sars)
Gt Gaidius tenuispinus (G.O. Sars)
Hn Heterorhabdus norvegicus (Boeck)
Hs Heterocope septentrionalis Juday & Muttkowski
L Limnocalanus sp.
Lm Limnocalanus macrurus G.O. Sars
M Metridia sp.
Ml Metridia longa (Lubbock)
Mlu Metridia lucens Boeck
Mp Metridia pacifica Brodsky
Mpo Mormonilla polaris G.O. Sars
Mpy Microcalanus pygmaeus (G.O. Sars)
P Pseudocalanus sp.
Pm Pseudocalanus minutus (Kroyer)
Pp Paracalanus parvus (Claus)
S Spinocalanus sp.
Sb Scaphocalanus brevicornis G.O. Sars
Sc Senecella calanoides
SC Scolecithricella sp.
Sl Spinocalanus longicornis G.O. Sars
Sm Scaphocalanus magnus (Scott)
Smi Scolecithricella minor (Brady)
SP Scaphocalanus sp.
Tb Temorites brevis G.O. Sars

Order: Harpacticoida

A Ameira sp.
Aa Amonardia arctica (Scott)
D Danielssenia sp.

Phylum:	Arthropoda (cont'd)		
		Df	Danielssenia fusiformis (Brady)
		Ds	Danielssenia stefanssoni Willey
		E	Ectinosoma sp.
		Ef	Ectinosama finmarchicum (Scott)
		En	Ectinosoma neglectum G.O. Sars
		H	Harpacticus sp.
		Hs	Harpacticus superflexus Willey
		Hu	Harpacticus uniremis Kroyer
		L	Laophonte sp.
		M	Microsetella sp.
		P	Pseudobradya sp.
		Pm	Pseudobradya minor (T. & A. Scott)
		T	Tachidius sp.
		Tf	Tisbe furcata (Baird)
		Zs	Zaus spinatus Goodsir
Order:	Cyclopoida		
		C	Cyclops sp.
		Cb	Cyclops bicuspidatus
		Cs	Cyclopina schneideri Scott
		Csc	Cyclops scutifer Sars
		Ct	Cyclops thomasi S.A. Forbes
		Cv	Cyclops vernalis Fischer
		CY	Cyclopina sp.
		O	Oncaea sp.
		Ob	Oncaea borealis G.O. Sars
		Oh	Oithona helgolandica Claus
		OI	Oithona sp.
		Om	Oncaea minuta Giesbrecht
		On	Oncaea notopus Giesbrecht
		Os	Oithona similis Claus
Order:	Monstrilloida		
		M	Monstrilla sp.
Subclass:	Cirripedia		
Order:	Thoracica		
		B	Balanus sp.
Subclass:	Malacostraca		
Order:	Mysidacea		
		M	Mysis sp.
		Ml	Mysis litoralis (Banner)
		Mo	Mysis oculata
		Mr	Mysis relicta Loven
Order:	Cumacea		
		D	Diastylis sp.
		Do	Diastylis oxyrhyncha
		Dr	Diastylis rathkei (Kroyer)
Order:	Isopoda		
		Me	Mesidotea entomon (Linnaeus)
Order:	Amphipoda		
		A	Apherusa sp.
		Ab	Acanthostepheia behringiensis (Lockington)
		Ag	Apherusa glacialis (Hansen)
		Al	Aceroides latipes (G.O. Sars)
		Am	Acanthostepheia malmgreni (Goes)
		An	Anonyx nugax (Phipps)
		B	Boeckosimus sp.
		Cd	Chironesimus debruyni
		Dp	Dulichia porrecta (Bate)
		G	Gammaracanthus sp.
		GA	Gammarus sp.
		Gl	Gammaracanthus loricatus (Sabine)
		Gs	Gammarus setosus Dementieva
		Gw	Gammarus wilkitzki (Birula)
		H	Hyperia sp.
		Hg	Hyperia galba (Montagu)
		Hm	Hyperia medusarum (Mueller)
		Hme	Haliragea megalops (Bucholtz)
		Hym	Hyperoche medusarum (Kroyer)
		M	Metopa sp.

Phylum: Arthropoda (cont'd)
MO Monoculodes sp.
O Oedocerus sp.
Og Onisimus glacialis
ON Onisimus sp.
OR Orchomene sp.
P Pontoporeia sp.
Pa Pontoporeia affinis Lindstrom
Pab Parathemisto abyssorum Boeck
Pg Pseudalibrotus glacialis G.O. Sars
Pi Pontogeneia inermis (Kroyer)
Pl Parathemisto libellula (Lichtenstein)
Plt Pseudalibrotus littoralis (Kroyer)
Pp Paroediceros propinquus (Goes)
Order: Euphausiacea
E Euphausia sp.
Ti Thysanoessa inermis (Kroyer)
Tl Thysanoessa longipes Brandt
Tr Thysanoessa raschi (M. Sars)
Order: Decapoda
Suborder: Natantia
Section: Caridea
Eg Eualus gaimardi (H. Milne-Edwards)
P Pandalus sp.
S Sabinea sp.
Ss Sabinea septemcarinata (Sabine)
Suborder: Reptantia
Section: Brachyura
Class: Insecta
Order: Ephemeroptera
(Family: Heptogeniidae)
Order: Diptera
(Family: Chironomidae)
Hs Heterocope septemtrionalis
T Tendipididae sp.

Phylum: Bryozoa

Phylum: Chaetognatha
Eh Eukrohnia hamata (Mobius)
Se Sagitta elegans Verrill
Sm Sagitta maxima (Conant)

Phylum: Echinodermata
Class: Asteroidea
Class: Echinoidea
Class: Ophiuroidea

Phylum: Chordata
Class: Copelata
F Fritillaria sp.
Fb Fritillaria borealis Lohmann
O Oikopleura sp.
Ov Oikopleura vanhoeffeni Lohmann

ICHTHYOPLANKTON

Phylum: Vertebrata
Class: Osteichthyes
Subclass: Neopterygii
Order: Isospondyli
Suborder: Clupeoidea
Family: Clupeidae
PCHR Clupea harengus pallasi Valenciennes (Pacific herring)
Family: Osmeridae
RNSM Osmerus mordax (Mitchill) (rainbow smelt)

Order: Anacanthini
Family: Gadidae
ARCD Boreogadus saida (arctic cod)
SFCD Eleginus gracilis (Tilesius) (saffron cod)
Order: Percomorphi
Suborder: Blennioidea
Family: Stichaeidae
LUMP Lumpenus sp. (prickleback)
Suborder: Cottoidea
Family: Cottidae
FHSC Myoxocephalus quadricornis quadricornis (fourhorn sculpin)
SC Myoxocephalus sp. (sculpin)
Order: Heterosomata
Family: Pleuronectidae
STFL Platichthys stellatus (starry flounder)

Description of Gear Type Codes

BN - Bongo Net
BRN - BiRge closing Net
C-23 PN - Coarse-silk conical Plankton Net
CB - Clarke Bumpus net
F-23PN - Finer silk conical Plankton Net
FN - Faber Net
IKT - Isaacs-Kidd midwater Trawl
KB - Kemmerer Bottle
MN - Miller Net
N - Neuston net
NB - Niskin Bottle
NBS - Niskin Bag-Sampler
NN - Nansen-type Net
NS - Not Specified
PB - Plastic Bottle
PN - conical Plankton Net
PP - PhytoPlankton net
PVC - PVC sampler
S corer - Sipre ice corer
S-30 PN - Silk conical Plankton Net
SN - Scor Net
STN - STramin Net
Subsampler - nuclear submarine sampler
VD - Van Dorn sampler
WN - Wisconsin Net

Note: In general, numerical prefixes to the codes indicate mesh size (u) and mouth diameter (cm) of the plankton net. The suffixes H, O and V indicate Horizontal, Oblique and Vertical tows, respectively. Further details are provided in Appendix I.

TABLE 2: MEASUREMENTS, METHODOLOGY AND QUALITY RATING

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
13-0001	PHYTOPLANKTON										
	Centrales CH,CJ,CO,Co, Cr,Ct, Mg	#/F-23 PNH	#/s	1	relative	S; S; NS; NS; NS; loc. rel.	NS	NS	found	2:all	
		#/S-30 PNV?	#/s	1	relative	NS; S; NS; NS; NS	NS	NS	found, rather plentiful	2:all	NS whether collected in plankton net or dredge
		identification	genus, species	2	relative	S/NS; S; NS; NS; NS; loc. rel.	NS	NS	found, rather plentiful; taxonomic references stated	2:all	
	Pennales Lt,Na,Nba,Ng Npo,Nq,Nt,Pg	#/C-23 PNH	#/s	1	relative	S; S; NS; NS; NS	NS	NS	found	2:all	
		#/S-30 PNV?	#/s	1	relative	NS; S; NS; NS; NS	NS	NS	found; NS	2:all	Nba,Ng: NS whether collected in plankton net or dredge
		identification	species	2	relative	S/NS; S; NS; NS; NS	NS	NS	found; taxonomic references stated	2:all	
		length, overall	mm	1	1	S/NS; S; NS; NS; NS	NS	NS	NS	2:all	
	Dinophyceae C	#/F-23 PNH	#/s	1	relative	S; S; NS; NS; NS	NS	NS	found	2:all	
		identification	genus	1	relative	S; S; NS; NS	NS	NS	found	2:all	
	ZOOPLANKTON										
	Foreminiferida Cf,Qs,Sb,Tn	#/S-30 PNV	#/s	1	3+	S; S; NS; NS; NS	NS	NS	#, few, several	2:all	
		identification	species	1	3+	S; S; NS; NS; NS	NS	NS	taxonomic references stated	2:all	
		length, overall	mm	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, overall	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		colour	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
	Trachymedusae Ad	#/S-30 PNV	#/s	1	2	S; S; NS; NS; NS	NS	NS	#	2:all	
		identification	species	1	2	S; S; NS; NS; NS	NS	NS	taxonomic references stated	2:all	
		height, overall	mm	1	NS	NS	NS	NS	NS	2:all	Remark 1
	Ploima Sj	#/T-23 PNH	#/s	1	relative	S; S; NS; NS; NS; loc. rel.	NS	NS	large numbers	2:all	
		identification	species	1	relative	S; S; NS; NS; NS; loc. rel.	NS	NS	large numbers; NS	2:all	
		length, overall	u	1	NS	NS	NS	NS	NS	2:all	Remark 2
		length, foot	u	1	NS	NS	NS	NS	NS	2:all	Remark 2
		length, toes	u	1	NS	NS	NS	NS	NS	2:all	Remark 2
		width, overall	u	1	NS	NS	NS	NS	NS	2:all	Remark 2
		shape, overall	NA	1	NS	NS	NS	NS	NS	2:all	Remark 2
		shape, head	NA	1	NS	NS	NS	NS	NS	2:all	Remark 2
		shape, dors. antennae	NA	1	NS	NS	NS	NS	NS	2:all	Remark 2
		shape, lat. antennae	NA	1	NS	NS	NS	NS	NS	2:all	Remark 2
		shape, foot	NA	1	NS	NS	NS	NS	NS	2:all	Remark 2

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
13-0001 (cont'd)	Calanoida Al,Cf,Ch,Eh, En,Ml,Pm	#/C-23 PNH	#/s	1	79	S; S; NS; NS; NS; loc. rel.	NS	NS	NS	2:all	
		#/C-23 PNV	#/s	1	3-4	S; S; NS; NS; NS	NS	NS	range	2:all	
		#/F-23 PNH	#/s	1	13	S; S; NS; NS; NS; loc. rel.	NS	NS	NS	2:all	
		#/S-30 PNV	#/s	1	4+	S; S; NS; NS; NS	NS	NS	#, present	2:all	
		identification	species	4	99+	S; S; NS; NS; NS; loc. rel.	NS	NS	taxonomic references stated for some	2:all	
		length, overall	mm	2	6	S; S; NS; NS; NS	NS	NS	NS	2:all	
		age	adult; relative	3	47+	S; S; NS; NS; NS; loc. rel.	NS	NS	stage; immature, young, juvenile, mature	2:all	
		sex	NA	4	90+	S; S; NS; NS; NS	NS	NS	NS	2:all	
		spermatophores	presence	1	2	S; S; NS; NS; NS; loc. rel.	NS	NS	NS	2:all	
	Harpacticoida As	#/C-23 PNH	#/s	1	1	S; S; NS; NS; NS; loc. rel.	NS	NS	NS	2:all	
		identification	species	1	1	S; S; NS; NS; NS; loc. rel.	NS	NS	taxonomic references stated for some	2:all	
		length, overall	mm	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, overall	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, A1	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, A2	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, Mxp2	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, P1	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, P2	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, P4	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		shape, P5	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
		sex	NA	1	NS	NS	NS	NS	NS	2:all	Remark 1
	Cyclopoida Os	#/C-23 PNH	#/s	1	20	S; S; NS; NS; NS; loc. rel.	NS	NS	#	2:all	
		#/S-30 PNV	#/s	1	6+	S; S; NS; NS; NS	NS	NS	#, present	2:all	
		identification	species	2	26+	S; S; NS; NS; NS; loc. rel.	NS	NS	taxonomic references stated for some	2:all	
	Copelata F	#/C-23 PNH	#/s	1	relative	S; S; NS; NS; NS; loc. rel.	NS	NS	present	2:all	
		identification	genus	1	relative	S; S; NS; NS; NS; loc. rel.	NS	NS	present; NS	2:all	
50-0001	ZOOPLANKTON										
	Anthomedusae Ln	#/50 NNV	#/haul	2	2	S; S; S; NS; NS	NS	METHOD 1	#	2:all	
		identification	species	2	2	S; S; S; NS; NS	NS	NS	author cited	2:all	
		width, overall	mm	NS	7	NS	NS	NS	range	2:all	Remark 3

METHOD 1: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor and standardized to 100-0m vertical hauls.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
50-0001 (cont'd)		height, overall	mm	NS	7	NS	NS	NS	range	2:all	Remark 3
	Trachymedusae Ac	#/50 NNV	#/haul	14	304	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#	2:all	
		identification	species	14	304	S; S; S; NS; NS; S/NS stn. depth	NS	NS	author cited	2:all	
		length, gonads	mm	NS	NS	NS	NS	NS	range	2:all	Remark 3; applies also to 51-0001
		width, overall	mm	NS	NS	NS	NS	NS	range	2:all	Remark 3; applies also to 51-0001
		height, overall	mm	NS	NS	NS	NS	NS	range	2:all	Remark 3; applies also to 51-0001
		shape, overall	NA	NS	NS	NS	NS	NS	NS	2:all	Remark 3; applies also to 51-0001
	Narcomedusae Al	#/50 NNV	#/haul	7	35	S; S; S; NS; NS	NS	METHOD 1	#	2:all	
		identification	species	7	35	S; S; S; NS; NS	NS	NS	author cited	2:all	
		width, overall	mm	NS	NS	NS	NS	NS	range	2:all	Remark 3; applies also to 51-0001
	Thecosomata Sh	#/50 NNV	#/haul	14	4327+	S; S; S; NS; NS	NS	METHOD 1	#, present	2:all	
		identification	species	14	4327+	S; S; S; NS; NS	NS	NS	NS	2:all	
	Pelecypoda un.	#/50 NNV	#/haul	6	87+	S; S; S; NS; NS	NS	METHOD 1	#, present	2:all	
		identification	class	6	87+	S; S; S; NS; NS	NS	NS	NS	2:all	
	Annelida un.	#/50 NNV	#/haul	4	275+	S; S; S; NS; NS	NS	METHOD 1	#, present	2:all	
		identification	phylum	4	275+	S; S; S; NS; NS	NS	NS	NS	2:all	
		age	larva	4	275+	S; S; S; NS; NS	NS	NS	stage	2:all	
	Ostracoda un.	#/50 NNV	#/haul	5	23	S; S; S; NS; NS	NS	METHOD 1	#	2:all	
		identification	subclass	5	23	S; S; S; NS; NS	NS	NS	NS	2:all	
	Copepoda un.	#/50 NNV	#/haul	13	11136	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#	2:all	
		identification	subclass	13	11136	S; S; S; NS; NS; S/NS stn. depth	NS	NS	NS	2:all	
		age	nauplius	13	11136	S; S; S; NS; NS; S/NS stn. depth	NS	NS	stage	2:all	
	Calanoida C,Cc,Cf,Ch,En, Ml,Mpy,Pm	#/50 NNV	#/haul	14	18409+	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#, present	2:all	
		identification	species	14	18409+	S; S; S; NS; NS; S/NS stn. depth	NS	NS	author cited	2:all	

METHOD 1: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor and standardized to 100-0m vertical hauls.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
50-0001 (cont'd)		length, overall	mm	1	1	S; S; S; NS; NS	NS	NS	NS	2:all	
		age	copepodite #; relative	13	1856+	S; S; S; NS; NS; S/NS stn. depth	NS	NS	stage #; juvenile	2:all	
		sex	NA	1	1	S; S; S; NS; NS	NS	NS	NS	2:all	
	Cyclopoida 0, OI	#/50 NNV	#/haul	14	64024+	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#, present	2:all	
		identification	genus	14	64024+	S; S; S; NS; NS; S/NS stn. depth	NS	NS	NS	2:all	
	Cirripedia un.	#/50 NNV	#/haul	5	99	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#	2:all	
		identification	subclass	5	99	S; S; S; NS; NS; S/NS stn. depth	NS	NS	NS	2:all	
		age	nauplius, cyprid	5	99	S; S; S; NS; NS S/NS stn. depth	NS	NS	stage	2:all	
	Amphipoda un.	#/50 NNV	#/haul	8	32	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#	2:all	
		identification	order	8	32	S; S; S; NS; NS; S/NS stn. depth	NS	NS	NS	2:all	
	Ophiuroidea un.	#/50 NNV	#/haul	3	relative	S; S; S; NS; NS	NS	METHOD 1	present	2:all	
		identification	subclass	3	relative	S; S; S; NS; NS	NS	NS	present	2:all	
		age	ophiopluteus	3	NS	S; S; S; NS; NS	NS	NS	stage	2:all	
	Copelata un.	#/50 NNV	#/haul	10	1696+	S; S; S; NS; NS; S/NS stn. depth	NS	METHOD 1	#, present	2:all	
		identification	class	10	1696+	S; S; S; NS; NS; S/NS stn. depth	NS	NS	NS	2:all	
51-0001	ZOOPLANKTON										
	Trachymedusae Ac	#/50 NNV	#/haul	17	218	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
		identification	species	17	218	S; S; S/NS; NS; NS	NS	NS	author cited	2:all	
		length, gonads	mm	NS	NS	NS	NS	NS	NS	2:all	Remark 3; applies also to 50-0001
		width, overall	mm	NS	NS	NS	NS	NS	NS	2:all	Remark 3; applies also to 50-0001
		height, overall	mm	NS	NS	NS	NS	NS	NS	2:all	Remark 3; applies also to 50-0001
		shape, overall	NA	NS	NS	NS	NS	NS	NS	2:all	Remark 3; applies also to 50-0001
	Narcomedusae Al	#/50 NNV	#/haul	2	4	S; S; S; NS; NS	NS	METHOD 2	#	2:all	
		identification	species	2	4	S; S; S; NS; NS	NS	NS	author cited	2:all	

METHOD 1: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor and standardized to 100-0m vertical hauls.

METHOD 2: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0001 (cont'd)		width, overall	mm	NS	NS	NS	NS	NS	range	2:all	Remark 3; applies also to 50-0001
	Thecosomata	#/50 NNV	#/haul	19	9812	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
	Sh	identification	species	19	9812	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
	Pelecypoda	#/50 NNV	#/haul	11	280+	S; S; S/NS; NS; NS	NS	METHOD 2	#, present	2:all	
	un.	identification	class	11	280+	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
		age	larva	11	280+	S; S; S/NS; NS; NS	NS	NS	stage	2:all	
	Annelida	#/50 NNV	#/haul	2	relative	S; S; S; NS; NS	NS	METHOD 2	present	2:all	
	un.	identification	phylum	2	relative	S; S; S; NS; NS	NS	NS	present; NS	2:all	
		age	larva	2	relative	S; S; S; NS; NS	NS	NS	stage; present	2:all	
	Ostracoda	#/50 NNV	#/haul	8	80+	S; S; S; NS; NS	NS	METHOD 2	#, present	2:all	
	un.	identification	subclass	8	80+	S; S; S; NS; NS	NS	NS	NS	2:all	
	Copepoda	#/50 NNV	#/haul	19	15991	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
	un.	identification	subclass	19	15991	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
		age	nauplius	19	15991	S; S; S/NS; NS; NS	NS	NS	stage	2:all	
	Calanoida	#/50 NNV	#/haul	19	49171+	S; S; S/NS; NS; NS	NS	METHOD 2	#, present	2:all	
	Al,C,Cf,Ch,Co,	identification	genus, species	19	49171+	S; S; S/NS; NS; NS	NS	NS	author cited	2:all	
	Eh,En,Gb,Gt,	age	relative	19	6276	S; S; S/NS; NS; NS	NS	NS	juvenile	2:all	
	Hn,Ml,Mpy,Pm,										
	Sm,Smi										
	Cyclopoida	#/50 NNV	#/haul	19	99160	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
	O,Ol	identification	genus	19	99160	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
	Cirripedia	#/50 NNV	#/haul	9	1001	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
	un.	identification	subclass	9	1001	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
		age	cyprid	9	1001	S; S; S/NS; NS; NS	NS	NS	stage	2:all	
	Amphipoda	#/50 NNV	#/haul	8	64	S; S; S/NS; NS; NS	NS	METHOD 2	#	2:all	
	un.	identification	order	8	64	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
	Euphausiacea	#/50 NNV	#/haul	1	2	S; S; S; NS; NS	NS	METHOD 2	#	2:all	
	un.	identification	order	1	2	S; S; S; NS; NS	NS	NS	NS	2:all	
		age	furcilia	1	2	S; S; S; NS; NS	NS	NS	stage	2:all	
	Echinoidea	#/50 NNV	#/haul	1	relative	S; S; S; NS; NS	NS	METHOD 2	present	2:all	
	un.	identification	class	1	relative	S; S; S; NS; NS	NS	NS	present; NS	2:all	
		age	echinopluteus	1	relative	S; S; S; NS; NS	NS	NS	stage; present	2:all	

METHOD 2: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0001 (cont'd)	Ophiuroidea un.	#/50 NNV	#/haul	5	1+	S; S; S; NS; NS	NS	METHOD 2	#, present	2:all	
		identification	subclass	5	1+	S; S; S; NS; NS	NS	NS	NS	2:all	
		age	ophiopluteus	5	1+	S; S; S; NS; NS	NS	NS	stage	2:all	
	Copelata un.	#/50 NNV	#/haul	17	9724	S; S; S/NS; NS; NS	NS	METHOD 2	#		
		identification	order	17	9724	S; S; S/NS; NS; NS	NS	NS	NS	2:all	
51-0002	ZOOPLANKTON										
	Anthomedusae Eb,Ef,He,S	#/NS PNH	#/m	1	36	NS; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/79-30 PNH	#/m	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	genus, species	2	37	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Leptomedusae Im	#/79-30 PNH	#/m	1	4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	species	1	4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae A,Ad	#/NS CBH	#/m	1	5+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#, present	2:1,2,4,5,8	
		#/NS PNH	#/m; #/m	2; 2	7; 31	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/NS PNV	#/m; #/m	1; 2	2; 29	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/79-30 PNH	#/m	1	30	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/79-30 PNV	#/m	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	few	2:1,4,5,8	
		#/282-30 PNH	#/m	1	220	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	

METHOD 2: less abundant organisms - complete enumeration; abundant forms - count on an aliquot part of the sample multiplied by the appropriate factor.

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		#/282-30 PNV	#/s; #/ss	1; 1	relative; 5	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,4,5,8	
		identification	genus, species	11	329+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	4	257	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	Narcomedusae A,Al	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNH	#/s; #/ss	1; 1	6; 10	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/NS PNV	#/s; #/ss	1; 1	1; 1	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/79-30 PNH	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNH	#/ss	1	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	genus, species	5	20+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Siphonophora Da	#/NS PNV	#/ss	2	6	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	7	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cydippida Mc	#/NS PNH	#/s	1	9	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		identification	species	1	9	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Beroida Bc	#/NS PNH	#/s; #/ss	1; 1	1; 12	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/NS PNV	#/ss	1	1	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	species	2	14	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Thecosomata S,Sh	#/NS PNH	#/s; #/ss	1; 1	relative; 34	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,2,4,5,8	
		#/NS PNV	#/ss	2	57	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/s; #/ss	1; 1	1; 17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	genus, species	7	109+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gymnosomata C,Cl	#/NS PNH	#/s; #/ss	2; 2	relative; 79	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,2,4,5,8	
		#/79-30 PNV	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/282-30 PNH	#/ss	1	43	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	genus, species	5	123+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Polychaeta un	#/79-30 PNH	#/ss	1	5	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	2	6	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	6	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Diplostraca P,Pl	#/NS CBH	#/s	1	relative	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/79-30 PNH	#/ss	1	3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	3	5+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Myodocopa C,Cb	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNV	#/ss	1	2	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	4	3+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		sex	NA	1	2	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
	Copepoda un.	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		#/NS PNH	#/ss	1	3	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/NS PNV	#/ss	2	359	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/79-30 PNH	#/ss	1	104	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNH	#/ss	1	116	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	3	124	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	9	706+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	9	706+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Al,Cg,Ch, Db,Dt,Eg,Gt, Hn,Lm,Ml,Mpy, Pm	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNH	#/s; #/ss	3; 2	relative; 987	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,2,4,5,8	
		#/NS PNV	#/ss	2	727	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/79-30 PNH	#s; #/ss	1; 1	relative; 793	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	many; #	2:1,2,4,5,8	
		#/79-30 PNV	#/s	3	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,4,5,8	
		#/282-30 PNH	#/ss	1	21	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/s; #/ss	1; 3	relative; 482	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		identification	species	17	3010+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	17	3010	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	17	421	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
	Harpacticoida E	#/282-30 PNV	#/ss	1	4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	1	4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cyclopoida CY,Ob,Om,un	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNH	#/s	2	relative	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNV	#/ss	2	401	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/79-30 PNH	#/s; #/ss	1; 1	relative; 39	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,2,4,5,8	
		#/79-30 PNV	#/s	2	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,4,5,8	
		#/282-30 PNH	#/ss	1	37	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/s; #/ss	1; 3	relative; 282	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present; #	2:1,4,5,8	
		identification	order, genus, species	14	759+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	4	758	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		sex	NA	4	383	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
	Thoracica B	#/NS CBH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/NS PNV	#/ss	1	3	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	3	4+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius, larva, cyprid	3	4+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Isopoda un.	#/NS PNV	#/ss	1	1	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	order	1	1	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	NS	2:4,5,8	
	Amphipoda Hg,Hm,Pab,Pl	#/NS CBH	#/s	1	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		#/NS PNH	#/s; #/ss	2; 2	7; 103	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/NS PNV	#/s; #/ss	1; 2	1; 5	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/79-30 PNH	#/ss	1	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/s; #/ss	1; 1	1; 1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	species	8	122	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)		age	relative	1	2	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	young	2:4,5,8	
	Decapoda P,Sa	#/NS PNH	#/ss	1	24	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	2	25	S/NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	late larva	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Insecta un.	#/NS PNV	#/ss	1	1	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	class	1	1	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Chaetognatha Eh,Sa	#/NS CBH	#/s	1	11	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,2,4,5,8	
		#/NS PNH	#/s; #/ss	1; 2	5; 88	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#; #	2:1,2,4,5,8	
		#/NS PNV	#/s	1	relative	NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	present	2:1,2,4,5,8	
		#/79-30 PNH	#/ss	1	25	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	4	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	6	133+	S/NS; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
51-0002 (cont'd)	Copelata F,Fb,O,Ov	#/NS CBH	#/m	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	present	2:1,2,4,5,8	
		#/NS PNV	#/ss	2	284	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNH	#/ss	1	69	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/s; #/ss	1; 2	1; 103	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	genus, species	7	457+	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	73	NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/NS PNH	#/ss	1	4	NS; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/282-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		age	larva	2	5	S/NS; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
60-0003	ZOOPLANKTON Anthomedusae Eb,Ef,Hc,Sp	#/NS PN	#/s	1	relative	NS; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	several	2:1,2,4,5,8	
		#/NS-100 STNH	#/s	1	12+	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#, several	2:1,2,4,5,8	
		identification	species	2	12+	S/NS; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		height, overall	mm	2	2+	S/NS; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
60-0003 (cont'd)	Leptomedusae P1	#/NS PN	#/s	1	1	NS; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5, 8	
		#/NS-100 STNH	#/s	1	1	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5, 8	
		identification	#/s	2	2	S/NS; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		width, overall	mm	2	2	S/NS; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Trachymedusae A	#/NS-100 STNH	#/s	1	relative	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	many	2:1,2,4,5, 8	
		identification	genus	1	relative	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	many; ABS taxonomic manual used	2:4,5,8	
	Thecosomata S	#/NS-100 STNH	#/s	1	10	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5, 8	
		identification	genus	1	10	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Calanoida Cg,Ch,Eg,Lm,Pm	#/NS-100 STNH	#/s	1	2+	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#, present, most abundant	2:1,2,4,5, 8	
		#/571-50 PNH	#/s	1	relative	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	few, very many	2:1,2,4,5, 8	
		identification	species	2	2+	S; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	2	relative	S; S; WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage #; present, few, very many, most abundant	2:4,5,8	
		sex	NA	2	relative	S; S, WA/NS; NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present, few, very many, most abundant	2:4,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
60-0003 (cont'd)	Mysidacea Mr?	#/571-50 PNH	#/s	1	5	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	species	1	5	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	adult; relative	1	2	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage; young, adolescent	2:4,5,8	
		sex	NA	1	2	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,8	
	Amphipoda Hm,Hym,Pl	#/NS-100 STNH	#/s	1	9	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	species	1	9	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	6	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	young	2:4,5,8	
		sex	NA	1	2	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,8	
	Euphausiacea Tr	#/NS-100 STNH	#/s	1	relative	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	several	2:1,2,4,5,8	
		identification	species	1	relative	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	several; ABS taxonomic manual used	2:4,5,8	
		age	relative	1	relative	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	young; several	2:4,5,8	
	Decapoda un.	#/NS-100 STNH	#/s	1	7	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	order	1	7	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	7	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
60-0003 (cont'd)	Chaetognatha un.	#/NS-100 STNH	#/a	1	3	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	phylum	1	3	S; NS; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
60-0067	ZOOPLANKTON Copepoda un.	#/subsampler	total # in 7 series	1	<30	METHOD 7	nets and contents frozen in toto	METHOD 8	#	2:1,2,4-6,8	<30 specimens were present in Series IX, only one of the seven series to which the data apply
		identification	order	1	<30	METHOD 7	nets and contents frozen in toto	METHOD 9	NS	2:4-6,8	
	Calanoida Cg,Ch,Co,Eq, Ml,Sm	#/subsampler	total # in 1 series	1	85	METHOD 7	nets and contents frozen in toto	METHOD 8	#	2:1,2,4-6,8	
		identification	species	1	85	METHOD 7	nets and contents frozen in toto	METHOD 9	author cited	2:4-6,8	
		length, overall	mm	1	29	METHOD 7	nets and contents frozen in toto	METHOD 9	NS	2:4-6,8	
		age	copepodite #; adult; relative	NS	NS	METHOD 7	nets and contents frozen in toto	METHOD 9	stage; stage; #; immature	2:4-6,8	Remark 1
		sex	NA	NS	NS	METHOD 7	nets and contents frozen in toto	METHOD 9	NS	2:4,6,8	Remark 1
61-0001	ZOOPLANKTON Trachymedusae Ad	#/73-30 PNV	#/a	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 7: S; rel.; NS; NS; no; loc./date rel.; carousel timed to move at hourly intervals; one net placed in every other receptacle; 12 collections made over 24h period after which sampler turned off automatically.

METHOD 8: samples thawed in 5-10% F, lower end of net cut off and organisms removed with aid of fine jet of water and forceps, sorted into major groups represented and counted.

METHOD 9: samples thawed in 5-10% F, lower end of net cut off and organisms removed with aid of fine jet of water and forceps, sorted into major groups represented and identified.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
61-0001 (cont'd)	Siphonophora Dm	#/73-30 PNV	#/s	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Thecosomata Sh	#/73-30 PNV	#/ss	1	2	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	2	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Polychaeta un.	#/73-30 PNH	#/ss	1	10	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/73-30 PNV	#/ss	8	181	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	9	191	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	9	191	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Acarina un.	#/73-30 PNV	#/ss	2	4	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order	2	4	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Diplostraca En,Pl	#/73-30 PNV	#/ss	4	18	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	4	18	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
61-0001 (cont'd)	Copepoda un.	#/73-30 PNH	#/ss	1	1325	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/73-30 PNV	#/ss	9	2241	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	10	3566	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	10	3566	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Calanoida A,Ac,Cg,Ch,Db, Dt,Eh,Lm,Ml, Mpy,Pm	#/73-30 PNH	#/ss	1	98	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/73-30 PNV	#/ss	9	3397	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	10	3495	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	10	3495	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	10	1370	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	NS	2:4,8	
	Harpacticoida Df,E,L,un.	#/73-30 PNH	#/ss	1	73	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/73-30 PNV	#/ss	6	112	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, genus, species	7	185	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cyclopoida C,Ce,CY,Ob,Oe	#/73-30 PNH	#/ss	1	1	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/73-30 PNV	#/ss	7	73	S; S, WA; NS; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
61-0001 (cont'd)		identification	genus, species	8	74	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	5	66	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	4	31	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
	Thoracica B	#/73-30 PNV	#/ss	4	97	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	4	97	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	4?	97?	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	Age is stated for 1 station only but assume other *Balanus* must also be planktonic stages
	Decapoda un.	#/73-30 PNV	#/ss	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Chaetognatha Se	#/73-30 PNV	#/ss	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Copelata Fb	#/73-30 PNV	#/ss	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001	ZOOPLANKTON Anthomedusae Hc	#/233-50 PNV	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	present	2:1,4,5,8	
		identification	species	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	present; ABS taxonomic manual used	2:4,5,8	
	Leptomedusae O	#/233-50 PNV	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	several	2:1,4,5,8	
		identification	genus	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	several; ABS taxonomic manual used	2:4,5,8	
		width, overall	mm	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	several; NS	2:4,5,8	
	Trachymedusae Ad	#/233-50 PNH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	very many	2:1,2,4,5,8	
		#/233-50 PNV	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	few	2:1,4,5,8	
		identification	species	2	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	few, very many; ABS taxonomic manual used	2:4,5,8	
	Siphonophora Da	#/233-50 PNH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	present	2:1,2,4,5,8	
		#/233-50 PNV	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	1+	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		height, overall	mm	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	NS	2:4,5,8	
	Polychaete un.	#/73-30 PNV	#/ss	1	4	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001 (cont'd)		#/233-50 PNV	#/ss	5	11	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 10	#	2:1,4,5,8	
		identification	class	6	15	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	6	15	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Acarina un.	#/233-50 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Diplostraca E,En,P,Pl	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/s; #/ss	1; 5	relative; 82	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 10	present; #	2:1,4,5,8	
		identification	genus, species	5	83+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/ss	2	319	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/ss	5	21	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 10	#	2:1,4,5,8	
		identification	subclass	7	340	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	7	340	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 10: medusae sorted out in 3 stations; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001 (cont'd)	Calanoida Ac,Cg,Ch,Ob, Dt,Eg,Eh,Lm, Ml,Pm	#/73-30 PNV	#/ss	2	162	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNH	#/s	2	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5, 8	
		#/233-50 PNV	#/s; #/ss	2; 5	relative; 2898	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 10	present, very few, few, many; #	2:1,4,5,8	
		identification	species	10	3060+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	7	3060+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	7	1528+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
		egg number	presence	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	present, very few	2:1,4,5,8	
	Harpacticoida Ds,E,Tf	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/ss	2	3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 11	#	2:1,4,5,8	
		identification	genus, species	3	4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
		sex	NA	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 10: medusae sorted out in 3 stations; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 11: medusae sorted out; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DAT SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001 (cont'd)	Cyclopoida CY,Ob,Om	#/73-30 PNV	#/ss	2	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/s; #/ss	2; 3	relative; 4	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 12	present, very few; #	2:1,4,5,8	
		identification	genus, species	6	6+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	4	5	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	2	3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,8	
	Cirripedia un.	#/233-50 PNH	#/s	2	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present, few	2:1,2,4,5,8	
		identification	subclass	2	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	present, few; ABS taxonomic manual used	2:4,5,8	
		age	cyprid	2	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage; present, few	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/ss	3	9	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 13	#	2:1,4,5,8	
		identification	genus	4	10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	4	10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 12: medusae sorted out in 2 stations; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 13: medusae sorted out in 4 stations; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001 (cont'd)	Amphipoda Gw,Hm,Pg,Pi	#/233-50 PNH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	present	2:1,2,4,5,8	
		#/233-50 PNV	#/s; #/ss	1; 1	39; 8	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 11	#; #	2:1,4,5,8	
		identification	species	3	47	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Decapoda un.	#/233-50 PNH	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	order	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Insecta un.	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-50 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 11	#	2:1,4,5,8	
		identification	class	2	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Chaetognatha Se	#/233-50 PNH	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	several	2:1,2,4,5,8	
		#/233-50 PNV	#/s; #/ss	2; 2	relative; 3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 12	present, many; #	2:1,4,5,8	
		identification	species	4	3+	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 11: medusae sorted out; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 12: medusae sorted out in 2 stations; ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
62-0001 (cont'd)		size, overall	relative	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	large; several	2:4,5,8	
	Copelata O	#/233-50 PNV	#/s	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	few	2:1,4,5,8	
		identification	genus	1	relative	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	few; ABS taxonomic manual used	2:4,5,8	
	ICHTHYOPLANKTON										
	Other un.	#/233-50 PNH	#/s	1	2	S; S, WA; S; 1; no; loc. rel.	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		age	larva	1	2	S; S, WA; S; 1; no; loc. rel.	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
69-0066	PHYTOPLANKTON										
	Centrales Ca,Ccm,Ccn, Cct,Cd,Cdb, Cf,CH,Ca,Cai, Cau,R,Rh,Rhh, T,Tn	#/NS PNH?	NS	2	relative	NS; S; NS; NS; NS; date rel.	METHOD 14	NS	present	2:1,2,4,5,8	
		#/NS PNV	NS	3	relative	NS; S?/NS; NS; NS; NS; date rel.	METHOD 14	NS	present	2:1,2,4,5,8	
		identification	genus, species	4	relative	NS; S/S?/NS; NS; NS; NS; date rel.	METHOD 14	NS	present; NS	2:4,5,8	
	Pennales FR,N,Ncl,Nme, NT,Ntr,Tn	#/NS PNH?	NS	3	relative	NS; S; NS; NS; NS	METHOD 14	NS	present	2:1,2,4,5,8	
		#/NS PNV	NS	3	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present	2:1,2,4,5,8	
		identification	genus, species	4	relative	NS; S/S?/NS; NS; NS; NS	METHOD 14	NS	present; NS	2:4,5,8	
	Chrysophyceae Db,Dmp,Et	#/NS PNH?	NS	2	relative	NS; S; NS; NS; NS	METHOD 14	NS	present	2:1,2,4,5,8	
		#/NS PNV	NS	3	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present	2:1,2,4,5,8	
		identification	species	3	relative	NS; S/S?/NS; NS; NS; NS	METHOD 14	NS	present; NS	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 14: preserved in gluteraldehyde in Whirl-pak bags.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
69-0066 (cont'd)	Dinophyceae Gc,P,Pb,Pbe, Pm,Pp,un.	#/NS PNH?	NS	3	relative	NS, S; NS; NS; NS; date rel.	METHOD 14	NS	present	2:1,2,4,5,8	
		#/NS PNV	NS	2	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	present	2:1,2,4,5,8	
		identification	order, genus, species	4	relative	NS; S/S?; NS; NS; NS	METHOD 14	NS	present; NS	2:4,5,8	
	ZOOPLANKTON										
	Foraminiferida un.	#/NS PNV	NS	2	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present	1:3	
		identification	order	2	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present; NS	1:3	
	Radiolaria un.	#/NS PNV	NS	1	relative	NS; NS; NS; NS; NS; date rel.	METHOD 14	NS	present	1:3	
		identification	subclass	1	relative	NS; NS; NS; NS; NS; date rel.	METHOD 14	NS	present; NS	1:3	
	Tintinnida un.	#/NS PNH?	NS	1	relative	NS, S; NS; NS; NS	METHOD 14	NS	present	1:3	
		#/NS PNV	NS	3	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present	1:3	
		identification	order	3	relative	NS; S/S?/NS; NS; NS; NS	METHOD 14	NS	present; NS	1:3	
	Rotifera un.	#/NS PNV	NS	2	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	present	1:3	
		identification	phylum	2	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	present; NS	1:3	
	Polychaeta un.	#/NS PNV	NS	1	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	present	1:3	
		identification	class	1	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	present; NS	1:3	
		age	larva	1	relative	NS; S?; NS; NS; NS; date rel.	METHOD 14	NS	stage; present; NS	1:3	
	Copepoda un.	#/NS PNH?	NS	1	relative	NS, S; NS; NS; NS	METHOD 14	NS	present	1:3	
		#/NS PNV	NS	3	relative	NS; S?/NS; NS; NS; NS	METHOD 14	NS	present	1:3	
		identification	subclass	4	relative	NS; S/S?/NS; NS; NS; NS	METHOD 14	NS	present; NS	1:3	

METHOD 14: preserved in gluteraldehyde in Whirl-pak bags.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
70-0002	ZOOPLANKTON Foraminiferida Gp	#/202-S PNH	#/haul	45	1613	METHOD 15	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		#/202-S PNV	#/haul	77	44426	S; S, relative, estimated angle; S/NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		# tests/202-S PNV	$\#/m^3$	77	1263.98	S; S, relative, estimated angle; S/NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		identification	species	81	46039+	METHOD 16	10% F buffered with hexamethyl tetramine	METHOD 17	detailed taxonomic review given	2:4,5,8	
		biomass, total dry	g/m^3	86	155	METHOD 16	10% F buffered with hexamethyl tetramine	METHOD 18	NS	2:1,2,4,5,8	
		width, maximum	u	5	1112	METHOD 137	10% F buffered with hexamethyl tetramine	METHOD 19	METHOD 20	2:5,8	
		width, minimum	u	2	6	S; S, relative, estimated angle; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	METHOD 19	NS	2:5,8	
		width, external pore	u	1	2	NS; NS; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	measured on cathode ray tube of JOEL-2 SEM @ x4000 magnif.	mean diam.; coeff. var.	2:5,8	
		width, internal pore	u	1	2	NS; NS; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	measured on cathode ray tube of JOEL-2 SEM @ x4000 magnif.	mean diam.; coeff. var.	2:5,8	
		width, proloculus	u	1	24	NS; NS; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	METHOD 21	mean diam.; S.D.; coeff. var.	2:5,8	
		number, external pores	#/25u surface	1	2	NS; NS; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	counted on JOEL-2 SEM @ x4000 magnif.	#	2:5,8	
		number, internal pores	#/25u surface	1	2	NS; NS; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	counted on JOEL-2 SEM @ x4000 magnif.	#	2:5,8	

METHOD 15: S; S, relative, estimated angle; S/NS; 1; TSK meter; net floated at ship's bow near bow-thruster.
METHOD 16: S; S, relative, estimated angle; S/NS; 1; TSK meter; surface net floated at ship's bow near bow-thruster.
METHOD 17: dried in 60°C oven for >24h; ashed with TRACERLAB low-temp. asher (ref. for method description given); examined with Wilde M5 microscope.
METHOD 18: NS as taken by Folsom Splitter; filtered onto Millipore filter; dried in 60°C oven for >24h; weighed, ashed with TRACERLAB low-temp. asher (ref. for method description given); reweighed using electronic balance.
METHOD 19: mounted on slide; measured @ x50 magnif. using Wilde M5 microscope with eyepiece graticule.
METHOD 20: max. diam.; recorded to nearest 20u; noted that frequencies of size ranges <200u may not be statistically reliable; variance; coeff. var.; student t-test.
METHOD 21: specimen mounted on glass side; coated with Al using Edwards High vacuum evaporator (ref. for method description given); x500 magnif.
METHOD 137: S/NS; S, relative, estimated angle/NS; S/NS; 1; TSK meter; surface net floated at ship's bow near bowthruster.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
70-0002 (cont'd)		appearance, surface	NA	2	6	S; S, relative, estimated angle; NS; 1; TSK meter	10% F buffered with hexamethyl tetramine	METHOD 22	NA	4	
		coiling	% dextral	66	%	METHOD 16	10% F buffered with hexamethyl tetramine	METHOD 23	mean %; coeff. var.	2:4,5,8	
		age	normal form, quadrate, kummerform	5	3871	METHOD 16	10% F buffered with hexamethyl tetramine	METHOD 23	NS	2:4,5,8	
70-0067	ZOOPLANKTON Anthomedusae Eb,Ef,Hc	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	species	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Rotifera A,B,K,KE	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	genus	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; NS	2:all	
	Polychaeta un.	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	2	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	class	2	relative	S; NS; NS; 3, NS; NS	NS	NS	present; NS	2:all	
		age	larva	2	relative	S; NS; NS; 3, NS; NS	NS	NS	stage; present	2:all	
	Diplostraca (Conchostraca), B1,D1,Lk	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>12?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	suborder, species	>12?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
		age	relative	>12?	relative	S; NS; NS; 3, NS; NS	NS	NS	immature; present	2:all	

METHOD 16: S; S, relative, estimated angle; S/NS; 1; TSK meter; surface net floated at ship's bow near bow-thruster.

METHOD 22: previously mounted specimens cleaned in Sparkleen detergent; rinsed x2 in dist. H_2O buffered with $NaCO_3$; exposed to atmos. of atomic O_2 for ≥ 8h in TRACERLAB low-temp. asher; photomicrographs taken with JOEL-2 SEM using a Polaroid camera; enlargements made @ x3000 magnif.

METHOD 23: tests removed with 000 brush; mounted on micro-paleontological slide precoated with gum tragacanth.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
70-0067 (cont'd)	Calanoida Ab,Al,Cf,Cg, Ch,Da,Do,Dp, Da,Ec,Egr,Eh, El,Ene,Er,Eri, He,Lm,P	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	30	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	genus, species	30	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Harpacticoida Da,Pm	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	species	>3?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Cyclopoida Cac,Ct,Cv,CY, Oa	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>12?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	genus, species	>12?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Mysidacea Ml,Mr	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>10?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	species	>10?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Isopoda Me	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	6	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	species	6	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
	Amphipoda GA,Gl,H,MO,P	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	>4?	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	genus, species	>4?	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	
		age	relative	>4?	relative	S; NS; NS; 3, NS; NS	NS	NS	immature; present	2:all	
	Chaetognatha Eh	#/202-10 MNH or 726-66 PN	# of occurrences at 30 stns.	1	relative	S; NS; NS; 3, NS; NS	NS	NS	present	2:all	
		identification	species	1	relative	S; NS; NS; 3, NS; NS	NS	NS	present; author cited	2:all	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0001	ZOOPLANKTON Anthomedusae Hc, Sp	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	3	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	3	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/73-30 PNV	#/s	2	17	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	3	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	3	31	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Narcomedusae Al	#/73-30 PNV	#/s	2	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/s	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	2	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0001 (cont'd)	Polychaeta un.	#/73-30 PNV	#/m; #/mm	3; 1	55; 8	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		#/233-30 PNV	#/m	4	26	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	4	89	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	4	89	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Diplostraca Pl	#/233-30 PNV	#/m	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/mm	4	1673	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/mm	4	58	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	4	1731	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	early nauplius, late nauplius, nauplius	4	1731	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Al,Cg,Ch, Dt,Eh,Lm,Ml, Mpy,Pm	#/73-30 PNV	#/m; #/mm	4; 4	905; 1551	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		#/233-30 PNV	#/m	4	2179	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	4	4635	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	ABS taxonomic manual used	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0001 (cont'd)		age	copepodite #	4	4635	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	4	1118	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida En,H,un.	#/73-30 PNV	#/m^3	2	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/m	1	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order, genus, species	3	11	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	young	2:4,5,8	
		sex	NA	1	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida Ob,Om	#/73-30 PNV	#/m^3	4	807	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/m	4	846	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	4	1653	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	4	1653	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	4	792	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Mysidacea un.	#/233-30 PNV	#/m	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0001 (cont'd)		identification	order	1	1	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Cumacea D	#/233-30 PNV	#/m	1	2	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	2	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Cirripedia un.	#/73-30 PNV	#/m	4	301	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/m	3	220	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	subclass	4	525	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	nauplius, larva, cyprid	4	525	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Amphipoda MO,un.	#/73-30 PNV	#/m	2	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/m	3	5	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order, genus	3	9	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Chaetognatha Se,un.	#/73-30 PNV	#/m	3	19	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/m	3	5	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum, species	3	34	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0001 (cont'd)	Copelata Fb,0	#/73-30 PNV	#/s; #/ss	3; 1	34; 4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		#/233-30 PNV	#/s	3	28	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	3	66	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/233-30 PNV	#/s	2	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		age	larva	2	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
71-0004	ZOOPLANKTON Anthomedusae Hc	#/73-30 PNV	#/s	2	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	3	7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Polychaeta un.	#/73-30 PNV	#/ss	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Diplostraca En,Pl	#/73-30 PNV	#/ss	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0004 (cont'd)		#/233-30 PNV	#/ss	2	41	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	2	45	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		sex	NA	1	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#s; #/ss	1; 1	11; 566	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	2	577	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	577	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Db,Dt,Eh, Lm,Pm	#/73-30 PNV	#/s; #/ss	2; 1	554; 2906	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/s; #/ss	1; 2	16; 2742	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	6218	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	3	6202	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	3	4413	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida A,D,En,L,Ze, un.	#/73-30 PNV	#/s; #/ss	1; 1	4; 30	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, genus, species	2	35	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
71-0004 (cont'd)		age	relative	2	27	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature, young	2:4,5,8	
		sex	NA	2	3	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida Ca, CY	#/73-30 PNV	#/aa	1	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	1	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cirripedia un.	#/233-30 PNV	#/aa	2	5	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	2	5	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	2	5	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
72-0001	ZOOPLANKTON										
	Foraminiferida Gp	#/75-C PNV	#/0.5a	6	17	S; S, relative, estimated; S; 1; TSK meter; NS atn depth	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		#/202-C PNV	#/0.5a	5	8	S; S, relative, estimated; S; 1; TSK meter; NS atn. depth	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		#/202-S PNV	#/0.5a	7	16	S; S, relative, estimated; S; 1; TSK meter; NS atn. depth	10% F buffered with hexamethyl tetramine	METHOD 17	#	2:1,2,4,5,8	
		identification	species	15	41	S; S, relative, estimated; S; 1; TSK meter; NS atn	10% F buffered with hexamethyl tetramine	METHOD 17	detailed taxonomic review given	2:4,5,8	

METHOD 3: aa taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: aa taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 17: dried in 60°C oven for $\geq$24h; ashed with TRACERLAB low-temp. asher (ref. for method description given); examined with Wilde M5 microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0001 (cont'd)		biomass, total dry	g/m^3	35	30	S; S, relative, estimated; S; 1; TSK meter; NS stn depth	10% F buffered with hexamethyl tetramine	METHOD 18	NS	2:1,2,4,5,8	
		width, maximum	u	NS	22	NS; S, relative, estimated angle; NS; 1; TSK meter; NS stn. depth	10% F buffered with hexamethyl tetramine	METHOD 19	METHOD 24	2:5,8	
		coiling	% dextral, sinistral	9	%	S; S, relative, estimated; S; 1; TSK meter; NS stn. depth	10% F buffered with hexamethyl tetramine	METHOD 23	NS	2:4,5,8	
		age	normal form, quadrate	NS	22	NS; S, relative, estimated angle; NS; 1; TSK meter; NS stn. depth	10% F buffered with hexamethyl tetramine	METHOD 23	NS	2:4,5,8	
72-0004	ZOOPLANKTON Anthomedusae H,Hc	#/73-30 PNV	#/m	9	30	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	genus, species	9	30	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Rotifera un.	#/73-30 PNV	#/m	4	21	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	4	21	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/m	1	3	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	1	3	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 18: NS as taken by Folsom Splitter; filtered onto Millipore filter; dried in 60°C oven for $\geq$24h; weighed, ashed with TRACERLAB low-temp. asher (ref. for method description given); reweighed using electronic balance.

METHOD 19: mounted on slide; measured @ x50 magnif. using Wilde M5 microscope with eyepiece graticule.

METHOD 23: tests removed with 000 brush; mounted on micro-paleontological slide precoated with gum tragacanth.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)	Mollusca un.	weight, wet	mg/m^3	1	NS	NS; NS; NS; NS; NS	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	determined by directly weighing preserved specimens	NS	2:1,2,4,5,8	
	Gastropoda un.	#/73-30 PNV	#/ss	4	18	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	4	18	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	4	18	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Pelecypoda un.	#/73-30 PNV	#/ss	4	2635	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	4	2635	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	4	2635	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Polychaeta un.	#/73-30 PNO	#/ss	1	8	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/73-30 PNV	#/ss	15	1269	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	16	1277	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		weight, wet	mg/m^3	1	NS	NS; NS; NS; NS; NS	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	determined by directly weighing preserved specimens	NS	2:1,2,4,5,8	
		age	larva; relative	16	1277	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage; young	2:4,5,8	
	Diplostraca En,Pl	#/73-30 PNO	#/ss	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)		#/73-30 PNV	#/ss	13	158	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	species	14	162	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult	2	5	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
		sex	NA	4	91	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Copepoda un.	#/73-30 PNO	#/ss	1	283	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/73-30 PNV	#/ss	17	11222	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	subclass	18	11505	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		weight, wet	mg/m^3	1	NS	NS; NS; NS; NS; NS	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	determined by directly weighing preserved specimens	NS	2:1,2,4,5,8	
		age	nauplius, early, late	18	11505	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Db,Dt,Eh, Lm,Pm	#/73-30 PNO	#/ss	1	563	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/73-30 PNV	#/s; #/ss	3; 17	939; 15697	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	species	18	17199	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	18	17199	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	18	5854	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)	Harpacticoida A,D,Da,E,En,P, Tf,Z,Za,un.	#/73-30 PNV	#/m; #/m³	1; 8	5; 78	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, genus, species	8	83	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; adult; relative	6	69	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; stage; immature, young	2:4,5,8	
		sex	NA	2	11	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida Ca,CY,un.	#/73-30 PNV	#/m³	12	55	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	order, genus, species	12	55	S; S, WA; S/NS; 1; no; S/NS loc.; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; adult; relative	9	44	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; stage; immature	2:4,5,8	
		sex	NA	4	14	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cirripedia un.	#/73-30 PNO	#/m³	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/73-30 PNV	#/m³	3	406	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	4	407	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		weight, wet	mg/m^3	1	NS	NS; NS; NS; NS; NS	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	determined by directly weighing preserved specimens	NS	2:1,2,4,5,8	
		age	nauplius, cyprid	4	407	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)	Thoracica B	#/73-30 PNV	#/ss	7	287	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	7	287	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius, larva, cyprid	7	287	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Cumacea D	#/73-30 PNV	#/ss	1	1	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	1	1	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Amphipoda Dp	#/73-30 PNV	#/s	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	immature	2:4,5,8	
	Other un.	#/73-30 PNV	#/ss	2	323	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	#	2:1,4,5,8	
		age	egg	2	323	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	stage	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/73-30 PNV	#/s	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		age	larva	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)	EPONTIC ALGAE Centrales Cb,CH,Cs,R,T, Ts,Tbi,Tn,un.	#/S core	# cells/L	3	2483943	METHOD 25; S/NS time	METHOD 26	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	3	2483943	preserved sections shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,Ace,Ad,AH, Akk,Akr,Aks, All,Alm,Am,AP, At,Ck,CO,Dls, Dln,DP,G,Gsp, Gf,Ld,N,Nc, Ncl,Ncr,Ncy, Nde,Ndr,Nf, Nfr,Ngd,Nge, Nhy,Nk,Nkj, Nls,Nle,Nli, Nn,No,Npe,Npo, Nqu,Nre,Nsa, Nsb,Nso,Nsp, Nsr,Nss,NT, Ntd,Ntg,Nti, Ntr,Nvm,P,Pa, Pan,Pc,PL,Pmi, Pq,Pqb,Pqc, Pqd,Pql,Pqm, Pqs,Psb,Pse, Psm,Psu,Si,St, Tm,un.	#/S core	# cells/L	3	41063062	METHOD 25	METHOD 26	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	3	41063062	preserved sections shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chlorophyceae un.	#/S core	# cells/L	2	217572+	METHOD 25	METHOD 26	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order	2	217572+	METHOD 25	METHOD 26	METHOD 29	taxonomic references stated	2:5,8	

METHOD 25: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw; NS stn. depth.

METHOD 26: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 27: each section agitated; each 1mL ss of bottom section and 10mL ss of other sections pipetted or poured into individual 10mL Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 29: each section agitated; each 1mL ss of bottom section and 10mL ss of other sections pipetted or poured into individual 10mL Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0004 (cont'd)	Euglenophyceae E	#/S core	# cells/L	1	36262	METHOD 25	METHOD 26	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8,	
		identification	genus	1	36262	METHOD 25	METHOD 26	METHOD 29	taxonomic references stated	2:5,8	
	Dinophyceae Gc,un.	#/S core	# cells/L	3	3296210	METHOD 25	METHOD 26	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, species	3	3296210	METHOD 25	METHOD 26	METHOD 29	taxonomic references stated	2:5,8	
72-0007	PHYTOPLANKTON Centrales CO,M,R	#/12-20 PNH	#/ss	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	abundant	2:1,2,4-6, 8	
		identification	genus	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	abundant; NS	2:4-6,8	
	Pennales AS,D,F,SY,T	#/12-20 PNH	#/ss	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	abundant	2:1,2,4-6, 8	
		identification	genus	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	abundant; NS	2:4-6,8	
	Chlorophyceae P,SP	#/12-20 PNH	#/ss	2	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	rare	2:1,2,4-6, 8	
		identification	genus	2	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	rare; NS	2:4-6,8	
	Chrysophyceae DN	#/12-20 PNH	#/ss	1	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	rare	2:1,2,4-6, 8	
		identification	genus	1	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1mL ss examined microscopically for each station; x100 magnif.	rare; NS	2:4-6,8	

METHOD 25: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw; NS stn. depth.

METHOD 26: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 27: each section agitated; each 1mL ss of bottom section and 10mL ss of other sections pipetted or poured into individual 10mL Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 29: each section agitated; each 1mL ss of bottom section and 10mL ss of other sections pipetted or poured into individual 10mL Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
72-0007 (cont'd)	Dinophyceae C	#/12-20 PNH	#/ss	2	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare	2:1,2,4-6,8	
		identification	genus	2	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare; NS	2:4-6,8	
	ZOOPLANKTON Ploima K,KE	#/12-20 PNH	#/ss	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	abundant	2:1,2,4-6,8	
		identification	genus	4	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	abundant; NS	2:4-6,8	
	Diplostraca (Cladocera)	#/12-20 PNH	#/ss	1	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare	2:1,2,4-6,8	
		identification	suborder	1	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare; NS	2:4-6,8	
	Copepoda un.	#/12-20 PNH	#/ss	3	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare, common	2:1,2,4-6,8	
		identification	subclass	3	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	rare, common; NS	2:4-6,8	
		age	nauplius	3	relative	S; NS; NS; slow; NS; NS stn. depth; ? date	10% F	3 x 1ml ss examined microscopically for each station; x100 magnif.	stage; rare, common; NS	2:4-6,8	
73-0001	PHYTOPLANKTON Pennales un.	#/73?-12 WNV	extrapolated cells/L	4	126.2	S; S; NS; NS; no; ?stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	order	4	126.2	S; S; NS; NS; no; ?stn. depth; ?date	5% F	3 x 1 ml ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Chlorophyceae un.	#/73?-12 WNV	extrapolated cells/L	3	39.6	S; S; NS; NS; no; ?stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	class	3	39.6	S; S; NS; NS; no; ?stn. depth; ?date	5% F	3 x 1 ml ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	

METHOD 30: 3 x 1ml ss taken; examined with microscope @ x100 magnif.; concentrations calculated from mean no./taxa and H_2O vol. strained.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0001 (cont'd)	ZOOPLANKTON										
	Rotifera un.	#/73?-12 WNV	extrapolated cells/L	3	19.8	S; S; NS; NS; no; ?stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	phylum	3	19.8	S; S; NS; NS; no; ?stn. depth; ?date	5% F	3 x 1 mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Copepoda un.	#/73?-12 WNV	extrapolated cells/L	1	3.9	S; S; NS; NS; no; ?stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	subclass	1	3.9	S; S; NS; NS; no; ?stn. depth; ?date	5% F	3 x 1 mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Other un.	#/73?-12 WNV	extrapolated cells/L	1	4.3	S; S; NS; NS; no; ?stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4,5,8	
		age	nauplius	1	4.3	S; S; NS; NS; no; ?stn. depth; ?date	5% F	3 x 1 mL ss taken; examined with microscope at x100 magnif.	stage	2:4,5,8	
73-0002	BACTERIA										
	un.	#/NBS	log CFU/L	17	53	METHOD 31	METHOD 32	METHOD 157	METHOD 33	2:1,4-6,8	
		identification	relative	17	53	METHOD 31	METHOD 32	METHOD 158	heterotrophs	2:4-6,8	
	PHYTOPLANKTON										
	Centrales CO,CY,M,Mi,P, Pg,ST,T,Tb,Tg, un.	#/VD	# cells/L	5	1.01×10^6	METHOD 159	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	5	1.01×10^6	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:1,2,5,8	

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 30: 3 x 1ml ss taken; examined with microscope @ x100 magnif.; concentrations calculated from mean no./taxa and H_2O vol. strained.

METHOD 31: S; S, WA; S; NS; S/NS stn. depth; collected aseptically prior to other oceanographic sampling.

METHOD 32: kept chilled; processed within 1/2h of collection; spin-plate technique used to dispense 0.1ml ss with cold 0.2mL pipette onto surface of cold agar plate [ZoBell Marine Broth 2216E (Difco) and Plate Count Agar (Difco) prepared previously with deionized water and stored at 5°C until used]; placed in 5°C refrigerator; when ss absorbed by medium, plate inverted; incubated at 5°C; transported in chilled condition.

METHOD 33: quadruplicate spin-plates made of each sample on both media; plates with uneven distribution of colonies discarded; colonies of 3 plates at a replicate set enumerated; #.

METHOD 34: 125ml ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 157: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters; colonies of 3 plates enumerated, averaged and mean value determined.

METHOD 158: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

METHOD 159: S, SL; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)	Pennales A,AH,C,Ca,CO, DP,G,N,Ncl,Ns, Nsu,NT,P,SU,T Tfe,TH,Tn,un.	#/VD identification	# cells/L order, genus, species	5 5	5.68x10^5 5.68x10^5	METHOD 159 preserved 125mL ss shaken; 35mL ss drawn off	METHOD 34 METHOD 28	METHOD 35 Leitz phase-contrast compound microscope @ x500 magnif.	METHOD 36 taxonomic references stated	2:1,5,8 2:1,2,5,8	
	Chlorophyceae S,un.	#/VD identification	# cells/L order, genus	3 3	17000 17000	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 taxonomic references stated	2:1,4,5,8 2:4,5,8	
	Chrysophyceae Db,DN	#/VD identification	# cells/L genus, species	1 1	7000 7000	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 taxonomic references stated	2:1,4,5,8 2:4,5,8	
	Cyanophyceae un.	#/VD identification	# cells/L phylum	4 4	14000 14000	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 taxonomic references stated	2:1,4,5,8 2:4,5,8	
	Euglenophyceae E,un.	#/VD identification	# cells/L phylum, genus	2 2	13x10^4 13x10^4	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 taxonomic references stated	2:1,4,5,8 2:4,5,8	
	Dinophyceae G,Gc,P,un.	#/VD identification	# cells/L phylum, genus, species	2 2	79000 79000	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 taxonomic references stated	2:1,4,5,8 2:4,5,8	
	Other un.	#/VD identification	# cells/L relative	3 3	45000 45000	METHOD 159 METHOD 159	METHOD 34 METHOD 34	METHOD 35 METHOD 37	METHOD 36 pigmented flagellates; taxonomic references stated	2:1,4,5,8 2:4,5,8	

METHOD 28: 35ml ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 10% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30ml dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate mounted in Hyrax on slide.

METHOD 34: 125ml ss preserved with 5ml 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 159: S, SL; S, WA; NS; NS; sampler agitated; 125ml ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)	**ZOOPLANKTON** Anthomedusae Ef,Hc,Sp	#/73-30 PNV	#/ss	3	4	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	2	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	4	11	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Leptomedusae O	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	genus	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/73-30 PNV	#/ss	4	47	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	74	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	2	22	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	4	143	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Narcomedusae Al	#/73-30 PNV	#/ss	2	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)		#/571-50 PNV	#/ss	2	10	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	17	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Siphonophora Da	#/73-30 PNV	#/ss	2	7	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	8	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Thecosomata Sh,un.	#/73-30 PNV	#/ss	3	7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	11	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, species	4	19	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)	Polychaeta un.	#/73-30 PNV	#/ss	6	97	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	6	97	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	6	97	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Diplostraca D	#/73-30 PNV	#/ss	1	18	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	55	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		identification	genus	2	73	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/ss	13	8624	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	13	8624	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	13	8624	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida A,Ac,Al,Cg,Ch, D,Dt,E,Eg,Eh, Lm,Ml,Mpy,Pm	#/73-30 PNV	#/ss	16	1914	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	2	127	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	2	907	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	16	2948	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	16	2948	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)		sex	NA	16	1689	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida E,En,T	#/73-30 PNV	#/ss	5	37	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	5	37	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	1	10	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida C,Ce,CY,Ob,Om, un.	#/73-30 PNV	#/ss	15	1410	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	22	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5, 8	
		identification	order, genus, species	15	1432	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	15	1431	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	11	371	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/ss	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)	Mysidacea M	#/73-30 PNV	#/ss	10	25	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	10	30	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cumacea D	#/73-30 PNV	#/ss	3	8	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	3	8	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	Amphipoda Ab,Al,Hg,Hm, MO,Pab,Pg,Pl	#/73-30 PNV	#/ss	6	32	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/ss	1	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/ss	1	12	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	8	49	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	3	25	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
		sex	NA	1	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Euphausiacea Tr	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)		#/571-50 PNV	#/sa	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	furcilia #	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage #	2:4,5,8	
	Decapoda Eg,Sa,un.	#/73-30 PNV	#/sa	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNV	#/sa	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	2	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Insecta un.	#/73-30 PNV	#/sa	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Chaetognatha Eh,Se	#/73-30 PNV	#/sa	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/sa	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/sa	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	8	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0002 (cont'd)	Chordate un.	#/73-30 PNV	#/sa	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Copelata Fb,Ov	#/73-30 PNV	#/sa	4	257	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/sa	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		#/571-50 PNV	#/sa	2	61	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	4	319	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/73-30 PNV	#/sa	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/571-50 PNH	#/sa	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,2,4,5,8	
		age	larva	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
73-0003A	BACTERIA un.	#/NBS	log CFU/L	1	relative	METHOD 40	METHOD 38	METHOD 158	METHOD 39	2:1,4-6,8	
		identification	relative	1	relative	METHOD 40	METHOD 38	METHOD 158	heterotrophs counts graphed	2:4-6,8	

METHOD 3: sa taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: sa taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 38: aseptically transferred to cold, sterile 1L polypropylene bottles; maintained in chilled condition; processed within 4h of collection; spin-plate technique used to dispense aliquot of sample with cold pipette onto surface of cold agar plate [ZoBell Marine Broth 2216E (Difco) with seawater salts previously dissolved in deionized water and solidified with 1.2% Bacto-agar (Difco)]; placed in 5°C incubator; when aliquot absorbed by medium, plate inverted; incubated at 5°C.

METHOD 39: quadruplicate spin-plates made of each sample; plates with uneven distribution of colonies discarded; colonies of 3 plates of a replicate set enumerated; counts graphed.

METHOD 40: S; S; NS; NS; S/NS stn. depth; collected aseptically prior to other oceanographic sampling.

METHOD 158: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)	ZOOPLANKTON										
	Cnideria un.	#/73-30 PNV	#/se	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	relative	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	medusa; ABS taxonomic manual used	2:4,5,8	
	Anthomedusae Hc	#/73-30 PNV	#/s; #/se	2; 1	46; 1	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	species	3	47	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Rotifera un.	#/73-30 PNV	#/se	2	8	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	2	8	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/s; #/se	1; 2	63; 3	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	phylum	2	66	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/se	2	299	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	2	299	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	299	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Pelecypoda un.	#/73-30 PNV	#/se	2	211	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	2	211	S; S; WA; S/NS; 1; no; NS stn. depth	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)		age	larva	2	211	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Polychaeta un.	#/73-30 PNV	#s; #/ss	1; 3	107; 1480	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	3	1587	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	3	1587	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Diplostraca E,En,Pl	#/73-30 PNV	#/ss	2	40	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	2	40	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
		sex	NA	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Copepods un.	#/73-30 PNV	#/s; #/ss	1; 3	384; 13484	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	3	13868	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	3	13868	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoids A,Ac,Al,Db,Dt, Eh,ER,Lm,Pm	#/73-30 PNV	#/s; #/ss	2; 3	1046; 12190	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	genus, species	3	13236	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)		age	copepodite #	3	13236	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	3	6835	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida A,De,E,Ef,L,P, Tf,Z,un.	#/73-30 PNV	#/s; #/ss	1; 3	30; 85	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	order, genus, species	3	115	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult; relative	2	32	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage; immature	2:4,5,8	
		sex	NA	2	16	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida Cs,CY,Ob,un.	#/73-30 PNV	#/s; #/ss	1; 3	5; 107	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	order, genus, species	3	112	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult; relative	2	50	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage; immature	2:4,5,8	
		sex	NA	2	21	S; S, WA; S/NS; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/s; #/ss	1; 2	7; 1323	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	genus	2	1330	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius, cyprid	2	1330	S; S, WA; S; 1; no; S/NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)	Mysidacea Mr	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Cumacea D	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Insecta un.	#/73-30 PNV	#/ss	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Other un.	#/73-30 PNV	#/s; #/ss	2	407; 2825	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	#	2:1,4,5,8	
		age	egg	2	3272	S; S, WA; S/NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	stage	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)	EPONTIC ALGAE										
	Centrales Cb,CH,CO,Ce,M, R,T,Te,Tg,Tn, Tno,un.	#/S core	# cells/L	1	1791335	METHOD 25	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	1	1791335	preserved sections shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,Ad,AH,Akk, Akr,Aks,All, AP,Apr,Bp,Ck, Cl,CO,Dla,DP, Dm,Fp,G,Ge, Gep,Gf,Ld,Lg, N,Na,Nac,Nc, Nce,Ncl,Ncp, Ncr,Ncy,Nda, Nde,Ndr,Ndt, Nf,Nfr,Ngd, Nge,Ngl,Ngr, Nhy,Nk,Nkj, Nla,Nlt,Nm,No, Npo,Nre,Nsb, Nso,Nsr,Nss, NT,Ntd,Nte, Ntg,Nti,Ntr, Nve,Nvm,P,Pan, Pc,PL,Pmi,Pq, Pqb,Pqc,Pqd, Pqs,Pqu,Psd, Pse,Psm,Psu, Si,Sj,Soe,St, Sto,SY,Tm,TR, un.	#/S core	# cells/L	1	35446067	METHOD 25	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	1	35446067	preserved sections shaken; 35 mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chlorophyceae un.	#/S core	# cells/L	1	18130	METHOD 25	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	

METHOD 25: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw; NS stn. depth.

METHOD 27: each section agitated; each 1mL ss of bottom section and 10mL ss of other sections pipetted or poured into individual 10mL Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 41: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-0003A (cont'd)		identification	class	1	18130	METHOD 25	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
	Euglenophyceae E	#/S core	# cells/L	1	7252	METHOD 25	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	genus	1	7252	METHOD 25	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
	Dinophyceae Gc,un.	#/S core	# cells/L	1	583813	METHOD 25	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	class, species	1	583813	METHOD 25	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
73-0003B	ZOOPLANKTON Tintinnida un.	#/73-30 PNV	#/m^3	14	402058	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	order	14	402058	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Anthomedusae Hc	#/73-30 PNV	#/m^3	1	1	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	species	1	1	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Leptomedusae O	#/73-30 PNV	#/m^3	4	24	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	genus	4	24	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 25: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw; NS stn. depth.

METHOD 27: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 29: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 41: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 42: calculated densities underestimated but probably < factor of 2; true filtration efficiency closer to 86%; refs for filtration efficiency given.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-00038 (cont'd)	Trachymedusae Ad	#/73-30 PNV	$\#/m^3$	1	1	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	species	1	1	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Rotifera un.	#/73-30 PNV	$\#/m^3$	12	63739	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	phylum	12	63739	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Polychaeta un.	#/73-30 PNV	$\#/m^3$	11	5166	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	class	11	5166	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
		age	larva	11	5166	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 5	stage	2:3-5,8	
	Diplostraca B,Dl,Dm,En,Pl	#/73-30 PNV	$\#/m^3$	18	4814	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	genus, species	18	4814	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Crustacea un.	#/73-30 PNV	$\#/m^3$	18	469183	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	class	18	469183	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
		age	nauplius	18	469183	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 5	stage	2:3-5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 42: calculated densities underestimated but probably < factor of 2; true filtration efficiency closer to 86%; refs for filtration efficiency given.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
73-00038 (cont'd)	Calanoida Ac,Al,Cg,Db, Do,Dt,Ea,Eh, El,Lm,P	#/73-30 MAY	$\#/m^3$	18	186357	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	genus, species	18	186357	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Harpacticoida un.	#/73-30 MAY	$\#/m^3$	6	116	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	order	6	116	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Cyclopoida un.	#/73-30 MAY	$\#/m^3$	10	1159	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	order	10	1159	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
	Cirripedia un.	#/73-30 MAY	$\#/m^3$	4	191	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		identification	order	4	191	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	ABS taxonomic manual used	2:3-5,8	
		age	cyprid	4	191	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 5	stage	2:3-5,8	
	Other un.	#/73-30 MAY	$\#/m^3$	18	29411	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	densities calculated assuming 100% filtration efficiency	METHOD 42	2:1-5,8	
		age	egg	18	29411	S; S, WA; NS; 1; no; NS stn. depth; ?date	buffered F	METHOD 6	stage	2:3-5,8	
74-0001	PHYTOPLANKTON										
	Centrales Ch,un.	#/73?-12 MAY	extrapolated cells/l	3	6.0	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6, 8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 30: 3 x 1ml ss taken; examined with microscope @ x100 magnif.; concentrations calculated from mean no./taxa and H_2O vol. strained.
METHOD 42: calculated densities underestimated but probably < factor of 2; true filtration efficiency closer to 80%; refs for filtration efficiency given.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0001 (cont'd)		identification	order, genus	3	4.8	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Pennales un.	#/73?-12 WNV	extrapolated cells/L	18	474.3	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	order	18	474.3	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Chlorophyceae un.	#/73?-12 WNV	extrapolated cells/L	17	4649.8	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	class	17	4649.8	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	ZOOPLANKTON Rotifera K,KE,N	#/73?-12 WNV	extrapolated cells/L	17	135	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	genus	17	135	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
	Copepoda un.	#/73?-12 WNV	extrapolated cells/L	1	1.6	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4-6,8	
		identification	subclass	1	1.6	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1 mL ss taken; examined with microscope at x100 magnif.	NS	2:4-6,8	
		age	adult	1	1.6	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1 mL ss taken; examined with microscope at x100 magnif.	stage	2:4-6,8	
	Other un.	#/73?-12 WNV	extrapolated cells/L	11	60.4	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	METHOD 30	3 ss examined; #	2:1,2,4,5,8	
		age	nauplius	11	60.4	S; S; NS; NS; NS; NS stn. depth; ?date	5% F	3 x 1mL ss taken; examined with microscope at x100 magnif.	stage	2:4,5,8	

METHOD 30: 3 x 1ml ss taken; examined with microscope @ x100 magnif.; concentrations calculated from mean no./taxa and H_2O vol. strained.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0001 (cont'd)	EPONTIC ALGAE Pennales un.	#/scraping	NS	1	relative	scraped off ice broken up during drilling	NS	examined by microscope	high density	2:all	
		identification	order	1	relative	scraped off ice broken up during drilling	NS	examined by microscope	high density; NS	2:all	
	EPONTIC INVERTEBRATES Crustacea un.	#/scraping	NS	1	relative	scraped off ice broken up during drilling	NS	examined by microscope	few	2:all	
		identification	class	1	relative	scraped off ice broken up during drilling	NS	examined by microscope	few; NS	2:all	
		age	nauplius	1	relative	scraped off ice broken up during drilling	NS	examined by microscope	stage; few; NS	2:all	
74-0003	PHYTOPLANKTON Centrales M	#/PVC	# chains/m^3; # cells/m^3	6	$94.99x10^4$ $18.99x10^6$	1L PVC; S; S; NS	METHOD 43	METHOD 44	#	2:1,4-6,8	
		identification	genus	6	$94.99x10^4$ $18.99x10^6$	1L PVC; S; S; NS	METHOD 43	METHOD 44	NS	2:4-6,8	
	Cyanophyceae un.	#/PVC	# chains/m^3	2	31490	1L PVC; S; S; NS	METHOD 43	METHOD 44	#	2:1,4-6,8	
		#/72-50 PNH	NS	4	155+	METHOD 45	METHOD 48	METHOD 46	NS	2:1,4-6,8	
		identification	class	4	31490; 155+	PVC = 1L PVC; S; S; NS 72-50 PNH = METHOD 45	PVC = METHOD 43 72-50 PNH = METHOD 48	METHOD 47	NS	2:4-6,8	
	ZOOPLANKTON Trematoda un.	#/72-50 PNH	#/s	1	1	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
		identification	class	1	1	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	

METHOD 43: 3mL Lugol (Utermohl's formula) added to each bottle so no air trapped beneath the cap; bottles tightly sealed; labelled; maintained in dark at room temp.
METHOD 44: each sample split (ref. for method description given); ss examined with Wilde M-11 inverted microscope and gridded counting plate.
METHOD 45: S; S; S; 1; S, Gurly TS Pigmy flow meter rigidly mounted in front of net mouth; net drawn vertically along side of boat; outside of mesh lightly sprayed with water; contents of net bucket rinsed with squeeze bottle into wide-mouthed sample jar.
METHOD 46: each sample split (ref. for method description given); ss examined with Wilde M-5 dissecting microscope and gridded counting plate.
METHOD 47: each sample split (ref. for method description given); ss examined either with Wilde M-11 inverted microscope or Wilde M-5 dissecting microscope and gridded counting plate.
METHOD 48: 2% F buffered; maintained in dark at room temp.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0003	Rotifera	#/72-50 PNH	#/m	9	78975	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
(cont'd)	K,Kl	identification	phylum	9	78975	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Pelecypoda	#/72-50 PNH	#/m	1	1	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	un.	identification	class	1	1	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Diplostraca	#/72-50 PNH	#/m	2	2	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	(Cladocera)	identification	suborder	2	2	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Ostracoda	#/72-50 PNH	#/m	1	1	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	un.	identification	subclass	1	1	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Calanoida	#/72-50 PNH	#/m	21	13276	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	Dm,Dp,Ene,Lm,	identification	order, species	21	13276	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	METHOD 49
	un.	age	relative	12	1285	METHOD 45	METHOD 48	METHOD 46	copepodite, juvenile	2:3-6,8	
	Harpacticoida	#/72-50 PNH	#/m	5	37	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	un.	identification	order	5	37	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Cyclopoida	#/72-50 PNH	#/m	10	2796	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	C,Cb	identification	genus, species	10	2796	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Mysidacea	#/72-50 PNH	#/m	17	105	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	Mr	identification	species	17	105	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Amphipoda	#/72-50 PNH	#/m	14	131	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	H,G,P	identification	genus	14	131	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Diptera	#/72-50 PNH	#/m	8	15	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
	Hs,T	identification	genus, species	8	15	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
		age	larva, pupa; adult	5	10	METHOD 45	METHOD 48	METHOD 46	stage	2:3-6,8	

METHOD 45: 5; 5; 5; 1; 5, Gurly 15 Pigmy flow meter rigidly mounted in front of net mouth; net drawn vertically along side of boat; outside of mesh lightly sprayed with water; contents of net bucket rinsed with squeeze bottle into wide-mouthed sample jar.

METHOD 46: each sample split (ref. for method description given); ss examined with Wild M-5 dissecting microscope and gridded counting plate.

METHOD 48: 2% F buffered; maintained in dark at room temp.

METHOD 49: Dp and Lm reported as Diaptomus pribolofensis and Limnocalanus macrurus, respectively.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0001 (cont'd)	Ephemeroptera (Heptageniidae)	#/72-50 PNH	#/s	5	5	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-6,8	
		identification	family	5	5	METHOD 45	METHOD 48	METHOD 46	NS	2:3-6,8	
	Other un.	#/72-50 PNH	#/s	8	10335	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-5,8	
		age	nauplius	8	10335	METHOD 45	METHOD 48	METHOD 46	stage	2:3-5,8	
	ICHTHYOPLANKTON Other un.	#/72-50 PNH	#/s	5	45	METHOD 45	METHOD 48	METHOD 46	#	2:1,3-5,8	
		age	larva	5	45	METHOD 45	METHOD 48	METHOD 46	stage	2:3-5,8	
74-0007A	BACTERIA un.	#/NDS	$\log_{10}$ CFU/l; on marine agar; on PC agar	16	510.99; 454.12	METHOD 50	METHOD 51	METHOD 150	METHOD 33	2:1,4-6,8	
		identification	relative	16	956.11	METHOD 50	METHOD 51	METHOD 150	heterotrophs	2:4-6,8	
	PHYTOPLANKTON Centrales CH,CY,M,Ma, Mi?,R,SI,T, Tg?,un.	#/VD	# cells/l	1	26×10^4.	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	order, genus, species	1	26×10^4.	preserved 125ml ss shaken; 5ml ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	

METHOD 28: 35ml ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 10ml dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 33: quadruplicate spin-plates made of each sample on both media; plates with uneven distribution of colonies discarded; colonies of 3 plates at a replicate set enumerated; #.

METHOD 34: 125ml ss preserved with 5ml 40% formaldehyde neutralized with $CaCO_3$; stored in polyethylene bottles.

METHOD 35: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 45: 5; 5; 5; 1; 5, Gurly 15 Pigmy flow meter rigidly mounted in front of net mouth; net drawn vertically along side of boat; outside of mesh lightly sprayed with water; contents of net bucket rinsed with squeeze bottle into wide-mouthed sample jar.

METHOD 46: each sample split (ref. for method description given); ss examined with Wild M-5 dissecting microscope and gridded counting plate.

METHOD 48: 2% F buffered; maintained in dark at room temp.

METHOD 50: 5; 5, M4; NS; NS; collected aseptically prior to the oceanographic sampling.

METHOD 51: aseptically transferred to cold, sterile 1l polypropylene bottles; processed within 4h of collection; spin-plate technique used to disperse aliquot of sample with cold pipette onto surface of cold agar plate [both ZoBell Marine Broth 2216E (Difco) and Plate Count Broth (Difco) previously dissolved separately in deionized water and solidified with 1.2% Bacto agar (Difco)]; placed in 5°C incubator; when aliquot absorbed by medium, plate inverted; incubated at 5°C.

METHOD 52: 5, 5l; 5, M4; NS; NS; sampler agitated; 125ml ss drawn off.

METHOD 150: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION ACCURACY		
74-0007A (cont'd)	Pennales A,AH,DP,Fcy,N, Ncl,NT,S,In, un.	#/VD	# cells/l	3	75000	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	order, genus, species	3	75000	preserved 125ml ss shaken; 35ml ss drawn off	METHOD 51	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chrysophyceae un.	#/VD	# cells/l	1	2000	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	class	1	2000	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Euglenophyceae E	#/VD	# cells/l	3	$14x10^4$	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	genus	3	$14x10^4$	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Dinophyceae A,Gc,P,un.	#/VD	# cells/l	3	90000	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	class, genus, species	3	90000	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Other un.	#/VD	# cells/l	3	$21.04x10^5$	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	relative	3	$21.04x10^5$	METHOD 52	METHOD 34	METHOD 37	pigmented flagellates, filamentous algae; NS	2:5,8	
	ZOOPLANKTON										
	Cnidaria un.	#/73-30 PNV	#/m	3	14	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	3	14	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Anthomedusae Eh,Hr	#/73-30 PNV	#/m	2	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 34: 125ml ss preserved with 5ml 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 51: aseptically transferred to cold, sterile 1l polypropylene bottles; processed within 4h of collection; spin-plate technique used to dispense aliquot of sample with cold pipette onto surface of cold agar plate [both ZoBell Marine Broth 2216E (Difco) and Plate Count Broth (Difco) previously dissolved separately in deionized water and solidified with 1.2% Bacto-agar (Difco)]; placed in 5°C incubator; when aliquot absorbed by medium, plate inverted; incubated at 5°C.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125ml ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)		#/569-50 PNO	#/mm	1	4	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	6	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/73-30 PNV	#/m	13	92	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	13	92	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Narcomedusae Al	#/73-30 PNV	#/m	6	28	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	6	28	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Siphonophora C,Da	#/73-30 PNV	#/m	3	6	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	4	7	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Anthozoa un.	#/73-30 PNV	#/m; #/mm	1; 2	2; 7	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	4	10	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	3	9	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)	Beroide Bc	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/s; #/ss	6; 8	128; 86	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	12	214	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	12	214	S; S, WA; S; 1, no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Thecosomata Sh	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Polychaeta un.	#/73-30 PNV	#/m; #/ss	9; 7	89; 33	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	13	122	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	trochophore, larva	13	121	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)	Myodocopa Cb	#/73-30 PNV	#/m	5	18	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	13	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	5	31	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		sex	NA	5	31	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	NS	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/m; #/mm	2; 14	5; 9031	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 1	#	2:1,4,5,8	
		identification	subclass	15	9036	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	15	9035	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida A,Al,Cc,Cg,Ch, Co,Dt,Eb,Eg, Gt,Hn,Lm,Ml, Mp,Mpy,Pm,Sm, Smi	#/73-30 PNV	#/m; #/mm	15;14	778; 2912	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/569-50 PNO	#/m; #/mm	1	183; 1146	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	15	5019	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	15	5019	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	15	3402	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida E,He,M,Pm,un.	#/73-30 PNV	#/m; #/mm	8; 9	125; 212	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)		identification	genus, species	12	337	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius; relative	3	3	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage; immature	2:4,5,8	
		sex	NA	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida CY,Ob,Om	#/73-30 PNV	#/m; #/m³	4; 15	14; 3562	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	15	3576	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	15	3576	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	15	1776	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Monstrilloida M	#/73-30 PNV	#/m	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Cirripedia un.	#/73-30 PNV	#/m³	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	cyprid	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/m³	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)		identification	genus	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Mysidacea M,Ml	#/73-30 PNV	#/m	5	10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	565	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	6	575	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Isopoda un.	#/73-30 PNV	#/m; #/ms	9; 3	13; 10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	1	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order	11	24	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	cryptoniscid, larva	10	17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Amphipoda Ag,Am,M,MO, Pab,Pg,Pl	#/73-30 PNV	#/m	9	29	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/m	1	76	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	10	105	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	6	64	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature, young, younger	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)	Euphausiacea Ti,Tl,Tr,un.	#/73-30 PNV	#/s	5	76	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/s	1	494	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order, species	5	570	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Decapoda un.	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Chaetognatha Eh,Se,un.	#/73-30 PNV	#/s	12	55	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/569-50 PNO	#/s	1	320	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum, species	13	375	S; S, WA; S/NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Copelata Fb,Ov	#/73-30 PNV	#/s; #/ss	9; 6	222; 199	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/569-50 PNO	#/s; #/ss	1; 1	1; 2	S; S, WA; NS; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	10	424	S; S, WA; S/N; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Other un.	#/73-30 PNV	#/s; #/ss	1; 12	2; <1916	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007A (cont'd)		age	egg, larva	12	<1913	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
74-0007C	BACTERIA										
	un.	#/NBS	$\log_{10}$ CFU/L: on marine agar agar; on PC agar	8	262.31; 208.53	METHOD 50	METHOD 51	METHOD 158	METHOD 33	2:1,4-6,8	
		identification	relative	8	470.84	METHOD 50	METHOD 51	METHOD 158	heterotrophs	2:4-6,8	
	PHYTOPLANKTON										
	Centrales Cc,Cf,CH,Co, CY,Ma,Mg,Mi, Sa,Sh,T,Tb,Tq, un.	#/VD	# cells/L	5	5.18×10^5+	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	order, genus, species	5	5.18×10^5+	preserved 125ml ss shaken; 35ml ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,Af,AH,Ak,AP, CM,CO,De,Dl, Dlc,Do,DP,Ds, Fc,Fcy,Fo,G, Gk,Gw,GY,L,N, Nb,Nc,Ncl,Nd?, Ndc,Ndr,Nfr, Nhu,Nk?,Np, Nsb,Nsc,Nsi, NT,Nv,Psm,Pst, S,Sa,Sf,Sh,So, SY,Tfl,Tn,TR, un.	#/VD	# cells/L	5	5.04×10^5+	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
		identification	order, genus, species	5	5.04×10^5+	preserved 125ml ss shaken; 35ml ss drawn off	METHOD 51	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; AHS taxonomic manual used.

METHOD 28: 35ml ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30ml dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 33: quadruplicate spin-plates made of each sample on both media; plates with uneven distribution of colonies discarded; colonies of 3 plates at a replicate set enumerated; #.

METHOD 34: 125ml ss preserved with 5ml 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 50: S; S, WA; NS; NS; collected aseptically prior to the oceanographic sampling.

METHOD 51: aseptically transferred to cold, sterile 1l polypropylene bottles; processed within 4h of collection; spin-plate technique used to dispense aliquot of sample with cold pipette onto surface of cold agar plate [both ZoBell Marine Broth 2216E (Difco) and Plate Count Broth (Difco) previously dissolved separately in deionized water and solidified with 1.2% Bacto-agar (Difco)]; placed in 5°C incubator; when aliqunt absorbed by medium, plate inverted; incubated at 5°C.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125ml ss drawn off.

METHOD 158: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C	Chlorophyceae	#/VD	# cells/l	6	96000	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
(cont'd)	C,un.	identification	class, genus	6	96000	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Chrysophyceae	#/VD	# cells/l	3	37000+	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
	DN,Et,un.	identification	class, genus, species	3	37000+	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Cyanophyceae	#/VD	# cells/l	1	1000	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
	C	identification	genus	1	1000	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Euglenophyceae	#/VD	# cells/l	3	2.76×10^5	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
	E	identification	genus	3	2.76×10^5	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Dinophyceae	#/VD	# cells/l	5	5.75×10^5+	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
	A?,Gc,P,Pc?, Pm,un.	identification	class, genus, species	5	5.75×10^5+	METHOD 52	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Other	#/VD	# cells/l	6	10.58×10^5+	METHOD 52	METHOD 34	METHOD 35	METHOD 36	2:1,5,8	
	un.	identification	relative	6	10.58×10^5+	METHOD 52	METHOD 34	METHOD 37	pigmented flagellates; taxonomic references stated	2:5,8	
	ZOOPLANKTON										
	Foraminiferida	#/73-90 PNV	#/ss	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
	un.	identification	order	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	AHS taxonomic manual used	2:4,5,8	
	Radiolaria un.	#/73-90 PNV	# ss	1	10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; AHS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 34: 125ml ss preserved with 5ml 40% formaldehyde neutralized with $CaCO_3$; stored in polyethylene bottles.
METHOD 35: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.
METHOD 36: x500 magnif.; 630 microscope fields selected; # cells in Whipple grid counted.
METHOD 37: 125ml ss agitated; 10ml ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.
METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125ml ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		identification	subclass	1	10	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cnidaria un.	#/73-30 PNV	#/s	1	7	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	1	7	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/73-30 PNV	#/s	6	42	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	21	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	6	63	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Narcomedusae Al	#/73-30 PNV	#/s	2	2	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	1	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	3	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Siphonophora Da,Mb,un.	#/73-30 PNV	#/s	4	20	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order, species	4	20	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/s; #/ss	1; 2	3; 2	5; 5, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscore, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		identification	phylum	3	5	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/s; #/ss	2; 6	24; 101	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	18	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	7	143	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	7	143	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Thecosomata S,Sh	#/73-30 PNV	#/s	2	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	2	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Polychaeta un.	#/73-30 PNV	#/s; #/ss	6; 2	88; 2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	7	92	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	trochophore, larva	7	92	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Myodocopa Cb	#/73-30 PNV	#/s	4	17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	4	17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		age	relative	2	5	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
		sex	NA	4	10	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/ss	8	5530	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	47	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	subclass	8	5577	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	8	5577	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida A,Ac,Cg,Ch,Db, Dt,F,Eg,Eh,Hn, Lm,Ml,Mpy,Pm, Sm,Smi	#/73-30 PNV	#/s; #/ss	8; 7	672; 1262	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:1,4,5,8	
		#/233-30 PNV	#/s	1	81	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:1,4,5,8	
		identification	genus, species	8	2015	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	8	2015	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	8	863	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida Da,E,En,Ha,M, P,Pa,Tf	#/73-30 PNV	#/s; #/ss	8; 3	195; 53	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/s	1	1	5; S, WA; 5; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		identification	genus, species	8	249	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	3	21	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	6	75	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida Ub,Om	#/73-30 PNV	#/m^3	8	2066	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/m	1	191	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	8	2257	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	8	2257	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	8	1046	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cirripedia un.	#/73-30 PNV	#/m	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	subclass	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	cyprid	1	1	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/m	2	9	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	2	9	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		age	nauplius	2	9	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Cumacea Dr	#/73-30 PMV	#/m	1	12	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	12	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	12	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
		sex	NA	1	12	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Isopoda un.	#/73-30 PMV	#/m; #/ss	5; 2	15; 2	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order	5	17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	cryptoniscid, larva	5	17	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Amphipoda Ag,MU,Pab,Pl	#/73-30 PMV	#/m	6	22	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	6	22	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	4	10	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
	Euphausiacea Tr,un.	#/73-30 PMV	#/m; #/ss	1; 2	2; 23	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, species	3	25	S; S, WA; S; 1; no	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0007C (cont'd)		age	nauplius	2	23	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	
	Decapoda Ss	#/73-30 PNV	#/m	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	zoea	1	1	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	stage	2:4,5,8	
	Chaetognatha Fh,Se,un.	#/73-30 PNV	#/m	7	30	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		#/233-30 PNV	#/m	1	4	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum, species	7	34	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Copelata Fh,Ov	#/73-30 PNV	#/m; #/mm	4; 7	419; 75	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/m	1	14	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	7	508	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Other un.	#/73-30 PNV	#/m; #/mm	1; 8	2; <1234	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	#	2:1,4,5,8	
		#/233-30 PNV	#/m	1	21	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 5	#	2:1,4,5,8	
		age	egg, larva	8	<1255	S; S, WA; S; 1; no	5% F buffered with CaCO$_3$ or Na$_2$B$_4$O$_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0010A	ZOOPLANKTON										
	Anthomedusae Hc	#/73-30 PNV	#/m	2	15	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	15	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Rotifera un.	#/73-30 PNV	#/ms	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	1	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/m; #/ms	1; 1	43; 6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	phylum	2	49	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/m; #/ms	1; 1	2; 5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	class	2	7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Polychaeta un.	#/73-30 PNV	#/m; #/ms	2; 1	192; 5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	class	2	197	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	197	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0010A (cont'd)	Copepoda un.	#/73-30 PNV	#/s; #/ss	1; 2	204; 4169	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	subclass	2	4373	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius, larva	2	4373	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Al,Cg,Db, Df,Eh,Im,Pm	#/73-30 PNV	#/s; #/ss	2; 2	630; 389	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	species	2	1019	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	2	985	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #	2:4,5,8	
		sex	NA	2	936	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida A,E,P,un.	#/73-30 PNV	#/s; #/ss	1; 1	25; 10	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#; #	2:1,4,5,8	
		identification	order, genus	2	35	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	adult; relative	2	35	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage; immature	2:4,5,8	
		sex	NA	2	33	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Cyclopoida CY	#/73-30 PNV	#/s	1	13	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	13	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0010A (cont'd)		age	copepodite #; adult	1	11	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage #; adult	2:4,5,8	
		sex	NA	1	11	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/ss	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	EPONTIC ALGAE Centrales Cb,CH,Ckg,CO, Ce,M,Ms,T,Ta, Tg,Tn,un.	#/S core	# cells/L	2	2810305s	METHOD 53	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	2	2810305s	preserved sections shaken; 35ml ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 27: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 28: 35ml ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30ml dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 41: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 53: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0010A (cont'd)	Pennales A,AH,Ak,Akk, Akr,Aks,All, Alm,AP,Apr,Ck, Cl,CO,Dla,DP, Ds,Gep,N,Nn, Nc,Ncl,Ncp, Ncr,Ncy,Nda, Nde,Ndr,Nf, Nfr,Nge,Ngl, Nhy,Nk,Nkj,Nl, Nla,Nle,Nlt, Nm,No,Npo,Nqu, Nre,Nse,Nsr, Nss,NT,Nt,Ntd, Ntg,Nti,Ntr, Nva,P,Pan,Pc, PL,Pmi,Pq,Pqb, Pqc,Pqd,Pql, Pqs,Pqu,Psb, Psc,Psm,St,SY, Tm,un.	#/S core	# cells/L	2	55883328	METHOD 53	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	order, genus, species	2	55883328	preserved sections shaken; 35ml ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chlorophyceae un.	#/S core	# cells/L	2	870286	METHOD 53	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	class	2	870286	METHOD 53	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
	Euglenophyceae E	#/S core	# cells/L	1	36262	METHOD 53	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	
		identification	class	1	36262	METHOD 53	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
	Dinophyceae G,Gr,CY,un.	#/S core	# cells/L	2	1468604	METHOD 53	METHOD 41	METHOD 27	x500 magnif.; 89 microscope fields counted; #	2:1,4,5,8	

METHOD 27: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope.

METHOD 28: 35ml ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10ml 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30ml dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 29: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 41: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 53: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0010A (cont'd)		identification	class, genus, species	2	1468604	METHOD 53	METHOD 41	METHOD 29	taxonomic references stated	2:5,8	
74-0010C	BACTERIA B,C,D,E,K	#/Phleger core	#/g dry sediment	1	146360	METHOD 54	METHOD 55	METHOD 56	#	2:1,4-6,8	Remark 4 (75-0010B)
		identification	genus; relative	1	146360	METHOD 54	METHOD 55	METHOD 56	METHOD 57; NS	2:4-6,8	Remark 4 (75-0010B)
74-0011	ZOOPLANKTON										
	Cnidaria un.	#/505-60 FNH	mean # of 3 replicate samples	2	11	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	relative	2	11	METHOD 58	10% F	binocular microscope	medusa; NS	2:4-6,8	
	Ctenophora un.	#/505-60 FNH	mean # of 3 replicate samples	1	4	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	phylum	1	4	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Nematoda un.	#/505-60 FNH	mean # of 3 replicate samples	1	1	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	phylum	1	1	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Polychaeta un.	#/505-60 FNH	mean # of 3 replicate samples	2	58	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	class	2	58	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Oligochaeta un.	#/505-60 FNH	mean # of 3 replicate samples	1	14	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	class	1	14	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	

METHOD 29: each section agitated; each 1ml ss of bottom section and 10ml ss of other sections pipetted or poured into individual 10ml Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 41: sections placed in clean plastic containers; thawed at room temp.; immed. preserved in 2% F buffered with $CaCO_3$.

METHOD 53: 7.5cm SIPRE corer; tb; core length measured; sections cut with fine-toothed meat saw.

METHOD 54: S; NS; NS; collected in polycarbonate plastic liner (38mm OD, 1.5mm wall thickness, 270mm long) in Phleger corer; size 8 rubber stopper inserted and taped in place.

METHOD 55: packed in low-temp. equilibrated polystyrene Trans-Temp containers; shipping temp. range of 4-12°C; stored at 40°C in lab.

METHOD 56: ref. for Plate Count (PC) and Most Probable Number (MPN) methods description given; PC of total aerobic heterotrophs used Difco Tryptic Soy Broth with Oxoid Ion agar; MPN of facultatively anaerobic N_2 fixers used sucrose + mineral salts + growth factor medium, sets of tubes incubated aerobically and anaerobically; MPN of N_2-fixing clostridia used glucose + acetate + mineral salts + phenosafranin + soil extract medium incubated anaerobically; MPN of SO_4 reducers used lactate + yeast extract + mineral salts + steel wool medium incubated anaerobically.

METHOD 57: total aerobic heterotrophs, facultative N_2 fixers, N_2-fixing clostridia, SO_4 reducers.

METHOD 58: S; S; NS; NS; NS; sketch of net included; details of towing at each stn. included.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0011 (cont'd)	Ostracoda un.	#/505-60 FNH	mean # of 3 replicate samples	1	22	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	subclass	1	22	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Copepoda un.	#/505-60 FNH	mean # of 3 replicate samples	3	44519	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	subclass	3	44519	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Mysidacea un.	#/505-60 FNH	mean # of 3 replicate samples	3	193	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	order	3	193	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Isopoda un.	#/505-60 FNH	mean # of 3 replicate samples	1	7	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	order	1	7	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Amphipoda un.	#/505-60 FNH	mean # of 3 replicate samples	3	282	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	order	3	282	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Diptera (Chironomidae)	#/505-60 FNH	mean # of 3 replicate samples	2	57	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	family	2	57	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
	Chaetognatha un.	#/505-60 FNH	mean # of 3 replicate samples	1	10	METHOD 58	10% F	binocular microscope	range of # of organisms given; #	2:1,2,4-6,8	
		identification	phylum	1	10	METHOD 58	10% F	binocular microscope	NS	2:4-6,8	
74-0019	PHYTOPLANKTON										
	Pennales AS?,SY?	#/100-30 WNV	#/ml	4	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	rare (≤5/ml)	2:1,2,4-6,8	
		identification	genus	4	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	rare, NS	2:4-6,8	

METHOD 58: S; S; NS; NS; NS; sketch of net included; details of towing at each stn. included.
METHOD 59: 1ml aliquots on a Petroff-Hauser bacterial enumeration cell examined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0019 (cont'd)	Chlorophyceae un.	#/100-30 WNV	#/mL	1	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	rare (≤5/mL)	2:1,2,4-6, 8	
		identification	relative	1	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	filamentous green algae; rare; NS	2:4-6,8	
	ZOOPLANKTON Rotifera un.	#/100-30 WNV	#/mL	7	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	rare (≤5/mL)	2:1,2,4-6, 8	
		identification	phylum	7	relative	S; NS; NS; NS; NS; ?loc.; NS stn. depth; date rel.	50% (v/v) ethanol	METHOD 59	rare; NS	2:4-6,8	
74-0027	PHYTOPLANKTON Centrales Cbr,Cct,Cd, Cdi,Cg,Clo,Cm, Co,Cmo,Pg,R, Sc,Tg,Tn	#/VD	#/s	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present, common, dominant	2:1,4-6,8	Logol's solution should probably be Lugol.
		identification	genus, species	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present, common, dominant; NS	2:4-6,8	Logol's solution should probably be Lugol.
	Pennales At,Foc,Nde, Nso,Tnt	#/VD	#/s	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present, common, dominant	2:1,4-6,9	Logol's solution should probably be Lugol.
		identification	species	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present, common, dominant; NS	2:4-6,8	Logol's solution should probably be Lugol.
	Chrysophyceae D	#/VD	#/s	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	common, dominant	2:1,4-6,8	Logol's solution should probably be Lugol.
		identification	genus	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	common, dominant; NS	2:4-6,8	Logol's solution should probably be Lugol.
	Dinophyceae Ce,Pp	#/VD	#/s	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present	2:1,4-6,8	Logol's solution should probably be Lugol.
		identification	species	3	relative	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	present; NS	2:4-6,8	Logol's solution should probably be Lugol.

METHOD 59: 1mL aliquots on a Petroff-Hauser bacterial enumeration cell examined.

METHOD 60: each 500mL sample placed in volumetric cylinder; 3mL Logol's iodine solution added; settled for 2 days; supernatant siphoned off leaving approx. 20mL sample; trasferred to plastic vial; kept in dark at 5°C.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
74-0027 (cont'd)	ZOOPLANKTON Cnidaria un.	#/60-25 and 240-50 PNH and PNV	#/s	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present	2:1,2,4-6, 8	
		identification	relative	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	medusa; present; NS	2:4-6,8	
	Anthomedusae Sp	#/60-25 and 240-50 PNH and PNV	#/s	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present	2:1,2,4-6, 8	
		identification	species	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present; NS	2:4-6,8	
	Stauromedusae Cc	#/60-25 and 240-50 PNH and PNV	#/s	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present	2:1,2,4-6, 8	
		identification	species	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present; NS	2:4-6,8	
	Cydippida Pp	#/60-25 and 240-50 PNH and PNV	#/s	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present	2:1,2,4-6, 8	
		identification	species	2	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present; NS	2:4-6,8	
	Rotifera un.	#/60-25 and 240-50 PNH and PNV	#/s	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common	2:1,2,4-6, 8	
		identification	phylum	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common; NS	2:4-6,8	
	Copepoda un.	#/60-25 and 240-50 PNH and PNV	#/s	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common, dominant	2:1,2,4-6, 8	
		identification	subclass	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common, dominant; NS	2:4-6,8	
		age	nauplius	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	stage; common, dominant; NS	2:4-6,8	
	Calanoida A,Cg,Eh,Lm, Mpy,Pm	#/60-25 and 240-50 PNH and PNV	#/s	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common, dominant	2:1,2,4-6, 8	
		identification	genus, species	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present, common, dominant; NS	2:4-6,8	
	Cyclopoida Os	#/60-25 and 240-50 PNH and PNV	#/s	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present	2:1,2,4-6, 8	
		identification	species	3	relative	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chambers	present; NS	2:4-6,8	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0004	PHYTOPLANKTON										
	Centrales M	#/PVC	estimated # cells/m^3	2	3.91×10^7	1L PVC; S; NS; NS; S/? loc.; NS stn. depth	METHOD 61	METHOD 62	#	2:1,2,4-6,8	
		identification	genus	2	3.91×10^7	1L PVC; S; NS; NS; S/? loc.; NS stn. depth	METHOD 61	METHOD 62	NS	2:4-6,8	
	Pennales NT,T	#/PVC	estimated # cells/m^3	2	66.94×10^5	1L PVC; S; NS; NS; NS stn. depth	METHOD 61	METHOD 62	#	2:1,2,4-6,8	
		identification	genus	2	66.94×10^5	1L PVC; S; NS; NS; NS stn. depth	METHOD 61	METHOD 62	NS	2:4-6,8	
	Dinophyceae C	#/PVC	estimated # cells/m^3	1	3.55×10^6	1L PVC; S; NS; NS; NS stn. depth	METHOD 61	METHOD 62	#	2:1,2,4-6,8	
		identification	genus	1	3.55×10^6	1L PVC; S; NS; NS; NS stn. depth	METHOD 61	METHOD 62	NS	2:4-6,8	
	Other un.	#/PVC	estimated # cells/m^3	12	315.64×10^{11}	1L PVC; S; NS; NS; S/? loc.; NS stn. depth	METHOD 61	METHOD 63	#	2:1,2,4,5,8	
		identification	relative	12	315.64×10^{11}	1L PVC; S; NS; NS; S/? loc.; NS stn. depth	METHOD 61	METHOD 63	nannoplankton; NS	2:4,5,8	
	ZOOPLANKTON										
	Rotifera K,KE	#/72-50 PNH	#/m	5	912942	METHOD 64	METHOD 65	METHOD 66	Rotifera generally counted from 1% ss	2:all	
		identification	genus	5	912942	METHOD 64	METHOD 65	METHOD 67	NS	2:all	
	Pelecypoda un.	#/72-50 PNH	#/m	1	20	METHOD 64	METHOD 65	enumerated with Wilde M-5 dissection microscope	#	2:all	
		identification	class	1	20	METHOD 64	METHOD 65	identified with Wilde M-11 microscope	NS	2:all	
	Copepoda un.	#/72-50 PNH	#/m	4	147885	METHOD 64	METHOD 65	METHOD 66	#	2:all	
		identification	subclass	4	147885	METHOD 64	METHOD 65	METHOD 67	NS	2:all	
		age	egg, nauplius, copepodite	4	147885	METHOD 64	METHOD 65	METHOD 67	stage	2:all	

METHOD 61: 60ml ethyl alcohol (6% v/v) added to bottle so no air trapped beneath the cap; bottles tightly sealed; maintained in dark at ambient temp.
METHOD 62: sample transferred to 2L Erlenmeyer flask; swirled vigourously; 1ml aliquot examined @ x100 magnif. with Wilde M-11 inverted microscope.
METHOD 63: sample transferred to 2L Erlenmeyer flask; swirled vigourously; 0.05ml aliquot examined @ x400 magnif. with Wilde M-11 inverted microscope.
METHOD 64: S; S; NS; 0.44; Gurly TS Pigmy flow meter rigidly mounted in front of net mouth; S/? loc.; NS stn. depth.
METHOD 65: 6% v/v ethyl alcohol maintained in dark at ambient temp.
METHOD 66: each sample split (ref. for method description given); enumerated with Wilde M-5 dissection microscope.
METHOD 67: each sample split (ref. for method description given); identified with Wilde M-11 microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0004 (cont'd)	Calanoida Ac,Al,Cp,Dp, Eca,Ene,Lm,Pm, Sc	#/72-50 PNH	#/m	11	17519	METHOD 64	METHOD 65	METHOD 66	#	2:all	
		identification	species	11	17519	METHOD 64	METHOD 65	METHOD 67	NS	2:all	
		age	nauplius, copepodite	11	17519	METHOD 54	METHOD 65	METHOD 67	stage	2:all	
	Harpacticoida un.	#/72-50 PNH	#/m	2	132	METHOD 64	METHOD 65	METHOD 66	#	2:all	
		identification	order	2	132	METHOD 64	METHOD 65	METHOD 67	NS	2:all	
	Cyclopoida Cb	#/72-50 PNH	#/m	5	4362	METHOD 64	METHOD 65	METHOD 66	#	2:all	
		identification	species	5	4362	METHOD 64	METHOD 65	METHOD 67	NS	2:all	
		age	nauplius, copepodite	5	4362	METHOD 64	METHOD 65	METHOD 67	stage	2:all	
	Mysidacea Mr	#/72-50 PNH	#/m	2	38	METHOD 64	METHOD 65	enumerated with Wilde M-5 dissection microscope	#	2:all	
		identification	species	2	38	METHOD 64	METHOD 65	identified with Wilde M-11 microscope	NS	2:all	
	Amphipoda un.	#/72-50 PNH	#/m	1	1	METHOD 64	METHOD 65	enumerated with Wilde M-5 dissection microscope	#	2:all	
		identification	order	1	1	METHOD 64	METHOD 65	identified with Wilde M-11 microscope	NS	2:all	
	ICHTHYOPLANKTON Other un.	#/72-50 PNH	#/m	1	1	METHOD 64	METHOD 65	enumerated with Wilde M-5 dissection microscope	#	2:1-5,8	
		age	larva	1	1	METHOD 64	METHOD 65	identified with Wilde M-11 microscope	stage	2:3-5,8	
75-0006	PHYTOPLANKTON Centrales CH,Cco,Ccv,Cw, M,Id,In	#/73? MNH	# cells/m^3	7	2097.8 x10^3	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	#	2:all	
		identification	genus, species	7	2097.8 x10^3	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	NS	2:all	
	Pennales FR,Nae,NT	#/73? MNH	# cells/m^3	5	948.1x10^3	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	#	2:all	
		identification	genus, species	5	948.1x10^3	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	NS	2:all	

METHOD 64: S; S; NS; 0.44; Gurly TS Pigmy flow meter rigidly mounted in front of net mouth; S/? loc.; NS stn. depth.
METHOD 65: 6% v/v ethyl alcohol maintained in dark at ambient temp.
METHOD 66: each sample split (ref. for method description given); enumerated with Wilde M-5 dissection microscope.
METHOD 67: each sample split (ref. for method description given); identified with Wilde M-11 microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0006 (cont'd)	ZOOPLANKTON Thecosomata Sh	#/73? MNH	#/m^3	2	0.33	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	#	2:all	
		identification	species	2	0.33	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	NS	2:all	
	Calanoida Cf,Ch,Eh,Lm	#/73? MNH	#/m^3	6	82.64	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	#	2:all	
		identification	species	7	82.64+	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	NS	2:all	
	Amphipoda Pl	#/73? MNH	#/m^3	2	0.18	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	#	2:all	
		identification	species	2	0.18	S; NS; NS; 2-2.5; NS; sample washed into SS bucket	formaldehyde	NS	NS	2:all	
75-0009	ZOOPLANKTON Thecosomata S	#/IKT	#/m	2	26	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	genus	2	26	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Gymnosomata Cl	#/IKT	#/m	5	26	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	species	5	26	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Myodocopa Pg	#/IKT	#/m	1	9	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	species	1	9	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0009 (cont'd)	Calanoida Cg,Ch,Lm,un.	#/IKT	#/s	4	ap. 659	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	order, species	4	ap. 659	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	2	49	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage #	2:4,5,8	
		sex	NA	2	49	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Mysidacea Ml,un.	#/IKT	#/s	5	ap. 56	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	order, species	5	ap. 56	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Amphipoda Hm,Gl,Gw,Pl	#/IKT	#/s	8	40	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,2,4,5,8	
		identification	species	8	40	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		size, overall	relative	1	3	S; NS; NS; NS; NS; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	large, very large	2:4,5,8	
75-0010A	ZOOPLANKTON Polychaeta un.	#/73-30 PNV	#/ss	1	11	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	11	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	11	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0010A (cont'd)	Copepoda un.	#/73-30 PNV	#/ss	1	321	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	1	321	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	321	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Ac,Db,Lm,Pm	#/73-30 PNV	#/s	1	148	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	148	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	148	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
		sex	NA	1	146	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Harpacticoida un.	#/73-30 PNV	#/s	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	order	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
	Cyclopoida CV	#/73-30 PNV	#/s	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	adult	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0010A (cont'd)		sex	NA	1	1	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Other un.	#/73-30 PNV	#/ss	1	322	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	#	2:1,4,5,8	
		age	egg	1	322	S; S, WA; NS; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	stage	2:4,5,8	
75-0010B	BACTERIA B,C,D,E,K	#/Petersen grab	#/g dry sediment	1	146360	METHOD 68	METHOD 55	METHOD 56	#	2:1,2,4-6,8	Remark 4 (74-0010C)
		identification	genus; relative	1	146360	METHOD 68	METHOD 55	METHOD 56	METHOD 57	2:4-6,8	Remark 4 (74-0010C)
	PHYTOPLANKTON Centrales	#/VD	# cells/L	2	9.49×10^5+	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
	Cc,Cce,CH,Cn, CO?,CY,M,T,un.	identification	order, genus, species	2	9.49×10^5+	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.
METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.
METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.
METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.
METHOD 52: S, 5L; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.
METHOD 55: packed in low-temp. equilibrated polystyrene Trans-Temp containers; shipping temp. range of 4-12°C; stored at 40°C in lab.
METHOD 56: ref. for Plate Count (PC) and Most Probable Number (MPN) methods description given; PC of total aerobic heterotrophs used Difco Tryptic Soy Broth with Oxoid Ionagar; MPN of facultatively anaerobic N_2 fixers used sucrose + mineral salts + growth factor medium, sets of tubes incubated aerobically and anaerobically; MPN of N_2-fixing clostridia used glucose + acetate + mineral salts + phenosafranin + soil extract medium incubated anaerobically; MPN of SO_4 reducers used lactate + yeast extract + mineral salts + steel wool medium incubated anaerobically.
METHOD 57: total aerobic heterotrophs, facultative N_2 fixers, N_2-fixing clostridia, SO_4 reducers.
METHOD 68: S; NS; NS; approx. 1-3kg wet weight collected; transferred to gas-impermeable plastic bags.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0010B (cont'd)	Pennales A,AH,Al,Alc, AP,Ar,CO,De, Dl,DP,Dm,Fcy, Gf,Gk,Gw,GY,L, N,Nc,Ncl,Ndi, Ndr,Nf,Nh,Nhu, Np,Nr,Nm,Nmc, Nme,NT,P?,Pc, PL,S,Sf,SU,SY, un.	#/VD	# cells/L	2	1.21×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	2	1.21×10^5	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chrysophyceae Et,un.	#/VD	# cells/L	2	23000+	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, species	2	23000+	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Euglenophyceae E	#/VD	# cells/L	1	relative	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36; present	2:1,4,5,8	
		identification	genus	1	relative	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	present; taxonomic references stated	2:5,8	
	Dinophyceae A,D,Dm,Gc,P, Pm,un.	#/VD	# cells/L	2	1.89×10^5+	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, genus, species	2	1.89×10^5+	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Other un.	#/VD	# cells/L	2	5.54×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36; present	2:1,4,5,8	
		identification	relative	2	5.54×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	pigmented flagellates; NS	2:5,8	

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A	BACTERIA B,C,D,E,K	#/NBS	$\log_{10}$ CFU/L: on Lib X; on PC agar	4	169.62; 104.88	METHOD 69	METHOD 70	METHOD 158	METHOD 33	2:1,4-6,8	
		#/Petersen grab	#/g dry sediment	1	5586	METHOD 71	METHOD 55	METHOD 56	#	2:1,2,4-6,8	
		identification - NBS	relative	4	274.5	METHOD 69	METHOD 70	METHOD 158	heterotrophs	2:4-6,8	
		identification - Petersen grab	genus; relative	1	5586	METHOD 71	METHOD 55	METHOD 56	METHOD 57	2:4-6,8	
	PHYTOPLANKTON Centrales	#/VD	# cells/L	3	5.87×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
	Cce,CH,Cs,Cso, Cw,Ez,Ms?,Pg, T,Tn,un.	identification	order, genus, species	3	5.87×10^5	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,AH,AP,At,Dl,	#/VD	# cells/L	3	4.16×10^5+	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
	DP,Fcy,Fo,G, Gk,N,Nc,Ncl, Nd?,Ndr,Nfr, Nk?,Np,Nse,NT, Ps,S,Sh,Tn,un.	identification	order, genus, species	3	4.16×10^5+	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chlorophyceae C	#/VD	# cells/L	1	relative	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 33: quadruplicate spin-plates made of each sample on both media; plates with uneven distribution of colonies discarded; colonies of 3 plates at a replicate set enumerated; #.

METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

METHOD 55: packed in low-temp. equilibrated polystyrene Trans-Temp containers; shipping temp. range of 4-12°C; stored at 40°C in lab.

METHOD 56: ref. for Plate Count (PC) and Most Probable Number (MPN) methods description given; PC of total aerobic heterotrophs used Difco Tryptic Soy Broth with Oxoid Ionagar; MPN of facultatively anaerobic N_2 fixers used sucrose + mineral salts + growth factor medium, sets of tubes incubated aerobically and anaerobically; MPN of N_2-fixing clostridia used glucose + acetate + mineral salts + phenosafranin + soil extract medium incubated anaerobically; MPN of SO_4 reducers used lactate + yeast extract + mineral salts + steel wool medium incubated anaerobically.

METHOD 57: total aerobic heterotrophs, facultative N_2 fixers, N_2-fixing clostridia, SO_4 reducers.

METHOD 69: S; S, WA; NS; NS; NS stn. depth; collected aseptically prior to other oceanographic sampling.

METHOD 70: aseptically transferred to cold, sterile 1L polypropylene bottles; processed within 4h of collection; spin-plate technique used to disperse aliquot of sample with cold pipette onto surface of cold agar plate [both Lib X agar (ref. given) and Plate Count Broth (Difco) previously prepared and dissolved in 1L of 17% Instant Ocean and deionized water, respectively, PCB solidified with 1.2% Bacto-agar (Difco)]; placed in 5°C incubator; when aliquot absorbed by medium, plate inverted; incubated at 5°C.

METHOD 71: approx. 1-3kg wet weight collected; transferred to gas-impermeable plastic bags.

METHOD 158: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)		identification	genus	1	relative	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	present; taxonomic references stated	2:5,8	
	Chrysophyceae Db,Et,un.	#/VD	# cells/L	3	54000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, species	3	54000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Euglenophyceae E	#/VD	# cells/L	2	41000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	genus	2	41000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Dinophyceae A,Gc,P,Pm,un.	#/VD	# cells/L	3	91000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, genus, species	3	91000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Other un.	#/VD	# cells/L	3	15×10^4	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	relative	3	15×10^4	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	pigmented flagellates; NS	2:5,8	
	ZOOPLANKTON Porifera un.	#/73-30 PNV	#/s	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/73-30 PNV	#/s	3	21	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	3	21	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 52: S, 5L; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)	Narcomedusae Al	#/73-30 PNV	#/m	2	16	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	16	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Siphonophora Da,Mb	#/73-30 PNV	#/m	2	34	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	34	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Anthozoa un.	#/73-30 PNV	#/ss	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gastropoda un.	#/73-30 PNV	#/m; #/ss	1; 2	17; 15	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	3	32	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	3	32	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Thecosomata Sh	#/73-30 PNV	#/m	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Polychaeta P,un.	#/73-30 PNV	#/m; #/ss	2; 2	51; 34	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)		identification	class, genus	3	85	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	3	85	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Myodocopa Cb	#/73-30 PNV	#/s; #/ss	2; 1	106; 14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	2	120	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
		sex	NA	1	11	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Copepoda un.	#/73-30 PNV	#/ss	3	1125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	3	1125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	3	1125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Calanoida Cg,Ch,Co,Dt, Eg,Gt,Hn,Lm, Ml,Mp,Mpy,Pm, S,Sb,Sl,Sm, Smi,Tb	#/73-30 PNV	#/s; #/ss	3; 3	424; 710	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	3	1134	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #	3	1134	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage #	2:4,5,8	
		sex	NA	3	456	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)	Harpacticoida E,He,M,un.	#/73-30 PNV	#/s; #/ss	3; 2	27; 5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, genus, species	3	32	S; S, WA, S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Cyclopoida O,Ob,Om,On,Os	#/73-30 PNV	#/ss	3	1164	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	3	1164	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; relative	3	1162	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; immature	2:4,5,8	
		sex	NA	3	469	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Thoracica B	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Cumacea Do	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
	Isopoda un.	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)		identification	order	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Amphipoda Am,Pab	#/73-30 PNV	#/s	3	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	3	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
	Euphausiacea T,Ti	#/73-30 PNV	#/s	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	calyptopis	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Chaetognatha Eh,Se	#/73-30 PNV	#/s; #/ss	3; 2	5; 19	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	3	24	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Ophiuroidea un.	#/73-30 PNV	#/ss	2	125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	2	125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	ophiopluteus	2	125	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012A (cont'd)	Copelata Fb,O,Ov	#/73-30 PNV	#/s; #/ss	1; 3	39; 258	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	3	297	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Other un.	#/73-30 PNV	#/s; #/ss	1; 1	ap. 50; 50	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	#	2:1,4,5,8	
		age	egg	2	ap. 100	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	stage	2:4,5,8	
75-0012B	ZOOPLANKTON										
	Anthomedusae Hc	#/73-30 PNV	#/s	2	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	6	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Rotifera un.	#/73-30 PNV	#/s; #/ss	2; 1	3; 5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	phylum	3	8	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Nematoda un.	#/73-30 PNV	#/s	2	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	2	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Pelecypoda un.	#/73-30 PNV	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-00128 (cont'd)		age	larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Polychaete un.	#/73-30 PNV	#/s; #/ss	1; 2	15; 640	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	class	3	655	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	larva	3	655	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	Tardigrada un.	#/1000-100 PNH	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	phylum	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Diplostraca B,D,Pl	#/73-30 PNH	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/73-30 PNV	#/s; #/ss	3; 1	4; 1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	genus, species	5	[illegible]	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Copepoda un.	#/73-30 PNH	#/s	1	25	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/73-30 PNV	#/s; #/ss	3; 3	17; 935	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	subclass	7	977	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	nauplius	7	977	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	

METHOD 3: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-00128 (cont'd)	Calanoida Cg,D,Db,Dt,E, Eh,Lm,Pm	#/73-30 PNV	#/s; #/ss	3; 3	203; 3084	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		#/1000-100 PNH	#/s	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	7	3369	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	copepodite #; adult; relative	6	3368	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage #; stage; immature	2:4,5,8	
		sex	NA	5	377	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	NS	2:4,5,8	
	Harpacticoida un.	#/73-30 PNV	#/s; #/ss	1; 1	36; 7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order	2	43	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	7	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	Cyclopoida CY,un.	#/73-30 PNH	#/s	1	52	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/73-30 PNV	#/s; #/ss	3; 2	86; 17	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	order, genus	6	155	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	3	15	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	Mysidacea MI	#/73-30 PNV	#/s	2	21	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012B (cont'd)		identification	species	2	21	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	9	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	immature	2:4,5,8	
	Amphipoda Gl,M,Pa,Pg	#/73-30 PNH	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/73-30 PNV	#/s	1	3	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		#/1000-100 PNH	#/s	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus, species	3	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	adult; relative	3	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage; immature	2:4,5,8	
		sex	NA	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	NS	2:4,5,8	
	Insecta un.	#/73-30 PNV	#/s	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	class	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	2	4	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
	Copelata Fb	#/73-30 PNV	#/ss	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.
METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.
METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012B (cont'd)	Other un.	#/73-30 PNV	#/a; #/as	1; 3	4; 256	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	#	2:1,4,5,8	
		age	egg	1	260	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	stage	2:4,5,8	
	ICHTHYOPLANKTON SC	#/1000-100 PNH	#/a	1	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	genus	1	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
		age	larva	1	4	S; S, WA; S; 1; no; NS atn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	stage	2:4,5,8	
75-0012D	PHYTOPLANKTON										
	Centrales Cce,CY,Mg,Mi?, Pg,R,ST,T,un.	#/VD	# cells/L	3	$3.27x10^5$	METHOD 52; NS atn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	3	$3.27x10^5$	preserved 125mL as shaken; 35mL as drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,Af,AH,Ap,Ar, Ca,CY,De,Ds,E, N,Nbi?,Ncl,NT, SU,SY,Tfl,un.	#/VD	# cells/L	3	25000	METHOD 52; NS atn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	3	25000	preserved 125mL as shaken; 35mL as drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Euglenophyceae T	#/VD	# cells/L	1	1000	METHOD 52; NS atn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	genus	1	1000	METHOD 52; NS atn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	

METHOD 4: as taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 28: 35mL as centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 34: 125mL as preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL as agitated; 10mL as pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125mL as agitated; 10mL as pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125mL as drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012D (cont'd)	Other un.	#/VD	# cells/L	2	3000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	relative	2	3000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	pigmented flagellates; NS	2:5,8	
75-0012E	**BACTERIA** un.	#/NBS	$\log_{10}$ CFU/L: on Lib X; on PC agar	5	162.39; 106.38	METHOD 69	METHOD 70	METHOD 158	METHOD 33	2:1,4-6,8	
		identification	relative	5	268.77	METHOD 69	METHOD 70	METHOD 158	heterotrophs	2:4-6,8	
	PHYTOPLANKTON Centrales C,Cce,Cd,CH, CO?,Cs,Cso,Cw, Ms,Pg,Sc,T,Tn, un.	#/VD	# cells/L	3	154.09x 10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	3	154.09x 10^5	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Pennales A,AH,AP,AL,CH, CO,De,DI,DP, Fcy,Fo,Gk,GY, N,Ncl,Ndr,Nk?, Np,Nr,Nse,NT, S,SY,T,un.	#/VD	# cells/L	3	2.33×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	order, genus, species	3	2.33×10^5	preserved 125mL ss shaken; 35mL ss drawn off	METHOD 28	Leitz phase-contrast compound microscope @ x500 magnif.	taxonomic references stated	2:5,8	
	Chrysophyceae Db,Et,un.	#/VD	# cells/L	3	2.52×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, species, relative	3	2.52×10^5	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	choano-flagellates; taxonomic references stated	2:5,8	
	Euglenophyceae	#/VD	# cells/L	1	64000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	

METHOD 28: 35mL ss centrifuged @ 2000 rpm for 20 min.; supernatant decanted; 5-10mL 30% H_2O_2 added; cells resuspended; tubes placed in water bath (100°C) for 1h approx.; cooled to room temp.; washed x2 with 30mL dist. H_2O; centrifuged; supernatant decanted; few drops of suspension placed on coverslip; dried on hot plate; mounted in Hyrax on slide.

METHOD 33: quadruplicate spin-plates made of each sample on both media; plates with uneven distribution of colonies discarded; colonies of 3 plates at a replicate set enumerated; #.

METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

METHOD 69: S; S, WA; NS; NS; NS stn. depth; collected aseptically prior to other oceanographic sampling.

METHOD 70: aseptically transferred to cold, sterile 1L polypropylene bottles; processed within 4h of collection; spin-plate technique used to dispense aliquot of sample with cold pipette onto surface of cold agar plate [both Lib X agar (ref. given) and Plate Count Broth (Difco) previously prepared and dissolved in 1L of 17% Instant Ocean and deionized water, respectively, PCB solidified with 1.2% Bacto-agar (Difco)]; placed in 5°C incubator; when aliquot absorbed by medium, plate inverted; incubated at 5°C.

METHOD 158: incubation continued for 3 weeks; acridine orange stain used; plates examined by epifluorescent microscopy with appropriate light filters.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012F (cont'd)		identification	genus	1	64000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Dinophyceae Da,P,un.	#/VD	# cells/L	3	45000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	class, genus, species	3	45000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	taxonomic references stated	2:5,8	
	Other un.	#/VD	# cells/L	3	49000	METHOD 52; NS stn. depth	METHOD 34	METHOD 35	METHOD 36	2:1,4,5,8	
		identification	relative	3	49000	METHOD 52; NS stn. depth	METHOD 34	METHOD 37	pigmented flagellates; NS	2:5,8	
	ZOOPLANKTON Anthomedusae Hc	#/1000-100 PNH	#/s	2	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Leptomedusae Mo	#/1000-100 PNH	#/s	2	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	5	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Trachymedusae Ad	#/1000-100 PNH	#/s	2	157	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	157	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Thecosomata Sh	#/1000-100 PNH	#/ss	2	203	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 34: 125mL ss preserved with 5mL 40% formaldehyde neutralized with $CaCO_3$; stored in polyethlylene bottles.

METHOD 35: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; counted with Leitz inverted microscope @ x500 magnif.

METHOD 36: x500 magnif.; 638 microscope fields selected; # cells in whipple grid counted.

METHOD 37: 125mL ss agitated; 10mL ss pipetted or poured into Zeiss sedimentation chamber; settled for 12-24h; where necessary slide mounts were prepared; identified with Leitz phase-contrast compound microscope @ x500 magnif.

METHOD 52: S, SL; S, WA; NS; NS; sampler agitated; 125mL ss drawn off.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0012E (cont'd)		identification	species	2	203	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
	Gymnosomata Cl	#/1000-100 PNH	#/a	2	140	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 5	#	2:1,4,5,8	
		identification	species	2	140	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 6	ABS taxonomic manual used	2:4,5,8	
	Amphipoda Pl	#/1000-100 PNH	#/a	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	relative	1	14	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	immature	2:4,5,8	
	Decapoda Sa	#/1000-100 PNH	#/a	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	#	2:1,4,5,8	
		identification	species	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	ABS taxonomic manual used	2:4,5,8	
		age	late larva	1	1	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 3	stage	2:4,5,8	
	ICHTHYOPLANKTON Other un.	#/1000-100 PNH	#/a	2	10	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	#	2:1,4,5,8	
		age	larva	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	stage	2:4,5,8	
		length, overall	cm	1	2	S; S, WA; S; 1; no; NS stn. depth	5% F buffered with $CaCO_3$ or $Na_2B_4O_7$	METHOD 4	NS	2:4,5,8	

METHOD 3: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 4: ss taken either by pipette or Folsom Splitter; where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

METHOD 5: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope; ABS taxonomic manual used.

METHOD 6: where necessary slide mounts were prepared; Zeiss dissecting microscope, compound microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0026	PHYTOPLANKTON Pennales un.	#/VD	# cells/L	4	4400	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	#	2:1,2,4-6,8	Logol's solution should probably be Lugol.
		identification	order	4	4400	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	NS	2:4-6,8	Logol's solution should probably be Lugol.
	Chrysophyceae un.	#/VD	# cells/L	4	3700	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	#	2:1,2,4-6,8	Logol's solution should probably be Lugol.
		identification	class	4	3700	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	NS	2:4-6,8	Logol's solution should probably be Lugol.
	Dinophyceae un.	#/VD	# cells/L	4	25500	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	#	2:1,2,4-6,8	Logol's solution should probably be Lugol.
		identification	class	4	25500	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	NS	2:4-6,8	Logol's solution should probably be Lugol.
	Cryptophyceae un.	#/VD	# cells/L	4	3000	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	#	2:1,2,4-6,8	Logol's solution should probably be Lugol.
		identification	class	4	3000	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	NS	2:4-6,8	Logol's solution should probably be Lugol.
	ZOOPLANKTON Tintinnida un.	#/VD	# cells/L	4	1400	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	#	2:1,4-6,8	Logol's solution should probably be Lugol.
		identification	order	4	1400	S; S; NS; NS; NS stn. depth	METHOD 60	examined using sediment chambers and Wilde inverted microscope	NS	2:4-6,8	Logol's solution should probably be Lugol.
	Cnidaria un.	#/60-25 and.or 240-50 PNV	#/m^3	4	110	S; NS; NS; NS; NS; NS stn. depth	8% neutralized r	examined in Sedgwick-Rafter counting chamber	#	2:1,2,4-6,8	
		identification	relative	4	110	S; NS; NS; NS; NS; NS stn. depth	8% neutralized r	examined in Sedgwick-Rafter counting chamber	medusa; NS	2:4-6,8	
	Copepoda un.	#/60-25 and.or 240-50 PNV	#/m^3	4	12850	S; NS; NS; NS; NS; NS stn. depth	8% neutralized r	examined in Sedgwick-Rafter counting chamber	#	2:1,2,4-6,8	

METHOD 60: each 500mL sample placed in volumetric cylinder; 3mL Logol's iodine solution added; settled for 2 days; supernatant siphoned off leaving approx. 20mL sample; transferred to plastic vial; kept in dark at 5°C.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0026 (cont'd)		identification	subclass	4	12850	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chamber	NS	2:4-6,8	
		age	nauplius	4	6480	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chamber	stage	2:4-6,8	
	Echinoidea un.	#/60-25 and.or 240-50 PNV	#/m^3	4	3830	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chamber	#	2:1,2,4-6,8	
		identification	class	4	3830	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chamber	NS	2:4-6,8	
		age	echinopluteus	4	3830	S; NS; NS; NS; NS; NS stn. depth	8% neutralized F	examined in Sedgwick-Rafter counting chamber	stage	2:4-6,8	
75-0028	PHYTOPLANKTON Other un.	#/PB	# cells/L	3	59.93x10^5	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	phyto- and zooplankton data combined	2:1,2,5,8	
		#/VD	# cells/L	4	618238	S, 2L; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	phyto- and zooplankton data combined	2:1,5,8	
	ZOOPLANKTON Other un.	#/PB	# cells/L	3	59.93x10^5	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	phyto- and zooplankton data combined	2:1,2,5,8	
		#/VD	# cells/L	4	618238	S, 2L; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	phyto- and zooplankton data combined	2:1,5,8	
	EPONTIC ALGAE Bacillariophyceae un.	#/NS	#/hole	NS	relative	NS	5% F in 1L polyethylene bottles	method reference given	present	2:1,2,5,8	
		identification	class	NS	relative	NS	5% F in 1L polyethylene bottles	method reference given	present; taxonomic reference given	2:5,8	
	Centrales CH,CO,CY,M	#/PB	#/melt pond	12	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus	12	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	Pennales AH,AP,CA,CM,CO,DP,F,GY,L,N,NT,P,PL,SY	#/PB	#/melt pond	12	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0028 (cont'd)		identification	genus	12	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	Chlorophyceae B,Bp,Cc,CH,Cl, Cm,Ew,Pt,Pe	#/NS	#/hole	NS	relative	NS	5% F in 1L polyethylene bottles	method reference given	present	2:1,2,5,8	
		#/PB	#/melt pond	7	relative	S; S; S/NS; NS	METHOD 160	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus, species	7	relative	S/NS; S; S/NS; NS	METHOD 160	method reference given	present; relative abundance percent; taxonomic reference given	2:5,8	
	Phaeophyceae F,Fd,Pl,S,Sa, Sk,Sf,St	#/NS	#/hole	NS	relative	NS	5% F in 1L polyethylene bottles	method reference given	present	2:1,2,5,8	
		#/PB	#/melt pond	7	relative	S; S; S/NS; NS	METHOD 160	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus, species	7	relative	S/NS; S; S/NS; NS	METHOD 160	method reference given	present; relative abundance percent; taxonomic reference given	2:5,8	
	Cyanophyceae A,C,U,un.	#/NS	#/hole	NS	relative	NS	5% F in 1L polyethylene bottles	method reference given	present	2:1,2,5,8	
		#/PB	#/melt pond	6	relative	S; S; S/NS; NS	METHOD 160	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus, species	6	relative	S/NS; S; S/NS; NS	METHOD 160	method reference given	epiphyte: present; relative abundance percent; taxonomic reference given	2:5,8	
	Dinophyceae C,P	#/PB	#/melt pond	2	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus	2	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	EPONTIC INVERTEBRATES										
	Mastigophora D	#/PB	#/melt pond	4	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	

METHOD 160: 5% F in 120mL and 1L (macrophytes) polyethylene bottles.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
75-0028 (cont'd)		identification	genus	4	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	Genus undecipherable in report
	Ciliophora un.	#/PB	#/melt pond	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	subphylum	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	Heterotrichida 5	#/PB	#/melt pond	4	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	genus	4	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	Crustacea un.	#/PB	#/melt pond	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	class	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
		age	nauplius	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
	Copepoda un.	#/PB	#/melt pond	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent	2:1,2,5,8	
		identification	subclass	1	relative	S; S; S/NS; NS	5% F in 120mL polyethylene bottles	method reference given	relative abundance percent; taxonomic reference given	2:5,8	
76-0003	PHYTOPLANKTON Centrales Ce,CH,CO,CY,M, Mi,Sa,Tb,Tn, un.	#/PB	# cells/L	14	1298509	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,4-6,8	
		identification	order, genus, species	14	1298509	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Pennales A,Af,Ao,D,F,N, NT,T,Ta,SY	#/PB	# cells/L	14	98903	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,4-6,8	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0003 (cont'd)		identification	genus, species	14	98903	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Chrysophyceae Cr,Da,E,Et,un.	#/PB	# cells/L	11	951	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,4-6,8	
		identification	order, genus, species	11	951	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Dinophyceae D,un.	#/PB	# cells/L	9	13520	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,4-6,8	
		identification	order, genus	9	13520	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Other un.	#/PB	# cells/L	11	35942	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,4,5,8	
		identification	relative	11	35942	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	cells, spores; NS	2:4,5,8	
	ZOOPLANKTON Foraminiferida un.	#/PB	# cells/L	4	10	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,3-6,8	
		identification	order	4	10	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100mL ethanol (10% v/v)	as taken; Wilde inverted microscope	taxonomist identified	2:all	
	Tintinnida un.	#/PB	# cells/L	3	6	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100ml ethanol (10% v/v)	as taken; Wilde inverted microscope	#	2:1,3-6,8	
		identification	order	3	6	S, 5 gal. nalgene; S; NS; NS; NS stn. depth	100ml ethanol (10% v/v)	as taken; Wilde inverted microscope	taxonomist identified	2:all	
	Cnidaria un.	#/156-50 WNH	mean #/m^3	1	0.1	S; NS; NS; NS; NS; NS stn. depth	12ml ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	phylum	1	0.1	S; NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Scyphozoa un.	#/156-50 WNV	mean #/m^3	3	0.4	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0003 (cont'd)		identification	class	3	0.4	S; S; NS; NS; NS; NS stn. depth	12ml ethanol	as taken	taxonomist identified	2:all	
	Rotifera un.	#/156-50 WNV	mean #/m^3	1	30.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	phylum	1	30.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Gastropoda un.	#/156-50 WNH	mean #/m^3	1	6.4	S; NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		#/156-50 WNV	mean #/m^3	2	1.7	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	class	3	8.1	S; S/NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Pelecypoda un.	#/156-50 WNV	mean #/m^3	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	class	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Polycheata un.	#/156-50 WNV	mean #/m^3	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	class	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
		age	larva	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	stage	2:all	
	Diplostraca? (Cladocera)	#/156-50 WNH	mean #/m^3	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	order	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	'order Calanoida' is taxonomically incorrect; taxonomist identified	2:all	
	Calanoida Ct,ER,Lm,Pm, SC,un.	#/156-50 WNH	mean #/m^3	5	22388.2	S; S/NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		#/156-50 WNV	mean #/m^3	12	6705.7	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	order, genus, species	17	29093.9	S; S/NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
		age	nauplius, copepodite	17	28354.2	S; S/NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	stage	2:all	
	Harpacticoida un.	#/156-50 WNH	mean #/m^3	1	0.1	S; NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		#/156-50 WNV	mean #/m^3	7	1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0003 (cont'd)		identification	order	8	1.1	S; S/NS; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Cyclopoida O,Oh,un.	#/156-50 WWV	mean #/m³	9	215.4	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	order, genus, species	9	215.4	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Cirripedia un.	#/156-50 WWV	mean #/m³	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	subclass	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
		age	cyprid	1	0.1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	stage	2:all	
	Mysidacea un.	#/156-50 WWH	mean #/m³	1	3.17	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		#/156-50 WWV	mean #/m³	3	1	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	order	4	4.17	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Amphipoda (Lysianassidae)	#/156-50 WWV	mean #/m³	2	0.2	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	family	2	0.2	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Euphausiacea un.	#/156-50 WWV	mean #/m³	3	0.8	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	order	3	0.8	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Chaetognatha S	#/156-50 WWV	mean #/m³	2	0.2	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	genus	2	0.2	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Copelata O	#/156-50 WWV	mean #/m³	1	0.5	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	5 tows combined to provide mean #	2:all	
		identification	genus	1	0.5	S; S; NS; NS; NS; NS stn. depth	12mL ethanol	as taken	taxonomist identified	2:all	
	Other un.	weight, dry	mg/m³	7	45.06	156-50 WWH and WWV; S; NS; NS; NS; NS stn. depth	12mL ethanol	NS	NS	2:1-5,8	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0004	PHYTOPLANKTON										
	Centrales CH,CY,Mi,Se,T, Tb,Tn	#/PB	# cells/L	4	44936	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus, species	4	44936	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Pennales Af,AS,D,F,N, NT,SY,T,Ta	#/PB	# cells/L	4	1684	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus, species	4	1684	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Chlorophyceae un.	#/PB	# cells/L	2	16.4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,4-6,8	
		identification	class	2	16.4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	NS	2:4-6,8	
	Chrysophyceae Cr,Et,un.	#/PB	# cells/L	3	1268.4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,4-6,8	
		identification	class, species; relative	3	1268.4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	spores: NS	2:4-6,8	
	Dinophyceae Ct,D	#/PB	# cells/L	2	3144	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus, species	2	3144	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	NS	2:4-6,8	
	ZOOPLANKTON										
	Tintinnida un.	#/PB	# cells/L	2	188	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	5 samples combined to provide mean #	2:1,3-6,8	
		identification	order	2	180	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	as taken; Wilde inverted microscope	NS	2:3-6,8	
	Calanoida Lm	#/156-10 MMH identification	#/m^3 species	2 2	109.74 109.74	METHOD 73 METHOD 73	METHOD 74 METHOD 74	as taken as taken	# NS	2:1,3-6,8 2:3-6,8	

METHOD 72: preserved with 60mL (6% v/v) or 100mL (10% v/v) ethanol; labelled; stored in cool dark place.
METHOD 73: S; S; NS; 0.44; Gurly 15 Pigmy flow meter mounted in front of net opening.
METHOD 74: transferred to 4oz. wide-mouth glass bottles; preserved with 12% v/v ethyl alcohol.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0004 (cont'd)	Cyclopoide Cac	age	nauplius	1	0.12	METHOD 73	METHOD 74	as taken	NS	2:3-6,8	
		#/156-10 MNH	#/m^3	1	0.35	METHOD 73	METHOD 74	as taken	#	2:1,3-6,8	
		identification	species	1	0.35	METHOD 73	METHOD 74	as taken	NS	2:3-6,8	
76-0005	PHYTOPLANKTON										
	Centrales CH,CO,CY,M,Mi, S,Sa,T,Tb,un.	#/PB	# cells/L	8	288185.8	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	order, genus, species	8	288185.8	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	METHOD 76	2:4-6	
	Pennales D,F,N,NT,SU, SY,T,TA	#/PB	# cells/L	7	5742.2	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus	7	5742.2	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	representative cells photographed; prints retained	2:4-6	
	Chrysophyceae E	#/PB	# cells/L	1	18	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus	1	18	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	representative cells photographed; prints retained	2:4-6	
	Dinophyceae D	#/PB	# cells/L	3	192	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	genus	3	192	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 72	METHOD 75	representative cells photographed; prints retained	2:4-6	
	Other un.	#/PB	# cells/L	2	1694	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	5 samples combined to provide mean #	2:1,4,5,8	

METHOD 72: preserved with 60mL (6% v/v) or 100mL (10% v/v) ethanol; labelled; stored in cool dark place.

METHOD 73: S; S; NS; 0.44; Gurly TS Pigmy flow meter mounted in front of net opening.

METHOD 74: transferred to 4oz. wide-mouth glass bottles; preserved with 12% v/v ethyl alcohol.

METHOD 75: 10mL aliquot filtered through Nucleopore 0.4u filter; attached to metal mounting pins with silver paint; plated with thin layer of gold; examined with ETEC Autoscan Model U-1 scanning electron microscope; light microscope also used.

METHOD 76: representative cells and unidentifiable organisms photographed; prints retained.

METHOD 77: preserved with 100mL (10% v/v) ethanol; labelled; stored in cool dark place.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0005 (cont'd)		identification	relative	2	1694	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	cells, spores; METHOD 76	2:4,5	
	ZOOPLANKTON										
	Foraminiferida un.	#/PB	# cells/L	1	2	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	order	1	2	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	METHOD 76	2:4-6	
	Tintinnida un.	#/PB	# cells/L	2	4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	5 samples combined to provide mean #	2:1,4-6,8	
		identification	order	2	4	S, 1L nalgene; S; NS; NS; NS stn. depth	METHOD 77	METHOD 75	METHOD 76	2:4-6	
	Calanoida Lm	#/156-10 MNH	#/m^3	2	260.1	METHOD 73	METHOD 74	METHOD 78	samples with <100 organisms counted completely	2:all	
		identification	species	2	260.1	METHOD 73	METHOD 74	appendages examined with Wilde M-11 microscope	NS	2:all	
		length, overall	mm	2	NS	METHOD 73	METHOD 74	NS	range	2:all	
76-0036	PHYTOPLANKTON										
	Centrales CH,M	#/VD	# cells/mL	2	0.8	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 80	METHOD 82	2:1,4-6,8	
		identification	genus	2	0.8	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	
	Pennales A,Af,D,Dp,N, Ncc,SY,T	#/VD	# cells/mL	3	83.5+	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 80	METHOD 82; #, present	2:1,4-6,8	
		identification	genus, species	3	83.5+	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	

METHOD 73: S; S; NS; 0.44; Gurly TS Pigmy flow meter mounted in front of net opening.
METHOD 74: transferred to 4oz. wide-mouth glass bottles; preserved with 12% v/v ethyl alcohol.
METHOD 75: 10mL aliquot filtered through Nucleopore 0.4u filter; attached to metal mounting pins with silver paint; plated with thin layer of gold; examined with ETEC Autoscan Model U-1 scanning electron microscope; light microscope also used.
METHOD 76: representative cells and unidentifiable organisms photographed; prints retained.
METHOD 77: preserved with 100mL (10% v/v) ethanol; labelled; stored in cool dark place.
METHOD 78: where necessary ss (range given) taken; examined with Wilde M-5 dissection microscope.
METHOD 79: transferred to clean, unused 1L polyethelene bottles; preserved with 60mL ethyl alcohol (6% v/v); placed in darkness and stored at ambient temp.
METHOD 80: sample shaken by hand; 5 or 25mL aliquot removed; allowed to settle for 24h in settling chamber; examined with Wilde M40 inverted microscope.
METHOD 81: 5-10mL ss filtered through Nucleopore filters; gold coated; examined with Phillips electron microscope.
METHOD 82: 3 fields scanned; # in each field counted; average # determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0036 (cont'd)	Chlorophyceae un.	#/VD	# cells/mL	1	0.1	S, 2L; S; S?; NS	METHOD 79	METHOD 80	METHOD 82	2:1,4-6,8	
		identification	class	1	0.1	S, 2L; S; S?; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	
	Chrysophyceae DN,M,un.	#/VD	# cells/mL	3	19.2	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 80	METHOD 82; #, present	2:1,4-6,8	METHOD 83
		identification	class, genus	3	19.2+	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	
		age	spore	3	18+	S, 2L; S; S?/NS; NS	METHOD 79	METHOD 81	stage	2:4-6,8	
	Cyanophyceae un.	#/VD	# cells/mL	1	14	S, 2L; S; S?; NS	METHOD 79	METHOD 80	METHOD 82	2:1,4-6,8	
		identification	class	1	14	S, 2L; S; S?; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	DN (Chrysophyceae) spores present but incorrectly listed as Cyanophyta
	Dinophyceae un.	#/VD	# cells/mL	2	0.9	S, 2L; S; S?; NS	METHOD 79	METHOD 80	METHOD 82	2:1,4-6,8	
		identification	class	2	0.9	S, 2L; S; S?; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	
	Cryptophyceae R	#/VD	# cells/mL	1	0.5	S, 2L; S; S?; NS	METHOD 79	METHOD 80	METHOD 82	2:1,4-6,8	
		identification	genus	1	0.5	S, 2L; S; S?; NS	METHOD 79	METHOD 81	1 taxonomist (name given)	2:4-6,8	
	ZOOPLANKTON Rotifera un.	#/80-12.7 BRNV	#/m	2	70	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		identification	phylum	2	70	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
	Ploima A,KE,K1	#/80-12.7 BRNH	#/m	1	672	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		#/80-12.7 BRNV	#/m	2	4092	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; examined in petri dish with Wilde M5 microscope	samples with <100 organisms counted completely; #	2:all	

METHOD 79: transferred to clean, unused 1L polyethelene bottles; preserved with 60mL ethyl alcohol (6% v/v); placed in darkness and stored at ambient temp.

METHOD 80: sample shaken by hand; 5 or 25mL aliquot removed; allowed to settle for 24h in settling chamber; examined with Wilde M40 inverted microscope.

METHOD 81: 5-10mL as filtered through Nucleopore filters; gold coated; examined with Phillips electron microscope.

METHOD 82: 3 fields scanned; # in each field counted; average # determined.

METHOD 83: phylum count greater than sum of species, unclear from where extras arise.

METHOD 84: transferred to 4oz. glass bottles; preserved with 6% (v/v) ethyl alcohol.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0036 (cont'd)		identification	genus, species	2	476A	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
	Crustacea un.	#/BO-12.7 BRNH	#/m	1	26	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		identification	class	1	26	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
		age	nauplius	1	26	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	stage	2:all	
	Diplostraca P	#/BO-12.7 BRNV	#/m	2	27	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		identification	order	2	27	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
	Ostracoda un.	#/BO-12.7 BRNV	#/m	1	26	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		identification	subclass	1	26	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
	Copepoda un.	#/BO-12.7 BRNH	#/m	1	120	S; S; S?; NS; NS	METHOD 84	METHOD 85	#	2:all	
		#/BO-12.7 BRNV	#/m	3	427	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	samples with <100 organisms counted completely; #	2:all	
		identification	subclass	3	547	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	1 taxonomist (name given)	2:all	
		age	nauplius	3	547	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	NS	2:all	
	Calanoida Dp,E,L,Lm,un.	#/BO-12.7 BRNH	#/m	1	60	S; S; S?; NS; NS	METHOD 84	METHOD 85	#	2:all	
		#/BO-12.7 BRNV	#/m	3	195	S; S; S?; NS; NS	METHOD 84	METHOD 85	#	2:all	
		identification	order, genus, species	3	255	S; S; S?; NS; NS	METHOD 84	METHOD 85	1 taxonomist (name given)	2:all	
		age	copepodite	3	95	S; S; S?; NS; NS	METHOD 84	METHOD 85	stage	2:all	
	Harpacticoida un.	#/BO-12.7 BRNH	#/m	1	5	S; S; S?; NS; NS	METHOD 84	METHOD 85	#	2:all	
		#/BO-12.7 BRNV	#/m	1	2	S; S; S?; NS; NS	METHOD 84	METHOD 85	#	2:all	
		identification	order	1	7	S; S; S?; NS; NS	METHOD 84	METHOD 85	1 taxonomist (name given)	2:all	
	Cyclopoida Cac,CY,un.	#/BO-12.7 BRNH	#/m	1	650	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	#	2:all	

METHOD 84: transferred to 4oz. glass bottles; preserved with 6% (v/v) ethyl alcohol.
METHOD 85: examined in petri dish with Wilde 5 dissection and M11 compound microscopes.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
76-0036 (cont'd)		#/80-12.7 BRNV	#/s	3	6477	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	samples with <100 organisms counted completely; #	2:all	
		identification	order, genus, species	3	7127	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	1 taxonomist (name given)	2:all	
		age	copepodite	3	6522	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; METHOD 85	stage	2:all	
	Diptera (Chironomidae)	#/80-12.7 BRNV	#/s	1	1	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		identification	family	1	1	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	1 taxonomist (name given)	2:all	
	Other un.	#/80-12.7 BRNH	#/s	1	7	S; S; S?; NS; NS	METHOD 84	examined in petri dish with Wilde M5 microscope	#	2:all	
		#/80-12.7 BRNV	#/s	2	742	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; examined in petri dish with Wilde M5 microscope	samples with <100 organisms counted completely; #	2:all	
		identification	relative	2	742	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; examined in petri dish with Wilde M5 microscope	rotifera and copepoda mainly; 1 taxonomist (name given)	2:all	
		age	egg	2	742	S; S; S?; NS; NS	METHOD 84	NS as taken by pipette; examined in petri dish with Wilde M5 microscope	stage	2:all	
77-0002	PHYTOPLANKTON Centrales CH,CY,R	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	genus	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	
	Pennales N	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	genus	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	

METHOD 84: transferred to 4oz. glass bottles; preserved with 6% (v/v) ethyl alcohol.

METHOD 85: examined in petri dish with Wilde 5 dissection and M11 compound microscopes.

METHOD 86: method reference given; sample agitated; 1-2.5mL is settled in phytoplankton settling chamber for 3h/cm of chamber height; examined with Wilde M40 inverted microscope @ x750 magnif.

METHOD 87: method reference given; sample agitated; 1-2.5mL is settled in phytoplankton settling chamber for 3h/cm of chamber height; examined with Wilde M40 inverted microscope @ up to x1875 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY COLLECTION	METHODOLOGY STORAGE	METHODOLOGY ANALYSIS	METHODOLOGY PRECISION/ACCURACY	RATING	REMARKS
77-0002 (cont'd)	Chlorophyceae un.	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	class	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	
	Chrysophyceae un.	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	class	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	
	Euglenophyceae un.	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	class	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	
	Cryptophyceae R	#/KB	# cells/mL	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 86	#	2:1,5,8	
		identification	genus	NA	NA	S; S; NS; NS; NS stn. depth; 125mL sample collected	Lugol's solution	METHOD 87	taxonomic references stated	2:5,8	
	ZOOPLANKTON										
	Anthomedusae Hc	#/NS-12 WNV and.or 392-60 FNH	NA	NA	NA	METHOD 88	5% F	METHOD 89	NS	2:1,5,8	
		identification	species	NA	NA	METHOD 88	5% F	METHOD 90	taxonomic references stated	2:5,8	
	Trachymedusae Ad	#/NS-12 WNV and.or 392-60 FNH	NA	NA	NA	METHOD 88	5% F	METHOD 89	NS	2:1,5,8	
		identification	species	NA	NA	METHOD 88	5% F	METHOD 90	taxonomic references stated	2:5,8	
	Beroida Bc	#/NS-12 WNV and.or 392-60 FNH	NA	NA	NA	METHOD 88	5% F	METHOD 89	NS	2:1,5,8	

METHOD 86: method reference given; sample agitated; 1-2.5mL ss settled in phytoplankton settling chamber for 3h/cm of chamber height; examined with Wilde M40 inverted microscope @ x750 magnif.

METHOD 87: method reference given; sample agitated; 1-2.5mL ss settled in phytoplankton settling chamber for 3h/cm of chamber height; examined with Wilde M40 inverted microscope @ up to x1875 magnif.

METHOD 88: S; NS; NS; NS; Tsurumi-Seiki (Model 10-1971) flow meter.

METHOD 89: NS ss taken by plankton splitter; examined with Wilde M5 binocular microscope @ x25 and/or x50 magnif.

METHOD 90: temporary mounts of whole or dissected specimens examined with Wilde M12 compound microscope @ x100, x400 or x1000 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0002 (cont'd)		identification	species	NA	NA	METHOD 88	5% F	METHOD 90	taxonomic references stated	2:5,8	
	Calanoida A,Lm,Pm	#/NS-12 WNV and.or 392-60 FNH	NA	NA	NA	METHOD 88	5% F	METHOD 89	NS	2:1,5,8	
		identification	genus, species	NA	NA	METHOD 88	5% F	METHOD 90	taxonomic references stated	2:5,8	
77-0003	ZOOPLANKTON										
	Cnidaria un.	#/NS N	#/m	1	relative	METHOD 91	METHOD 92	NS	present	2:all	
		#/75-NS MNH and.or 243-NS MNH	#/m	2	relative	METHOD 93	METHOD 92	NS	present	2:all	
		identification	relative	3	relative	METHOD 94	METHOD 92	NS	medusa; present; NS	2:all	
		size, overall	relative	3	relative	METHOD 94	METHOD 92	NS	small; present; NS	2:all	
	Trachymedusae Ad	#/NS N	#/m	2	relative	METHOD 91	METHOD 92	NS	present, % abundance	2:all	
		#/75-NS MNH and.or 243-NS MNH	#/m	14	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
		identification	species	16	relative	METHOD 94	METHOD 92	NS	present, % abundance; NS	2:all	
	Narcomedusae Al,un.	#/NS N	#/m	2	relative	METHOD 91	METHOD 92	NS	present, % abundance	2:all	
		#/75-NS MNH and.or 243-NS MNH	#/m	1	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
		identification	order, species	3	relative	METHOD 94	METHOD 92	NS	present, % abundance; NS	2:all	
	Siphonophora A?	#/NS N	#/m	1	relative	METHOD 91	METHOD 92	NS	present	2:all	
		identification	genus	1	relative	METHOD 91	METHOD 92	NS	present; NS	2:all	
	Thecosomata Sh	#/NS N	#/m	3	relative	METHOD 91	METHOD 92	NS	present, % abundance	2:all	
		#/75-NS MNH and.or 243-NS MNH	#/m	15	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
		identification	species	18	relative	METHOD 94	METHOD 92	NS	present, % abundance; NS	2:all	
	Gymnosomata Cl	#/NS N	#/m	1	relative	METHOD 91	METHOD 92	NS	present	2:all	
		#/75-NS MNH and.or 243-NS MNH	#/m	9	relative	METHOD 93	METHOD 92	NS	present	2:all	

METHOD 88: S; NS; NS; NS; Tsurumi-Seiki (Model 10-1971) flow meter.

METHOD 89: NS as taken by plankton splitter; examined with Wilde M5 binocular microscope @ x25 and/or x50 magnif.

METHOD 90: temporary mounts of whole or dissected specimens examined with Wilde M12 compound microscope @ x100, x400 or x1000 magnif.

METHOD 91: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed after and retrieved before Miller nets; initially deployed from ship's boom; later fastened with 2.5 cm polypropylene rope at bow, inboard towing wires on net frame shortened, net towed away from ship's side; sample removed from cod end without washing net down.

METHOD 92: when sufficient, sample split in Folsom Splitter; 1/8 preserved in 5% buffered F; rest stored in Whirl-pak bag or aluminum can and frozen.

METHOD 93: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed before neuston tows and recovered last; sample removed from cod end without washing net down.

METHOD 94: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; towing details given; sample removed from cod end without washing net down.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0003 (cont'd)		identification	species	10	relative	METHOD 94	METHOD 92	NS	present, NS	2:all	
		size, overall	relative	3	relative	METHOD 93	METHOD 92	NS	NS	2:all	small; present; NS
	Calanoida	#/NS N	#/s	3	relative	METHOD 91	METHOD 92	NS	present, % abundance	2:all	
	Al,C,Cc,Cg,Ch, Che,Eg,Eh,Hn,	#/75-NS MNH and.or 243-NS MNH	#/s	21	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
	Mpy,Pm,Smi,un.	identification	order, genus, species	24	relative	METHOD 94	METHOD 92	NS	present, % abundance; NS	2:all	
		age	copepodite #; nauplius	22	relative	METHOD 94	METHOD 92	NS	stage #; stage; present, % abundance; NS	2:all	
	Cyclopoi O,Ob,Om	#/75-NS MNH and.or 243-NS MNH	#/s	21	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
		identification	genus, species	21	relative	METHOD 93	METHOD 92	NS	present, % abundance; NS	2:all	
	Amphipoda	#/NS N	#/s	1	relative	METHOD 91	METHOD 92	NS	present, % abundance	2:all	
	Hg,Pmb,Pl,Plt	#/75-NS MNH and.or 243-NS MNH	#/s	15	relative	METHOD 93	METHOD 92	NS	present, % abundance	2:all	
		identification	species	16	relative	METHOD 94	METHOD 92	NS	present, % abundance; NS	2:all	
	Euphausiacea Ti	#/75-NS MNH and.or 243-NS MNH	#/s	2	relative	METHOD 93	METHOD 92	NS	present	2:all	
		identification	species	2	relative	METHOD 93	METHOD 92	NS	present; NS	2:all	
	Decapoda	#/NS N	#/s	1	relative	METHOD 91	METHOD 92	NS	present	2:all	
	(Brachyura)	identification	section	1	relative	METHOD 91	METHOD 92	NS	present; NS	2:all	
		age	larva	1	relative	METHOD 91	METHOD 92	NS	stage; present	2:all	
	Chaetognatha Eh,Sm	#/75-NS MNH and.or 243-NS MNH	#/s	15	relative	METHOD 93	METHOD 92	NS	present	2:all	
		identification	species	15	relative	METHOD 93	METHOD 92	NS	present; NS	2:all	
	Asteroidea un.	#/75-NS MNH and.or 243-NS MNH	#/s	1	relative	METHOD 93	METHOD 92	NS	present	2:all	
		identification	class	1	relative	METHOD 93	METHOD 92	NS	present; NS	2:all	
		age	post-larva	1	relative	METHOD 93	METHOD 92	NS	stage; present; NS	2:all	
	Copelata un.	#/75-NS MNH and.or 243-NS MNH	#/s	11	relative	METHOD 93	METHOD 92	NS	present	2:all	

METHOD 91: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed after and retrieved before Miller nets; initially deployed from ship's boom; later fastened with 2.5 cm polypropylene rope at bow, inboard towing wires on net frame shortened, net towed away from ship's side; sample removed from cod end without washing net down.

METHOD 92: when sufficient, sample split in Folsom Splitter; 1/8 preserved in 5% buffered F; rest stored in Whirl-pak bag or aluminum can and frozen.

METHOD 93: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed before neuston tows and recovered last; sample removed from cod end without washing net down.

METHOD 94: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; towing details given; sample removed from cod end without washing net down.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0003 (cont'd)		identification	class	11	relative	METHOD 93	METHOD 92	NS	present; NS	2:all	
	ICHTHYOPLANKTON										
	Other	#/NS N	#/s	2	relative	METHOD 91	METHOD 92	NS	present	2:all	
	un.	#/75-NS MNH and.or 243-NS MNH	#/s	5	relative	METHOD 93	METHOD 92	NS	present	2:all	
		age	relative	7	relative	METHOD 94	METHOD 92	NS	young; present	2:all	
77-0009	PHYTOPLANKTON										
	Centrales A,Cb,CH,CO,CY, M,Ma,Mi,R,S,ST	#/VD	#/ss	16	5402	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	genus, species	16	5402	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	
	Pennales A,AC,AM,As,Ca, CE,CM,CO,D,Ed, EP,EU,F,G,GY, L,N,Ngg,Nse, NT,PE,R,RH,ST, SU,SY,T,Tfe,Tl	#/VD	#/ss	16	81097	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	genus, species	16	81097	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	
	Chlorophyceae Pr?	#/VD	#/ss	1	1	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	species	1	1	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	
	Chrysophyceae C,CH,DN	#/VD	#/ss	10	1723	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	genus	10	1723	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	

METHOD 91: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed after and retrieved before Miller nets; initially deployed from ship's boom; later fastened with 2.5 cm polypropylene rope at bow, inboard towing wires on net frame shortened, net towed away from ship's side; sample removed from cod end without washing net down.

METHOD 92: when sufficient, sample split in Folsom Splitter; 1/8 preserved in 5% buffered F; rest stored in Whirl-pak bag or aluminum can and frozen.

METHOD 93: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; deployed before neuston tows and recovered last; sample removed from cod end without washing net down.

METHOD 94: S; S; NS; 2.57-3.09; NS; NS stn. depth; S/? date; towing details given; sample removed from cod end without washing net down.

METHOD 95: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for $\geq$3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif.

METHOD 96: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for $\geq$3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif., when required x1600 magnif. used.

METHOD 161: 1L preserved with 10mL Lugol's solution.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY COLLECTION	METHODOLOGY STORAGE	METHODOLOGY ANALYSIS	METHODOLOGY PRECISION/ACCURACY	RATING	REMARKS
77-0009 (cont'd)	Cyanophyceae un.	#/VD	#/ss	9	59	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	class	9	59	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	
	Dinophyceae C,D,un.	#/VD	#/ss	15	309	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1,2,4-6, 8	
		identification	class, genus	15	309	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:4-6,8	
	ZOOPLANKTON Protozoa un.	#/VD	#/ss	16	1322	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6, 8	
		identification	phylum	16	1322	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Mastigophora un.	#/VD	#/ss	8	99	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6, 8	
		identification	superclass	8	99	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Rhizopodea un.	#/VD	#/ss	16	1593	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6, 8	
		#/150-50 PNV	#/s	4	24	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:1-3,5,6, 8	
		identification - VD	class	16	1616	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
		identification - PNV	class	16	1616	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	

METHOD 95: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for ≥3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif.

METHOD 96: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for ≥3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif., when required x1600 magnif. used.

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

METHOD 161: 1L preserved with 10mL Lugol's solution.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)	Amoebida un.	#/VD	#/ss	1	1	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6,8	
		identification	order	1	1	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Radiolaria un.	#/VD	#/ss	5	24	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6,8	
		identification	subclass	5	24	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Ciliata un.	#/VD	#/ss	16	1525	S, 3L; S; NS; NS: ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6,8	
		identification	class	16	1525	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Tintinnida un.	#/VD	#/ss	16	1082	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 95	#	2:1-3,5,6,8	
		identification	order	16	1082	S, 3L; S; NS; NS; ? loc.; NS stn. depth; date rel.	METHOD 161	METHOD 96	NS	2:3,5,6,8	
	Anthomedusae Cf,L,Pk,Sp	#/150-50 PNV	#/s	14	96	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/s	12	40	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus, species	15	136	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	

METHOD 95: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for ≥3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif.

METHOD 96: sample vigourously shaken; 12.5mL aliquot transferred to counting cell; cell carefully closed to exclude bubbles; ss settled for ≥3h; examined with Leitz inverted compound microscope with phase contrast @ x320 magnif., when required x1600 magnif. used.

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

METHOD 161: 1L preserved with 10mL Lugol's solution.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)	Leptomedusae D	#/150-50 PNV	#/a	5	236	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/a	4	4	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus	6	240	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verificatin of taxonomy	2:3,5,6	
	Limnomedusae Gv	#/150-50 PNV	#/a	3	18	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/a	4	5	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	species	6	23	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	
	Trachymedusae A	#/150-50 PNV	#/a	16	791	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/a	14	5073	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus	16	5864	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	
	Narcomedusae Ac	#/150-50 PNV	#/a	12	225	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/a	1	5	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	species	12	230	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verificatin of taxonomy	2:3,5,6	
	Ctenophora un.	#/150-50 PNV	#/a	10	180	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)		#/571-100 PNH	#/s	3	56	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	phylum	11	234	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Sipuncula P	#/150-50 PNV	#/s	1	4	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus	1	4	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Thecosomata S,Sh	#/150-50 PNV	#/s	13	1475	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/s	10	109	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	order	14	1584	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of some molluscs	2:3,5,6,8	
	Gymnosomata C	#/150-50 PNV	#/s	13	422	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/s	9	106	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	order	15	528	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of some molluscs	2:3,5,6,8	
	Pelecypoda un.	#/150-50 PNV	#/s	8	442	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	class	8	442	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of some molluscs	2:3,5,6,8	
	Polychaeta (Spionidae),un.	#/150-50 PNV	#/s	16	82667	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)		#/571-100 PNH	#/m	1	24	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	class, family	16	02667	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
		age	trochophore	16	82632	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
	Diplostraca Np,P	#/150-50 PNV	#/m	3	28	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/m	1	9	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	order	4	37	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Copepoda un.	#/150-50 PNV	#/m	16	1078383	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/m	11	6117	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	subclass	16	1084500	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	
		age	nauplius, copepodid	16	1084500	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
	Calanoida C,Cf,Cg,Ch,ER, M,Ml,Mlu,Pm	#/150-50 PNV	#/m	16	67377	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/m	16	195063	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	genus, species	16	262440	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)	Harpacticoida un.	#/150-50 PNV	#/s	6	106	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	order	6	106	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6	
	Cyclopoida 0,0I	#/150-50 PNV	#/s	16	69808	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/s	2	168	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	genus	16	69976	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	external verification of taxonomy	2:3,5,6,9	
	Cirripedia un.	#/150-50 PNV	#/s	10	166	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/s	1	32	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	subclass	10	198	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
		age	nauplius	10	198	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
	Amphipoda H,O,OR, (Calliopidae, Lysianassidae)	#/150-50 PNV	#/s	10	101	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		#/571-100 PNH	#/s	5	18	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	
		identification	family, genus	12	119	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Euphausiacea E,un.	#/150-50 PNV	#/s	12	110	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6, 8	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)		#/571-100 PNH	#/s	1	8	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	order, genus	12	118	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
		age	furcilia	3	23	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
	Decapoda (Pandalidae)	#/150-50 PNV	#/s	1	1	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	family	1	1	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Insecta un.	#/571-100 PNH	#/s	8	620	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	class	8	620	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Bryozoa un.	#/150-50 PNV	#/s	1	2	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	phylum	1	2	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
		age	cyphonaut	1	2	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
	Chaetognatha 5	#/150-50 PNV	#/s	14	136	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/s	2	2	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus	14	138	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)	Asteroidea un.	#/150-50 PNV	#/m	1	32	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	class	1	32	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Ophiuroidea un.	#/150-50 PNV	#/m	1	16	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	class	1	16	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Copelata 0	#/150-50 PNV	#/m	8	795	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/m	4	11	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		identification	genus	10	806	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
	Other un.	biomass, total wet	NS	NS	NS	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 97	NS	2:1-3,5,8	
	ICHTHYOPLANKTON Other un.	#/571-100 PNH	#/m	2	4	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,8	
		age	larva	2	4	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,8	
	Pleuronectidae un.	#/150-50 PNV	#/m	1	1	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	
		#/571-100 PNH	#/m	1	2	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	#	2:1-3,5,6,8	

METHOD 97: sample strained through tared 100μ mesh; blotted dry; weighed on Sartorius 2255 balance.

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
77-0009 (cont'd)		identification	family	2	3	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	NS	2:3,5,6,8	
		age	larva	2	3	S; S; NS; NS; NS; ? loc.; NS stn. depth; date rel.	5% buffered F	METHOD 98	stage	2:3,5,6,8	
78-0002	ZOOPLANKTON Hydrozoa Hp	#/NS SNV	#/m^3	1	1.71	S; S; NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	1.71	S; S; NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	2	4.91	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Leptomedusae Hp	#/NS SNV	#/m^3	1	1.19	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	1.19	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Trachymedusae Ad	#/NS SNV	#/m^3	2	5.76	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	2	5.76	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	1	0.92	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Gastropoda un.	biomass, total dry	mg/m^3	1	0.64	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Thecosomata Sh	#/NS SNV	#/m^3	1	0.17	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	0.17	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	

METHOD 98: sample washed into 1L beaker; placed on white background; large organisms counted; ss (500-2000 individuals) taken with Folsom Splitter; ss placed in petri dish; large organisms examined @ x6.4 magnif., smaller @ x16 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
78-0002 (cont'd)	Gymnosomata Cl	#/NS SNV	#/m^3	2	0.34	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	2	0.34	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	1	0.32	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Calanoida Al,Cf,Lm,Ml, Mpy,Pm,Pp	#/NS SNV	#/m^3	3	571.55	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	3	571.55	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	3	87.24	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Mysidacea Mo	#/NS SNV	#/m^3	1	0.17	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	0.17	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	1	0.02	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Amphipoda Cd	#/NS SNV	#/m^3	1	0.14	S; S; NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	0.14	S; S; NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	1	0.25	S; S; NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Decapoda Pb	#/NS SNV	#/m^3	2	1.31	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	2	1.31	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
78-0002 (cont'd)		biomass, total dry	mg/m^3	2	0.56	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		age	larva	2	1.31	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	stage	2:all	
	Chaetognatha Se	#/NS SNV	$\#/m^3$	3	0.48	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	3	0.48	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	3	0.48	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	Copelata Ol	#/NS SNV	$\#/m^3$	2	0.74	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		identification	species	2	0.74	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	2	0.33	S; S; S/NS; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
	ICHTHYOPLANKTON Other un.	#/NS SNV	$\#/m^3$	1	0.34	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	#	2:all	
		biomass, total dry	mg/m^3	1	0.86	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
		age	larva	1	0.34	S; S; S; NS; NS; ? loc.; NS stn. depth	NS	NS	NS	2:all	
78-0018	ZOOPLANKTON Hydrozoa Hp	#/NS SNV	$\#/m^3$	1	7.38	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	7.38	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m^3	1	4.95	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
	Anthomedusae Sp	#/NS SNV	$\#/m^3$	1	0.24	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
78-0018 (cont'd)		identification	species	1	0.24	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
	Calanoida Al,Cf,Ml,Mpy, Pp,Pm,Smi	#/NS SNV	#/m³	1	65.96	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	65.96	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m³	1	8.01	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
	Amphipoda Hg	#/NS SNV	#/m³	1	0.48	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	0.48	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m³	1	0.19	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
	Chaetognatha Se	#/NS SNV	#/m³	1	0.71	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	
		identification	species	1	0.71	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
		biomass, total dry	mg/m³	1	0.61	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
	ICHTHYOPLANKTON Other un.	#/NS SNV	#/m³	1	0.24	S; S; NS; NS; NS; NS stn. depth	NS	NS	#	2:all	
		biomass, total dry	mg/m³	1	1.05	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
		age	larva	1	0.24	S; S; NS; NS; NS; NS stn. depth	NS	NS	NS	2:all	
79-0009	PHYTOPLANKTON Bacillariophyceae un.	#/NB	#/a	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	present in low numbers	2:1,3-6,8	
		identification	relative	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	other diatoms; present in low numbers; NS	2:3-6,8	
	Centrales Cd,Ez,Rm,Sc	#/NB	#/a	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	present, most abundant	2:1,3-6,8	
		identification	species	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	NS	2:3-6,8	
	Dinophyceae un.	#/NB	#/a	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	present in low numbers	2:1,3-6,8	

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
79-0009 (cont'd)		identification	relative	1	relative	S, 3 L; S; NS; NS; NS stn. depth	NS	NS	some dinoflagellates; present in low numbers; NS	2:3-6,8	
79-0026	EPONTIC ALGAE Bacillariophyceae un.	#/NS core	# cells/L: live; dead	2	7.75×10^4; 2.01×10^4	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,2,4-6, 8	
		identification	relative	2	9.76×10^4	METHOD 99	10% F	METHOD 100	diatoms; NS	2:4-6,8	
	Pennales At, Nfr	#/NS core	# cells/L	2	5.93×10^4	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,2,4-6, 8	
		identification	species	2	5.93×10^4	METHOD 99	10% F	METHOD 100	NS	2:4-6,8	
	Cyanophyceae un.	#/NS core	# filaments/L	1	0.26×10^4	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	#	2:1,2,4-6, 8	
		identification	relative	1	0.26×10^4	METHOD 99	10% F	METHOD 100	blue-green algae; NS	2:4-6,8	
80-0016	EPONTIC ALGAE Bacillariophyceae un.	#/block	# cells/L: live; dead	1	68.63×10^5; 275.72×10^4	METHOD 101	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,4-6,8	
		#/NS core	# cells/L: live; dead	2	56.53×10^5; 195.76×10^4	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,2,4-6, 8	
		identification	relative	3	1723.08×10^4	METHOD 102	10% F	METHOD 100	diatoms; NS	2:4-6,8	
	Centrales Cfr, Cs	#/NS core	# cells/L	1	40700	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,2,4-6, 8	
		identification	species	1	40700	METHOD 99	10% F	METHOD 100	NS	2:4-6,8	
	Pennales At, Dl, Ge, N, Ncl, Ncy, Ndr, Nfr, Nga, Nlg, Npl, NT, Ntr, Pq	#/block	# cells/L	1	441.42×10^4	METHOD 101	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,4-6,8	
		#/NS core	# cells/L	2	385.27×10^4	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	living and dead cells counted; dead cell defined	2:1,2,4-6, 8	

METHOD 99: NS, 7.5 cm; tb; duplicate cores collected; bottom 10cm of core melted; volume recorded.
METHOD 100: 20mL as cleaned; mounted; examined @ x1000 magnif.; photographs of important or interesting species retained.
METHOD 101: bottom of block cut by chainsaw; sectioned; bottom 10cm melted; volume recorded.
METHOD 102: duplicate cores collected and bottom of block cut by chainsaw; sectioned; bottom 10cm melted; volume recorded.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
80-0016 (cont'd)		identification	genus, species	3	826.69x 10^4	METHOD 102	10% F	METHOD 100	NS	2:4-6,8	
	Cyanophyceae un.	#/NS core	# filaments/L	1	2300	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	#	2:1,2,4-6, 8	
		identification	relative	1	2300	METHOD 99	10% F	METHOD 100	blue-green algae; NS	2:4-6,8	
	Other un.	#/block	# cells/L	1	100×10^4	METHOD 101	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	#	2:1,4-6,8	
		#/NS core	# cells/L	2	13.51x 10^4+	METHOD 99	10% F	10 or 50mL sedimented as examined with Zeiss phase-contrast microscope	#	2:1-2,4-6, 8	
		identification	relative	3	113.51x 10^4+	METHOD 102	10% F	METHOD 100	spores, flagellates; NS	2:4-6,8	
80-0025	ZOOPLANKTON										
	Anthomedusae Ef,Eh?,H,Hc, Mh?,Ro,Sp	identification	genus, species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 106	2:4-6,8	
		biomass, total wet	mg/m^3	10	6018.61+	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6	
	Leptomedusae O,Pl?,Pla?	identification	genus, species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 106	2:4-6,8	
		biomass, total wet	mg/m^3	10	18.61	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6, 8	
	Trachymedusae Ad	identification	species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	10	230.1	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6, 8	
		age	larva	2	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6,8	
	Narcomedusae Al	identification	species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	10	81.51	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6, 8	

METHOD 99: NS, 7.5 cm; tb; duplicate cores collected; bottom 10cm of core melted; volume recorded.
METHOD 100: 20mL as cleaned; mounted; examined @ x1000 magnif.; photographs of important or interesting species retained.
METHOD 101: bottom of block cut by chainsaw; sectioned; bottom 10cm melted; volume recorded.
METHOD 102: duplicate cores collected and bottom of block cut by chainsaw; sectioned; bottom 10cm melted; volume recorded.
METHOD 103: 500 MNH; S; S; 1.5; NS plus 240-50 PNV and 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows were always taken.
METHOD 104: samples sieved through 1.024mm mesh nylon screen; washed with water; examined under low-power binocular microscope.
METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.
METHOD 106: sampling limitations and biases discussed; ? = taxonomy not verified.
METHOD 107: ref. for limitations and biases given; mean and SD of 6 replicates given; balance accuracy = ±1mg.
METHOD 108: sampling limitations and biases discussed; taxonomy verified.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
80-0025 (cont'd)	Siphonophore un.	identification	order	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	1	0.58	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Ctenophora un.	identification	phylum	6	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	6	83.11	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
	Nematoda un.	identification	phylum	1	NS	METHOD 110	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	1	<1	METHOD 110	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Thecosomata S,Sh	identification	genus, species	4	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:3-6	
		biomass, total wet	mg/m^3	4	3.51+	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,3-6,8	
		age	veliger	2	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:all	
	Gymnosomata C	identification	genus	3	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	3	<1	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	veliger	3	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6,8	
	Pelecypoda un.	identification	class	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:all	
		biomass, total wet	mg/m^3	1	<1	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,3-6,8	
	Polychaeta un.	identification	class	8	NS	METHOD 112	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	8	0.24+	METHOD 112	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
		age	trochophore; relative	7	NS	METHOD 112	10% F	METHOD 104	stage; juvenile; METHOD 108	2:4-6,8	
	Phyllodocida (Aphroditidae)	identification	family	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	1	<1	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Oligochaeta un.	identification	family	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	

METHOD 103: 500 MNH; S; S; 1.5; NS plus 240-50 PNV and 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows were always taken.

METHOD 104: samples sieved through 1.024mm mesh nylon screen; washed with water; examined under low-power binocular microscope.

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.

METHOD 107: ref. for limitations and biases given; mean and SD of 6 replicates given; balance accuracy = ±1mg.

METHOD 108: sampling limitations and biases discussed; taxonomy verified.

METHOD 109: 500-61 PNV; S; S; 1: Inter Ocean Model 313 flow meter; 6 replicate tows taken.

METHOD 110: 240-50 PNV: S; S; 1: Inter Ocean Model 313 flow meter; 6 replicate tows taken.

METHOD 111: 500 MNH; S; S; 1.5; NS plus 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows taken.

METHOD 112: 240-50 PNV and 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows taken.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
80-0025 (cont'd)		biomass, total wet	mg/m^3	1	<1	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Crustacea un.	identification	order	1	NS	METHOD 109	10% F	METHOD 104	sampling limitations and biases discussed; mysid or euphausid	2:4-6,8	
		biomass, total wet	mg/m^3	1	1.13	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Myodocopa un.	identification	order	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	1	<1	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Calanoida A,Ac,Al,Cg,Ch, Dt,Eg,Lm,Ml, Pm,Sm,Smi, (Temoridae)	identification	family, genus, species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	10	1408.74	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
		age	nauplius	2	NS	METHOD 103	10% F	METHOD 104	stage; METHOD 108	2:4-6,8	
	Harpacticoida H,T	identification	genus	3	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	3	<1	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	copepodite	1	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6,8	
	Cyclopoida CY,Ob,Os	identification	genus	5	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	5	<1	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
	Cirripedia un.	identification	subclass	8	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	8	<1	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	nauplius, cyprid	8	NS	METHOD 111	10% F	METHOD 104	stage; METHOD 108	2:4-6,8	
	Mysidacea Ml	identification	species	6	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	6	461.26	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	

METHOD 103: 500 MNH; S; S; 1.5; NS plus 240-50 PNV and 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows were always taken.
METHOD 104: samples sieved through 1.024mm mesh nylon screen; washed with water; examined under low-power binocular microscope.
METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.
METHOD 107: ref. for limitations and biases given; mean and SD of 6 replicates given; balance accuracy = ±1mg.
METHOD 108: sampling limitations and biases discussed; taxonomy verified.
METHOD 109: 500-61 PNV; S; S; 1: Inter Ocean Model 313 flow meter; 6 replicate tows taken.
METHOD 111: 500 MNH; S; S; 1.5; NS plus 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows taken.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
80-0025 (cont'd)	Cumacea un.	identification	order	1	NS	METHOD 109	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	1	0.02	METHOD 109	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Isopoda un.	identification	order	3	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	3	<1	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
	Amphipoda Gw,H,Hg,On, Pab,Pl, (Hyperiidae, Gammaridae, Oedicerotidae)	identification	family, genus, species	10	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	10	12.66	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	
		age	relative	9	NS	METHOD 103	10% F	METHOD 104	juvenile; METHOD 108	2:4-6,8	
	Euphausiacea 1?	identification	genus	2	NS	METHOD 111	10% F	METHOD 104	METHOD 106	2:4-6	
		biomass, total wet	mg/m^3	2	0.38+	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	relative	1	NS	METHOD 111	10% F	METHOD 104	juvenile; METHOD 106	2:4-6-,8	
	Decapoda un.	identification	order	6	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	6	5.5	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	relative	6	NS	METHOD 111	10% F	METHOD 104	juvenile; METHOD 108	2:4-6,8	
	Chaetognatha Eh,Se	identification	order	10	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	10	66.04+	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
	Ophiuroidea un.	identification	class	4	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	4	0.01+	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	pluteus; relative	4	NS	METHOD 111	10% F	METHOD 104	stage; juvenile; METHOD 108	2:4-6,8	
	Copelata Fb,O	identification	class	6	NS	METHOD 103	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	6	42.99	METHOD 103	10% F	METHOD 105	METHOD 107	2:1,2,4-6,8	

METHOD 103: 500 MNH; S; S; 1.5; NS plus 240-50 PNV and 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows were always taken.
METHOD 104: samples sieved through 1.024mm mesh nylon screen; washed with water; examined under low-power binocular microscope.
METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.
METHOD 106: sampling limitations and biases discussed; ? = taxonomy not verified.
METHOD 107: ref. for limitations and biases given; mean and SD of 6 replicates given; balance accuracy = ±1mg.
METHOD 108: sampling limitations and biases discussed; taxonomy verified.
METHOD 109: 500-61 PNV; S; S; 1: Inter Ocean Model 313 flow meter; 6 replicate tows taken.
METHOD 111: 500 MNH; S; S; 1.5; NS plus 500-61 PNV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows taken.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
80-0025 (cont'd)	Other un.	identification	relative	7	NS	METHOD 111	10% F	METHOD 104	METHOD 108	2:4-6	
		biomass, total wet	mg/m^3	7	0.2	METHOD 111	10% F	METHOD 105	METHOD 107	2:1,4-6,8	
		age	larva	7	NS	METHOD 111	10% F	METHOD 104	stage; METHOD 108	2:4-6,8	
81-0003	BACTERIA un.	#/NBS	TC/L	3	645.87	S, 3.8 L; S; NS; NS; ? loc.; NS stn. depth	METHOD 113	METHOD 114	METHOD 115	2:1,2,4,5,8	
81-0015	PHYTOPLANKTON Centrales Cb,Ccm,Cd,Cf,Cfr,CH,CO,Ce,CY,Me,Pg,Tg,Tlc,un.	#/NB	# cells/L	4	2840893	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	order, genus, species	4	2840893	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Pennales Ae,AH,Akr,Aph,At,Cc,Dla,Ge,Ck,Ncc,Ncy,Ndn,Ndr,Nfr,Ngn,Nle,Nlg,Nn,Npl,Npr,Nr,NT,Ntr,Nva,Nvm,Pla,SU,un.	#/NB	# cells/L	4	3525383	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	order, genus, species	4	3525383	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	

METHOD 104: samples sieved through 1.024mm mesh nylon screen; washed with water; examined under low-power binocular microscope.

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.

METHOD 107: ref. for limitations and biases given; mean and SD of 6 replicates given; balance accuracy = ±1mg.

METHOD 108: sampling limitations and biases discussed; taxonomy verified.

METHOD 111: 500 MMH; S; S; 1.5; NS plus 500-61 PMV; S; S; 1; Inter Ocean Model 313 flow meter; 6 replicate tows taken.

METHOD 113: preserved in 0.2% gluteraldehyde in 20mL aliquots.

METHOD 114: processed within several hours of collection; method (ref. given) modified; polycarbonate membrane filter (Nucleopore) (0.2um pore size, 25mm diam.) prestained in 2% solution of irgalan black in 2% v/v acetic acid; 2-15mL as shaken on vortex mixer; added to filter funnel; 80% dye content acridine orange (concentration of 0.1% in 0.02 mol tris buffer - pH 7.2) added to sample to yield final stain concentration of 0.02%; after 2 min., sample filtered; rinsed with 5mL cell-free water; membrane placed on glass slide; wetted with drop of Cargille Type A immersion oil; covered with cover glass; examined with Zeiss WL microscope equipped with epifluorescent condenser, 50-watt mercury lamp, BG 12 excitation filter, No. 50 barrier filter, No. 500 beam splitter and reticule with 10mm grid.

METHOD 115: sufficient amount sample filtered to yield approx. 100 cells per grid field; 10 randomly selected grid fields of each sample membrane counted; mean value determined.

METHOD 116: preserved with acid Lugol's solution (ref. given) in glass bottles.

METHOD 117: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given).

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% on 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 119: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)	Chlorophyceae CA	#/NB	# cells/L	2	143335	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	genus	2	143335	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Chrysophyceae Db	#/NB	# cells/L	1	99960	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	species	1	99960	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Euglenophyceae Ep,un.	#/NB	# cells/L	4	20133	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	class, species	4	20133	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Dinophyceae A,GR,GY,P,Pd, un.	#/NB	# cells/L	4	362498	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	class, genus, species; relative	4	362498	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	cysts; METHOD 120	2:5	
	Craspedophyceae un.	#/NB	# cells/L	2	19589	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	class	2	19589	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Cryptophyceae un.	#/NB	# cells/L	1	7839	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	class	1	7839	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 120	2:5	
	Other un.	#/NB	# cells/L	4	2035086	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	relative	4	2035086	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 121	2:5	
	ZOOPLANKTON										
	Hydrozoa un.	#/150-22.5 PNV	#/m^3	2	10	S; S; S/S? NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	relative	2	10	S; S; S/S? NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	medusa; taxonomic references stated	2:4,5,8	
	Anthomedusae (Bougainvillidae), (Corynidae), Cb,Hy,Py	#/150-22.5 PNO	#/m^3	1	12	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	METHOD 123	2:1,2,4,5,8	
		#/150-22.5 PNV	#/m^3	2	18	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	

METHOD 116: preserved with acid Lugol's solution (ref. given) in glass bottles.

METHOD 117: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given).

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 119: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 121: silicoflagellates, microflagellates; algal spores, unidentified filaments; taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 123: duplicate hauls made; # calculated assuming net hauled vertically; precision not determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)		identification	family, species	2	30	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Narcomedusae AE	#/158-22.5 PNV	$\#/m^3$	1	1	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	genus	1	1	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Gastropoda un.	#/158-22.5 PNO	$\#/m^3$	1	668	S; S; S; NS; no	METHOD 122	METHOD 124	METHOD 125	2:1,2,4,5,8	
		identification	class	1	668	S; S; S; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
		age	larva	1	668	S; S; S; NS; no	METHOD 122	METHOD 124	stage; NS	2:4,5,8	
	Polychaeta un.	#/158-22.5 PNO	$\#/m^3$	1	7	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	METHOD 123	2:1,2,4,5,8	
		#/158-22.5 PNV	$\#/m^3$	2	8832	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	class	2	8839	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
		age	larva	2	8839	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	stage; NS	2:4,5,8	
	Copepoda un.	#/158-22.5 PNO	$\#/m^3$	1	148696	S; S; S; NS; no	METHOD 122	METHOD 124	METHOD 125	2:1,2,4,5,8	
		#/158-22.5 PNV	$\#/m^3$	2	6486	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	subclass	2	155182	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
		age	nauplius	2	155182	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	stage; NS	2:4,5,8	
	Calanoida C,ER,L,M,P	#/158-22.5 PNO	$\#/m^3$	1	78973	S; S; S; NS; no	METHOD 122	METHOD 124	METHOD 125	2:1,2,4,5,8	
		#/158-22.5 PNV	$\#/m^3$	2	77082	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	genus	2	156055	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 123: duplicate hauls made; # calculated assuming net hauled vertically; precision not determined.

METHOD 124: samples with large numbers of animals split - larger forms removed; sample split into ≥ 2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 125: duplicate hauls made; # calculated assuming net hauled vertically; when feasible, ≥ 250 organisms counted per sample; precision not determined.

METHOD 126: duplicate hauls made; when feasible, ≥ 250 organisms counted per sample; precision not determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)	Harpacticoida (Harpacticidae), H,un.	#/158-22.5 PNO	$\#/m^3$	1	92	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	METHOD 123	2:1,2,4,5,8	
		#/158-22.5 PNV	$\#/m^3$	2	658	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	order, family, genus	2	750	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
	Cyclopoida O,OI	#/158-22.5 PNV	$\#/m^3$	2	6252	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	genus	2	6252	S; S; S/S?/NS; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
	Mysidacea un.	#/158-22.5 PNV	$\#/m^3$	1	30	S; S; NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	order	1	30	S; S; NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Amphipoda A,Pp	#/158-22.5 PNV	$\#/m^3$	1	6	S; S; NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	genus, species	1	6	S; S; NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Chaetognatha S	#/158-22.5 PNV	$\#/m^3$	1	3	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	genus	1	3	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Ophiuroidea un.	#/158-22.5 PNO	$\#/m^3$	1	288	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	METHOD 123	2:1,2,4,5,8	
		identification	class	1	288	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	1	288	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	ICHTHYOPLANKTON Osteichthyes un.	#/158-22.5 PNV	$\#/m^3$	1	29	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class	1	29	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.
METHOD 123: duplicate hauls made; # calculated assuming net hauled vertically; precision not determined.
METHOD 124: samples with large numbers of animals split - larger forms removed; sample split into ≥ 2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.
METHOD 126: duplicate hauls made; when feasible, ≥ 250 organisms counted per sample; precision not determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)		age	larva	1	29	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Stichaidae un.	#/158-22.5 PNO	$\#/m^3$	1	1	S; S; S/S?/NS; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	METHOD 123	2:1,2,4,5,8	
		#/158-22.5 PNV	$\#/m^3$	1	77	S; S; S?; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	family	1	78	S; S; S/S?; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	1	78	S; S; S/S?; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	EPONTIC ALGAE Centrales Cb,Cce,Cd,Cfr,	#/S core	# cells/L	1	124976115	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
	Ck,CO,Cs,Ms, flc,un.	identification	order, genus, species	1	124976115	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Pennales Aco,Ae,Af,Ag, Akk,Akr,Ala,	#/S core	# cells/L	1	425116441	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
	All,AP, Apl, At,Cc,Ck,Dl, Dla,Dlc,Ge,N, Na,Nan,Nbb,Nc, Ncr,Ncy,Ndr, Nda,Ngn,Nhy, Nla,Nle,Nlg, Nme,Nn,Npl, Npo,Npy,Nr, Nre,Ns,Nsc, Nsi,Nsr,NT,Nt, Ntd,Nti,Ntr, Nva,Nvm,Pag, PL,Pq,Pqb,Pqc, Pae,Sq,un.	identification	order, genus, species	1	425116441	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 123: duplicate hauls made; # calculated assuming net hauled vertically; precision not determined.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 128: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS as taken; each as thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given)

METHOD 129: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS as taken; each as thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)	Chlorophyceae CA	#/S core	# cells/L	1	1200888	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	genus	1	1200888	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Dinophyceae P,un.	#/S core	# cells/L	1	444418	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	class, genus	1	444418	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Other un.	#/S core	# cells/L	1	6439981	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	relative	1	6439981	S; S; cores cut into different layers	METHOD 127	METHOD 129	microflagellates, algal spores?, unidentified filaments; METHOD 120	2:4,5	
	EPONTIC INVERTEBRATES										
	Nematoda un.	#/S core	#/L	1	2712	S; S; cores cut into different layers	METHOD 127	METHOD 130	METHOD 131	2:1,4,5,8	
		#/scraping	#/L	1	8	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	phylum	1	2712	S; S; cores cut into different layers	METHOD 127	METHOD 130	taxonomic references stated	2:4,5,8	
		identification - scraping	phylum	1	8	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Polychaeta (Spionidae),un.	#/S core	#/L	1	17	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 128: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS ss taken; each ss thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given)

METHOD 129: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS ss taken; each ss thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

METHOD 130: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples with large numbers of animals split - larger forms removed; sample split into >2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 131: when feasible, >250 organisms counted in each sample; precision not determined.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 10cm hole in ice; styrofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 133: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples examined with Wilde M8 stereo microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)		#/scraping	#/L	1	9	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	order, family	1	17	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	order, family	1	9	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age - core	larva	1	17	S; S; cores cut into different layers	METHOD 127	METHOD 133	stage; NS	2:4,5,8	
		age - scraping	larva	1	9	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Copepoda un.	#/scraping	#/L	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification	subclass	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	nauplius	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Calanoida ER,P	#/S core	#/L	1	4	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/L	1	762	METHOD 132	METHOD 122	METHOD 124	METHOD 131	2:1,2,4,5,8	
		identification - core	genus	1	4	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	genus	1	762	METHOD 132	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
	Harpacticoida (Ectinosomidae), H	#/S core	#/L	1	253	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/L	1	53	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	family, genus	1	253	S; S; cores cut into different layers	METHOD 127	METHOD 130	taxonomic references stated	2:4,5,8	
		identification - scraping	family, genus	1	53	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Amphipoda A,O,un.	#/scraping	#/L	1	40	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 124: samples with large numbers of animals split - larger forms removed; sample split into $\geq$ 2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 130: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples with large numbers of animals split - larger forms removed; sample split into $\geq$2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 131: when feasible, $\geq$250 organisms counted in each sample; precision not determined.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 30cm hole in ice; stryofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 133: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples examined with Wilde M8 stereo microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0015 (cont'd)		identification	order, genus	1	40	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
81-0027	ZOOPLANKTON Hydrozoa un.	identification	class	6	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	6	891.5479+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Anthomedusae B,Ef,Ho,Hp,Ro,S	identification	genus, species	7	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	7	780.1667+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Leptomedusae O	identification	genus	6	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	6	0.3584+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Trachymedusae Ad	identification	species	6	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	6	135.7275	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Narcomedusae Al	identification	species	6	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed NS	2:4-6,8	
		biomass, total wet	mg/m^3	6	18.4261	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Siphonophora Da	identification	species	1	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed NS	2:4-6,8	
		biomass, total wet	mg/m^3	1	0.2997	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Ctenophora un.	identification	phylum	11	$\leq$100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed NS	2:4-6,8	
		biomass, total wet	mg/m^3	11	331.1044	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 30cm hole in ice; stryofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 134: S; S, WA; S; 1-1.5; torpedo-style flow meters; closing bridle; General Oceanics tripping mechanism; 3 replicate tows taken.

METHOD 135: samples sieved through 0.569mm mesh nylon screen; NS as taken with Hensen-Stemple pipette when >100 of any group were present; no more than 100 individuals of any group processed.

METHOD 163: mean and SD of 3 replicates given; balance accuracy: ±1mg.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0027 (cont'd)	Beroida Bc	identification	species	1	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed NS	2:4-6,8	
		biomass, total wet	mg/m^3	1	8.939	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Gastropoda un.	identification	class; relative	9	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS; pteropoda	2:3-6,8	
		biomass, total wet	mg/m^3	9	7.129+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,3-6	
	Thecosomata Sh	identification	species	5	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:3-6,8	
		biomass, total wet	mg/m^3	5	13.3044	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,3-6	
	Gymnosomata Cl	identification	species	5	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	5	1.971	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Diplostraca (Cladocera), P	identification	suborder, genus	10	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	10	1.8199	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Ostracoda un.	identification	subclass	3	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	3	0.1621+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Calanoida Ac,Al,Cg,Ch, Dt,Eg,ER,Lm, Pm,SP,un.	identification	order, genus, species	12	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	12	5753.7202	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Cirripedia un.	identification	subclass	5	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	5	NS	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Mysidacea Ml,Mr	identification	species	8	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.
METHOD 134: S; S, WA; S; 1-1.5; torpedo-style flow meters; closing bridle; General Oceanics tripping mechanism; 3 replicate tows taken.
METHOD 135: samples sieved through 0.569mm mesh nylon screen; NS as taken with Hensen-Stemple pipette when >100 of any group were present; no more than 100 individuals of any group processed.
METHOD 163: mean and SD of 3 replicates given; balance accuracy: ±1mg.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0027 (cont'd)		biomass, total wet	mg/m^3	8	417.9839	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Isopoda un.	identification	order	2	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	2	NS	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Amphipoda (Gammaridae), Ag,Ga,H,Hg,Og, On,Pab,Pl	identification	family, genus, species	10	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	10	220.6889	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
		age	relative	5	NS	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
	Euphausiacea T	identification	genus	8	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	8	22.6119	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Decapoda un.	identification	order	10	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	10	14.6449+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Chaetognatha Se	identification	species	11	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	11	71.6082	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Asteroidea un.	identification	class	2	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	2	0.0139+	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
	Copelata Ov,un.	identification	class, species	6	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
		biomass, total wet	mg/m^3	6	505.5952	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.
METHOD 134: S; S, WA; S; 1-1.5; torpedo-style flow meters; closing bridle; General Oceanics tripping mechanism; 3 replicate tows taken.
METHOD 135: samples sieved through 0.569mm mesh nylon screen; NS as taken with Hensen-Stemple pipette when >100 of any group were present; no more than 100 individuals of any group processed.
METHOD 163: mean and SD of 3 replicates given; balance accuracy: ±1mg.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
81-0027 (cont'd)	Other un.	identification	relative	3	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS; benthic larvae	2:4-6,8	
		biomass, total wet	mg/m^3	3	0.7723	METHOD 134	10% F	METHOD 105	METHOD 163	2:1,4-6	
		age	larva	3	≤100	METHOD 134	10% F	METHOD 135	sampling limitations and biases discussed; NS	2:4-6,8	
81-0101	BACTERIA un.	#/NBS	TC/L; CFU/L	3	505.23; 38.75	S, 3.8 L; S; NS; NS; ? loc.; NS stn. depth	TC: METHOD 113 CFU: METHOD 136	TC: METHOD 114 CFU: METHOD 138	TC: METHOD 139 CFU: METHOD 140	2:1,4,5,8	
82-0032	ICHTHYOPLANKTON PCHR	#/750-50 WNH	#/m	11	107	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	11	107	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	11	107	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	11	107	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	
	RNSM	#/750-50 WNH	#/m	13	1530	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	13	1530	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	13	relative	METHOD 141	METHOD 142	METHOD 143	up to 10/day measured; NS	2:3-5,8	
		age	larva	13	1530	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	

METHOD 105: each species grouped together; wet weight determined on a Mettler PL1200 electronic balance.

METHOD 113: preserved in 0.2% gluteraldehyde in 20mL aliquots.

METHOD 114: processed within several hours of collection; method (ref. given) modified; polycarbonate membrane filter (Nucleopore) (0.2um pore size, 25mm diam.) prestained in 2% solution of irgalan black in 2% v/v acetic acid; 2-15mL ss shaken on vortex mixer; added to filter funnel; 80% dye content acridine orange (concentration of 0.1% in 0.02 mol tris buffer - pH 7.2) added to sample to yield final stain concentration of 0.02%; after 2 min., sample filtered; rinsed with 5mL cell-free water; membrane placed on glass slide; wetted with drop of Cargille Type A immersion oil; covered with cover glass; examined with Zeiss WL microscope equipped with epifluorescent condenser, 50-watt mercury lamp, BG 12 excitation filter, No. 50 barrier filter, No. 500 beam splitter and reticule with 10mm grid.

METHOD 134: S; S, WA; S; 1-1.5; torpedo-style flow meters; closing bridle; General Oceanics tripping mechanism; 3 replicate tows taken.

METHOD 135: samples sieved through 0.569mm mesh nylon screen; NS ss taken with Hensen-Stemple pipette when >100 of any group were present; no more than 100 individuals of any group processed.

METHOD 136: spin-plate technique used to dispense 0.1mL ss with 0.2mL sterile pipette onto surface of cold Lib X agar plate [prepared previously (ref. given) with 1L of 34% Instant Ocean; autoclaved; cooled; dispensed in 100mm petri dishes; stored at 2°C]; incubated at 5°C.

METHOD 138: incubation continued for 3 weeks; plates examined; mean value of replicate set determined.

METHOD 139: sufficient amount sample filtered to yield approx. 100 cells per grid field; 10 randomly selected grid fields of each sample membrane counted; mean value determined.

METHOD 140: triplicate spin-plates made of each sample; plates with uneven distribution of colonies discarded; colonies of a replicate set enumerated; #.

METHOD 141: S; S, WA; NS; 1; centre-mounted General Oceanics digital flow meter Model 2030 (not used during 1 towing period); date rel.; 5.5kg depressor on 1m line attached to tow rope 1m in front of net; 2 replicate tows generally performed.

METHOD 142: weak non-buffered F; following analysis stored in 15mL dark glass bottles with 5% non-buffered F.

METHOD 143: ≤18mm: measured with ocular micrometer to 0.1mm; >18mm: measured with ruler to nearest 0.5mm.

METHOD 163: mean and SD of 3 replicates given; balance accuracy: ±1mg.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0032 (cont'd)	ARCD	#/750-50 WNH	#/m	3	6	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	3	6	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	3	6	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	3	6	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	
	SFCD	#/750-50 WNH	#/m	3	37	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	3	37	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	3	37	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	3	37	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	
	LUMP	#/750-50 WNH	#/m	2	8	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	2	8	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	2	8	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	2	8	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	
	FHSC	#/750-50 WNH	#/m	4	9	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	4	9	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	4	9	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	4	9	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	
	STFL	#/750-50 WNH	#/m	2	21	METHOD 141	METHOD 142	NS	#	2:1-5,8	
		identification	species	2	21	METHOD 141	METHOD 142	NS	taxonomic references stated, illustrations included	2:3-5,8	
		length, overall	mm	2	21	METHOD 141	METHOD 142	METHOD 143	NS	2:3-5,8	
		age	larva	2	21	METHOD 141	METHOD 142	terminology described	stage	2:3-5,8	

METHOD 141: S; S, WA; NS; 1; centre-mounted General Oceanics digital flow meter Model 2030 (not used during 1 towing period); date rel.; 5.5kg depressor on 1m line attached to tow rope 1m in front of net; 2 replicate tows generally performed.

METHOD 142: weak non-buffered F; following analysis stored in 15mL dark glass bottles with 5% non-buffered F.

METHOD 143: $\leq$10mm: measured with ocular micrometer to 0.1mm; >10mm: measured with ruler to nearest 0.5mm.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0109	BACTERIA un.	#/NBS	TC/L	3	110.7	S, 3.8 L; S; NS; NS; ? loc.; NS stn. depth	METHOD 113	METHOD 114	METHOD 139	2:1,4,5,8	
82-0110	PHYTOPLANKTON Centrales Cb,Ccm,CH,Cks, Cs,Ms,T,un.	#/NB identification	# cells/L order, genus, species	3 3	49970 49970	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Pennales AH,At,Cc,Ge, Nan,Ncy,Ndr, Nds,Nfr,Ngn, Npl,Nse,NT, Psg,Pq,un.	#/NB identification	# cells/L order, genus, species	4 4	867900 867900	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Chlorophyceae CA	#/NB identification	# cells/L genus	3 3	38135 38135	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Euglenophyceae E	#/NB identification	# cells/L genus	1 1	1315 1315	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Dinophyceae Da,P,un.	#/NB identification	# cells/L class, genus, species	4 4	76270 76270	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Craspedophyceae un.	#/NB identification	# cells/L class	4 4	26300 26300	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	
	Cryptophyceae un.	#/NB identification	# cells/L class	1 1	5260 5260	S, 3 L; S; S/NS; NS S, 3 L; S; S/NS; NS	METHOD 116 METHOD 116	METHOD 117 METHOD 119	METHOD 118 METHOD 120	2:1,5 2:5	

METHOD 113: preserved in 0.2% gluteraldehyde in 20ml aliquots.

METHOD 114: processed within several hours of collection; method (ref. given) modified; polycarbonate membrane filter (Nucleopore) (0.2um pore size, 25mm diam.) prestained in 2% solution of irgalan black in 2% v/v acetic acid; 2-15mL ss shaken on vortex mixer; added to filter funnel; 80% dye content acridine orange (concentration of 0.1% in 0.02 mol tris buffer - pH 7.2) added to sample to yield final stain concentration of 0.02%; after 2 min., sample filtered; rinsed with 5mL cell-free water; membrane placed on glass slide; wetted with drop of Cargille Type A immersion oil; covered with cover glass; examined with Zeiss WL microscope equipped with epifluorescent condenser, 50-watt mercury lamp, BG 12 excitation filter, No. 50 barrier filter, No. 500 beam splitter and reticule with 10mm grid.

METHOD 116: preserved with acid Lugol's solution (ref. given) in glass bottles.

METHOD 117: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given).

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 119: each sample thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 139: sufficient amount sample filtered to yield approx. 100 cells per grid field; 10 randomly selected grid fields of each sample membrane counted; mean value determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)	Other un.	#/NB	# cells/L	4	528630	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 117	METHOD 118	2:1,5	
		identification	relative	1	528630	S, 3 L; S; S/NS; NS	METHOD 116	METHOD 119	METHOD 121	2:5	
	ZOOPLANKTON Hydrozoa un.	#/158-22.5 PNV	#/m^3	1	16	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	order	1	16	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	relative	1	16	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	juvenile; NS	2:4,5,8	
	Anthomedusae (Bougainvillidae), Cb,Py	#/158-22.5 PNV	#/m^3	3	45	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	family, species	3	45	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Trachymedusae Ad	#/158-22.5 PNV	#/m^3	3	21	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	species	3	21	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Narcomedusae Ae,Al	#/158-22.5 PNV	#/m^3	2	33	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	genus, species	2	33	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Gastropoda un.	#/158-22.5 PNV	#/m^3	2	53	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class	2	53	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	2	53	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	

METHOD 116: preserved with acid Lugol's solution (ref. given) in glass bottles.

METHOD 117: each sample thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given).

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 119: each sample thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

METHOD 121: silicoflagellates, microflagellates; algal spores, unidentified filaments; taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)	Polychaeta (Nereidae),un.	#/150-22.5 PNV	#/m^3	3	170	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class, family	3	170	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	3	170	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Copepoda un.	#/150-22.5 PNV	#/m^3	3	530	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	subclass	3	530	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	nauplius	3	530	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Calanoida C,Ch,I,M,P	#/150-22.5 PNV	#/m^3	3	22314	S; S; S; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	genus, species	3	22314	S; S; S; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
	Harpacticoida un.	#/150-22.5 PNV	#/m^3	2	8	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	order	2	8	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Cyclopoida O,OI	#/150-22.5 PNV	#/m^3	3	3586	S; S; S; NS; no	METHOD 122	METHOD 124	METHOD 126	2:1,2,4,5,8	
		identification	genus	3	3586	S; S; S; NS; no	METHOD 122	METHOD 124	taxonomic references stated	2:4,5,8	
	Isopoda un.	#/150-22.5 PNV	#/m^3	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	order	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Amphipoda un.	#/150-22.5 PNV	#/m^3	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	order	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 124: samples with large numbers of animals split - larger forms removed; sample split into ≥ 2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 126: duplicate hauls made; when feasible, ≥ 250 organisms counted per sample; precision not determined.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)	Chaetognatha Eh	#/150-22.5 PNV	#/m^3	1	5	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	species	1	5	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Ophiuroidea un.	#/150-22.5 PNV	#/m^3	1	3	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class	1	3	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	1	3	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Copelata un.	#/150-22.5 PNV	#/m^3	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	ICHTHYOPLANKTON Osteichthyes un.	#/150-22.5 PNV	#/m^3	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	duplicate hauls made; precision not determined	2:1,2,4,5,8	
		identification	class	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	larva	1	2	S; S; S; NS; no	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	EPONTIC ALGAE Centrales	#/S core	# cells/L	5	220624609	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	order, genus, species	5	220624509	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Pennales	#/S core	# cells/L	5	30284747-46	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/L; intercomparison between two enumerators indicated little difference.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 128: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS ss taken; each ss thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given)

METHOD 129: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS ss taken; each ss thoroughly mixed; poured into Zeiss 50mL sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)		identification	order, genus, species	5	30286767-46	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Chlorophyceae CA	#/S core	# cells/l	2	478287	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	genus	2	478287	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Chrysophyceae Db	#/S core	# cells/L	3	127537	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	species	3	127537	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Euglenophyceae E	#/S core	# cells/L	3	565382	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	genus	3	565382	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Dinophyceae Da,GY,P,Pd	#/S core	# cells/L	5	9823166	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	genus, species	5	9823166	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Craspedophyceae un.	#/S core	# cells/L	4	1137334	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	class	4	1137334	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	
	Other un.	#/S core	# cells/L	5	112377926	S; S; cores cut into different layers	METHOD 127	METHOD 128	METHOD 118	2:1,4,5	
		identification	relative	8	112377926	S; S; cores cut into different layers	METHOD 127	METHOD 129	METHOD 120	2:4,5	

METHOD 118: counted in area equivalent to 89 microscopic fields; each cell counted as individual whether solitary or part of chain or colony; general precision estimate of ±20% as 40-100 cells counted per sample (ref. given); precision poor for samples with <50000 cells/l; intercomparison between two enumerators indicated little difference.

METHOD 120: taxonomic references stated; intercomparison between two identifiers indicated little difference.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 128: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS as taken; each as thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given)

METHOD 129: thawed at room temperature in darkness; preserved with acid Lugol's solution (ref. given) in glass bottles; NS as taken; each as thoroughly mixed; poured into Zeiss 50ml sedimentation chambers; settled for up to 48h; examined with Zeiss phase-contrast inverted microscope @ up to x625 magnif. following Utermohl technique (ref. given); permanent slides prepared (ref. for method description given); examined @ up to x1250 magnif.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)	EPONTIC INVERTEBRATES										
	Tintinnida	#/S core	#/ℓ	1	relative	S; S; cores cut into different layers	METHOD 127	METHOD 133	present; precision not determined	2:1,4,5,8	
		identification	order	1	relative	S; S; cores cut into different layers	METHOD 127	METHOD 133	present; taxonomic references stated	2:4,5,8	
	Hydrozoa un.	#/scraping	#/ℓ	1	1	METHOD 132	METHOD 127	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification	class	1	1	METHOD 132	METHOD 127	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Turbellaria un.	#/S core	#/ℓ	1	364	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/ℓ	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	class	1	364	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	class	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age - core	post-larva	1	364	S; S; cores cut into different layers	METHOD 127	METHOD 133	stage; NS	2:4,5,8	
		age - scraping	post-larva	1	3	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Nematoda un.	#/S core	#/ℓ	3	15678	S; S; cores cut into different layers	METHOD 127	METHOD 130	METHOD 131	2:1,4,5,8	
		#/scraping	#/ℓ	1	1	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	phylum	3	15678	S; S; cores cut into different layers	METHOD 127	METHOD 130	taxonomic references stated	2:4,5,8	
		identification - scraping	phylum	1	1	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Polychaeta (Spionidae),un.	#/S core	#/ℓ	3	524	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/ℓ	2	2	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 130: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples with large numbers of animals split - larger forms removed; sample split into ≥ 2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 131: when feasible, ≥ 250 organisms counted in each sample; precision not determined.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 30cm hole in ice; styrofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 133: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples examined with Wilde M8 stereo microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)		identification - core	class, family	3	524	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	class, family	2	2	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age - core	larva	3	524	S; S; cores cut into different layers	METHOD 127	METHOD 133	stage; NS	2:4,5,8	
		age - scraping	larva	2	2	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Copepoda un.	#/S core	#/l	1	190	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/l	1	5	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	subclass	1	190	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	subclass	1	5	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age - core	nauplius	1	190	S; S; cores cut into different layers	METHOD 127	METHOD 133	stage; NS	2:4,5,8	
		age - scraping	nauplius	1	5	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Calanoida CV,P,un.	#/S core	#/l	3	1089	S; S; cores cut into different layers	METHOD 127	METHOD 130	METHOD 131	2:1,4,5,8	
		#/scraping	#/l	2	12	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	order, genus	3	1089	S; S; cores cut into different layers	METHOD 127	METHOD 130	taxonomic references stated	2:4,5,8	
		identification - scraping	order, genus	2	12	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
		age	copepodite	1	5	METHOD 132	METHOD 127	examined with Wilde M8 stereo microscope	stage; NS	2:4,5,8	
	Harpacticoida (Ectinosomidae)	#/S core	#/l	4	675	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/l	1	8	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 130: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples with large numbers of animals split - larger forms removed; sample split into $\geq$2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 131: when feasible, $\geq$250 organisms counted in each sample; precision not determined.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 30cm hole in ice; stryofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 133: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples examined with Wilde M8 stereo microscope.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
82-0110 (cont'd)		identification - core	family, genus	4	673	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	family, genus	1	8	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Amphipoda A,G1	#/S core	#/ℓ	1	6	S; S; cores cut into different layers	METHOD 127	METHOD 133	precision not determined	2:1,4,5,8	
		#/scraping	#/ℓ	1	2	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	precision not determined	2:1,2,4,5,8	
		identification - core	genus, species	1	6	S; S; cores cut into different layers	METHOD 127	METHOD 133	taxonomic references stated	2:4,5,8	
		identification - scraping	genus, species	1	2	METHOD 132	METHOD 122	examined with Wilde M8 stereo microscope	taxonomic references stated	2:4,5,8	
	Other un.	#/S core	#/ℓ	1	2405	S; S; cores cut into different layers	METHOD 127	METHOD 130	METHOD 131	2:1,4,5,8	
		identification	relative	1	2405	S; S; cores cut into different layers	METHOD 122	METHOD 130	METHOD 144	2:4,5,8	
		age	trochophore, larva	1	<2405	S; S; cores cut into different layers	METHOD 127	METHOD 130	stage, NS	2:4,5,8	
83-0059	BACTERIA un.	#/VO	# × $10^{11}/m^3$	13	relative	METHOD 145	METHOD 146	METHOD 147	METHOD 148	2:1-3,5,8	
		length, overall	um	NS	NS	METHOD 145	METHOD 146	METHOD 149	mean dimensions determined	2:3-5,8	
		width, overall	um	NS	NS	METHOD 145	METHOD 146	METHOD 149	mean dimensions determined	2:3-5,8	

METHOD 122: preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic or glass bottles.

METHOD 127: each sample stored frozen in polyethylene bags or nalgene jars.

METHOD 130: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples with large numbers of animals split - larger forms removed; sample split into $\geq$2 fractions with Folsom Splitter; examined with Wilde M8 stereo microscope.

METHOD 131: when feasible, $\geq$250 organisms counted in each sample; precision not determined.

METHOD 132: ice scraper (25cm diam. plastic funnel with plankton net attached to hinged aluminum rod) pushed down a 30cm hole in ice; styrofoam attached to scraper gave positive buoyancy to force apparatus against under-ice surface; each sample consisted of 2 revolutions of scraper around perimeter of hole; 2 samples usually taken.

METHOD 133: core samples thawed at room temperature in darkness and preserved with 10% F (buffered with $Na_2B_4O_7$) in plastic and glass bottles; all samples examined with Wilde M8 stereo microscope.

METHOD 144: turbellarian larvae and pelecypod trochophore larvae and rotifers; taxonomic references stated.

METHOD 145: S; S, metre block; NS; NS; ? loc.; NS stn. depth; date rel.

METHOD 146: duplicate 10ml ss preserved in 3.7% F; stored at room temp.

METHOD 147: assayed within 3 weeks; each ss treated with 0.001M $Na_4P_4O_7$ for 30 min.; sonicated for 15 sec. at probe intensity of 100W; filtered onto 0.20um pore-size Nucleopore filter; examined by epifluorescent microscopy using DAPI technique (ref. for method description given).

METHOD 148: analytical method evenly disperses bacteria on filter; results graphed.

METHOD 149: cells stained with acridine orange (ref. for method description given); examined using epifluorescent microscopy.

TABLE 2 (cont'd)

DATA SET I.D.	TAXA	MEASUREMENT	UNITS	NO. STNS.	NO. SAMPLES	METHODOLOGY				RATING	REMARKS
						COLLECTION	STORAGE	ANALYSIS	PRECISION/ACCURACY		
84-0033	BACTERIA un.	#/bucket	#/ml	3	3179005	S; S; NS; NS	METHOD 150	METHOD 151	#	2:1,2,4,5,8	
	PHYTOPLANKTON Centrales CH	identification	species	1	NS	S; S; NS; NS	Lugol's iodine solution	METHOD 152	METHOD 153	2:5,8	
		biovolume	um^3/ml	1	714554	S; S; NS; NS	Lugol's iodine solution	METHOD 154	NS	2:4,5,8	
85-0016	BACTERIA un.	#/VD	# x $10^{11}/m^3$	4	relative	METHOD 145	METHOD 146	METHOD 147	METHOD 148	2:1-3,5,8	
		#/Craib core	# x 10^8/g dry weight	4	relative	S; NS; NS; ? loc.; date rel.	METHOD 155	METHOD 156	METHOD 148	2:1-3,5,8	
		length, overall	um	NS	NS	METHOD 145	METHOD 146	METHOD 149	mean dimensions determined	2:3-5,8	
		width, overall	um	NS	NS	METHOD 145	METHOD 146	METHOD 149	mean dimensions determined	2:3-5,8	

METHOD 145: S; S, metre block; NS; NS; ? loc.; NS stn. depth; date rel.

METHOD 146: duplicate 10ml ss preserved in 3.7% F; stored at room temp.

METHOD 147: assayed within 3 weeks; each ss treated with 0.001M $Na_4P_4O_7$ for 30 min.; sonicated for 15 sec. at probe intensity of 100W; filtered onto 0.20um pore-size Nuclepore filter; examined by epifluorescent microscopy using DAPI technique (ref. for method description given).

METHOD 148: analytical method evenly disperses bacteria on filter; results graphed.

METHOD 149: cells stained with acridine orange (ref. for method description given); examined using epifluorescent microscopy.

METHOD 150: preserved with 2% F in sterile glass bottles; kept at 4°C.

METHOD 151: processed within 6 weeks; stained with DAPI (ref. for method description given); examined with epifluorescence microscope.

METHOD 152: examined using Utermohl technique and Wilde M40 inverted microscope @ x10, x40 and x100 magnif.

METHOD 153: NS; any CH cultures were viable so unclear whether other species present.

METHOD 154: examined using Utermohl technique and Wilde M40 inverted microscope; at least 100 individuals counted @ x10, x40 and x100 magnif.; biovolume calculated from average cell dimensions by approximations to appropriate geometrical figures.

METHOD 155: surface of undisturbed sediment in each core sampled to 0.5cm depth by aspirating 15ml sediment into needleless 30ml plastic syringe; preserved in 2% F.

METHOD 156: assayed within 3 weeks; 100ul of each well-mixed slurry placed in 5ml sterile bacteria-free water; treated with 0.02M $Na_4P_4O_7$ for 30 min.; sonicated for 30 sec. at probe intensity of 100W; filtered onto 0.20um pore-size Nuclepore filter; examined by epifluorescent microscopy using DAPI technique (ref. for method description given).

8.3 Table 3: Sampling Effort

Table 3 describes the sampling effort for each data set. Data sets are listed in chronological order according to I.D. number. Data sets for which no information or only limited information is available on specific sampling parameters (e.g., time, area sampled) are listed, but the level of detail provided reflects this lack of information. The information on each data set and the abbreviations used are clarified below by table column.

If several types of gear were used at a station on a single date, they are separated under gear type with a semi-colon. Other sampling parameters (hour, interval, no. samples and depth sampled) can be matched with the gear type as they are listed in the same sequence as the gear type and are also separated with a semi-colon. Where only one notation appears in these columns, the information applies to all the gear types.

Data Set I.D.

Each identification (I.D.) number is unique to a data set; all subsequent references to data sets are by I.D. numbers. The first two digits of the I.D. refer to the last two digits of the year during which the data were gathered; the last four digits of the I.D. are assigned to the data set.

Station Number/Location

The number assigned to the station by the collecting agency is listed in some cases; where the precise geographic location was not indicated, a general description of the location is provided.

Latitude (°N) and Longitude (°W)

The specific north and west co-ordinates for each station are indicated in these columns. For data sets where transects were run, the co-ordinates for the start and finish of each transect are given. Not Specified ("NS") is used if this information was not provided in the documentation for the data set. Where sampling co-ordinates were not detailed but the stations were plotted on maps, approximate positions were obtained by measuring the plotted locations and are indicated in Table 3 by a "?".

Station Depth

This column indicates the water depth in metres at the samplng station. Not Specified ("NS") is used if this information was not reported. If the depth was estimated from concurrent studies, it is noted with a "?".

Gear Type

The type of gear used to collect the sample is indicated in this column, often as a coded abbrevation. An explanation of the codes is presented at the end of this section, following Remarks. If a combination of gear was used during the study and each type could not be listed on an individual basis, the gear types are listed in combination using either "and" or "and.or."

Time Sampled - Date, Time, Interval

The sampling date is indicated to the level of detail provided in the documentation for the data set. Not Specified ("NS") is used if this information was not reported. If the date was estimated from concurrent studies, it is noted with a "?".

The sampling time is expressed as Greenwich Mean Time (GMT). Where local or other time frames were used, they were converted to GMT for incorporation into Table 3.

The duration of the sampling period is indicated in minutes in the interval column.

No. Samples

The number of samples collected by each type of gear is listed. In some cases, the number was described in the documentation only in relative terms, e.g. several. Not Specified ("NS") is used if this information was not reported.

Depth Sampled

The depth in the water column sampled by the gear is expressed in metres either as a single number (for water bottles and most horizontal net tows) or as a range (for some horizontal net tows and all oblique and vertical net tows). The range "B-0" means that the gear was towed from near-Bottom to the surface (0m). Not Specified ("NS") is used if this information was not reported. Not Applicable ("NA") appears for ice and sediment samplers. If several depths were sampled by the same gear type on a single date, these depths are listed and separated by commas.

Remarks

Explanatory notes and additional information are presented in this column.

Description of Gear Type Codes

BN - Bongo Net
BRN - BiRge closing Net
C-23 PN - Coarse-silk conical Plankton Net
CB - Clarke Bumpus net
F-23PN - Finer silk conical Plankton Net
FN - Faber Net
IKT - Isaacs-Kidd midwater Trawl
KB - Kemmerer Bottle
MN - Miller Net
N - Neuston net
NB - Niskin Bottle
NBS - Niskin Bag-Sampler
NN - Nansen-type Net
NS - Not Specified
PB - Plastic Bottle
PN - conical Plankton Net
PP - PhytoPlankton net
PVC - PVC sampler
S corer - Sipre ice corer
S-30 PN - Silk conical Plankton Net
SN - Scor Net
STN - STramin Net
Subsampler - nuclear submarine sampler
VD - Van Dorn sampler
WN - Wisconsin Net

Note: In general, numerical prefixes to the codes indicate mesh size (u) and mouth diameter (cm) of the plankton net. The suffixes H, O and V indicate Horizontal, Oblique and Vertical tows, respectively. Further details are provided in Appendix I.

TABLE 3: SAMPLING EFFORT

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
13-0001	29e	70°03'	141°00'	46	C-23 PNV	Apr. 02, 1914	NS	NS	3	37-0	Hauls through crack in ice.
	29g	70°20'	140°30'	275	S-30 PNV	Apr. 06, 1914	NS	NS	Several	92-0, 275-0, 275-92	Long. reported by Mann (1925) as 140°20'W but by other authors as 140°30'W. Hauls through cracks in ice.
	36 - Off Cape Lyon, Darnley Bay, N.W.T.	NS	NS	9	F-23 PNH	Aug. 23, 1914	NS	5	1	0	
	43e - Off Stapylton Bay, Dolphin and Union Strait N.W.T.	NS	NS	46	C-23 PNH	Sep. 14, 1915	NS	10	1	0	
	50b	68°55'	116°52'	18	C-23 PNH	July 17, 1916	NS	NS	1	0	
50-0001	33	70°19'	138°24'	NS	50 NNV	Aug. 19	0700	NS	1	91-0	
	34	70°36'	135°50'	73	50 NNV	Aug. 19	1300	NS	1	46-0	
	35	71°03'	133°05'	137	50 NNV	Aug. 19	1800	NS	1	91-0	
	36	71°31'	130°02'	201	50 NNV	Aug. 19	2300	NS	1	91-0	
	37	71°38'	129°06'	298	50 NNV	Aug. 20	0100	NS	1	100-0	
	38	71°49'	127°55'	420	50 NNV	Aug. 20	0500	NS	1	100-0	
	39	71°57'	126°58'	420	50 NNV	Aug. 20	0600	NS	1	100-0	
	40	72°02'	126°17'	31	50 NNV	Aug. 20	0900	NS	1	15-0	
	41	71°56'	125°42'	71	50 NNV	Aug. 20	1200	NS	1	67-0	
	44	72°18'	126°33'	34	50 NNV	Aug. 21	1800	NS	1	30-0	
	46	72°25'	128°45'	333	50 NNV	Aug. 22	0100	NS	1	100-0	
	51	71°38'	137°16'	2050	50 NNV	Aug. 23	1100	NS	1	86-0	
	49	71°24'	132°55'	1280	50 NNV	Aug. 23	2000	NS	1	100-0	
	53	71°57'	141°00'	2760	50 NNV	Aug. 24	2100	NS	1	100-0	
51-0001	63	70°14'	139°15'	435	50 NNV	Sep. 14	1323	NS	1	100-0	
	64	70°17'	137°42'	73	50 NNV	Sep. 14	1800	NS	1	100-0	
	65	70°36'	136°00'	206	50 NNV	Sep. 14	2300	NS	1	100-0	
	66	70°53'	134°16'	104	50 NNV	Sep. 15	0400	NS	1	100-0	
	67	70°59'	134°27'	287	76-50 NNV	Sep. 15	0610	NS	1	100-0	
	69	71°14'	134°48'	951	50 NNV	Sep. 15	1000	NS	1	100-0	
	70	71°28'	135°06'	1646	50 NNV	Sep. 15	1400	NS	1	100-0	
	71	71°48'	135°46'	201	50 NNV	Sep. 15	1900	NS	1	100-0	
	72	72°21'	136°33'	2551	50 NNV	Sep. 16	0200	NS	1	100-0	
	73	72°46'	134°52'	2515	50 NNV	Sep. 16	1100	NS	1	100-0	
	74	72°41'	132°02'	2066	50 NNV	Sep. 16	1900	NS	1	100-0	
	75	73°41'	129°42'	1024	50 NNV	Sep. 17	0200	NS	1	100-0	
	76	72°24'	129°25'	420	50 NNV	Sep. 17	1000	NS	1	100-0	
	77	72°41'	128°16'	212	50 NNV	Sep. 17	1400	NS	1	100-0	
	78	72°57'	126°48'	135	50 NNV	Sep. 17	1845	NS	1	100-0	
	79	71°30'	128°54'	193	76-50 NNV	Sep. 18	0610	NS	1	100-0	
	82	72°00'	128°54'	374	50 NNV	Sep. 18	2400	NS	1	100-0	
	86	72°30'	139°46'	2999	50 NNV	Sep. 20	1000	NS	1	100-0	
	92	70°32'	139°24'	878	50 NNV	Sep. 21	1100	NS	1	100-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
51-0002	7	69°46'	139°42'	35	282-30 PNV	Aug. 21	0245	NS	1	27-0	
	8	69°51'	139°08'	55	282-30 PNV	Aug. 21	0550	NS	1	50-0	
	12	70°40'	128°15'	20	NS PNH	Aug. 24	2100	NS	1	14	
	13	71°06'	127°13'	218	NS PNH	Aug. 25	0220	NS	1	77-63	
	14	71°30'	126°12'	456	NS PNV	Aug. 25	0900	NS	1	405-0	
	15	71°52'	125°19'	132	NS PNH	Aug. 25	1300	NS	1	27-23	
	16	72°51'	127°04'	176	NS PNV	Aug. 26	0200	NS	1	162-0	
	17	73°21'	126°00'	95	282-30 PNV	Aug. 26	1130	NS	1	81-0	
	18	72°41'	125°34'	18	282-30 PNH	Aug. 27	0015	NS	1	11-9	
	21	69°37'	117°11'	183	282-30 PNV	Aug. 30	1645	NS	1	188-0	
	22	69°24'	117°55'	395	79-30 PNV	Aug. 31	2211	NS	1	360-0	
	23	69°14'	118°26'	117	79-30 PNV	Sep. 01	0053	NS	1	90-0	
	27	70°24'	120°13'	344	79-30 PNV	Sep. 01	1830	NS	1	315-0	
	28	71°02'	120°40'	209	79-30 PNH	Sep. 01	2400	NS	1	59	
	29	71°18'	120°53'	121	NS PNH	Sep. 02	0400	NS	1	15	
	30	70°23'	129°27'	9	79-30 PNH	Sep. 06	0200	NS	1	2	
	43	70°15'	138°12'	357	NS CBH	Sep. 15	1020-1413, 1435-1808	10, 13	11	0,5,9,18,29, 41,54,90	
52-0002	11?	71°57'	125°49'	27	NS-50 PNV	Sep. 02	1800	NS	1	27-0	During the cruise 20 plankton tows were taken at the 23 stations occupied; 21 of these stations are located in the study area. Therefore, tows may not have been taken at a maximum of three stations listed here. Inquiries at University of Washington failed to reveal location of data and samples.
	12?	73°50'	124°40'	54	NS-50 PNV	Sep. 04	1300	NS	1	54-0	
	13?	74°13'	125°10'	54	NS-50 PNV	Sep. 05	0200	NS	1	54-0	
	14?	74°17'	125°42'	433	NS-50 PNV	Sep. 05	0400	NS	1	100-0	
	15?	74°20'	125°48'	567	NS-50 PNV	Sep. 05	0800	NS	1	100-0	
	16?	74°24'	125°43'	664	NS-50 PNV	Sep. 05	1200	NS	1	100-0	
	17?	74°02'	126°38'	301	NS-50 PNV	Sep. 05	1800	NS	1	100-0	
	18?	74°05'	127°13'	664	NS-50 PNV	Sep. 05	2100	NS	1	100-0	
	19?	73°48'	126°52'	363	NS-50 PNV	Sep. 06	0100	NS	1	100-0	
	20?	73°41'	128°00'	786	NS-50 PNV	Sep. 06	0600	NS	1	100-0	
	21?	73°44'	128°33'	1748	NS-50 PNV	Sep. 06	1300	NS	1	100-0	
	22?	72°37'	126°04'	78	NS-50 PNV	Sep. 07	0500	NS	1	78-0	
	23?	73°16'	128°25'	665	NS-50 PNV	Sep. 07	1500	NS	1	100-0	
	24?	72°04'	128°57'	743	NS-50 PNV	Sep. 07	2000	NS	1	100-0	
	25?	71°45'	129°32'	567	NS-50 PNV	Sep. 08	0100	NS	1	100-0	
	26?	71°56'	125°48'	38	NS-50 PNV	Sep. 08	1600	NS	1	38-0	
	27?	71°02'	130°20'	75	NS-50 PNV	Sep. 09	1300	NS	1	75-0	
	28?	70°26'	132°36'	76	NS-50 PNV	Sep. 09	2000	NS	1	76-0	
	29?	70°14'	134°57'	78	NS-50 PNV	Sep. 10	0100	NS	1	78-0	
	30?	70°13'	137°26'	113	NS-50 PNV	Sep. 10	0600	NS	1	100-0	
	31?	70°14'	139°37'	698	NS-50 PNV	Sep. 10	2200	NS	1	100-0	
53-0001	2	70°13'	124°35'	70	73?-17.5 PNV; NS-50 PNV	Aug. 09	0030	NS	1; 1	70-0; NS	
	3	70°35'	124°11'	430	73?-17.5 PNV; NS-50 PNV	Aug. 09	0445	NS	1; 1	98-0; 430-0	
	4	70°50'	123°24'	482	73?-17.5 PNV; NS-50 PNV	Aug. 09	1000	NS	1; 1	98-0; 482-0	
	6	71°04'	123°12'	23	73?-17.5 PNV; NS-50 PNV	Aug. 09	1825	NS	1; 2	23-0	
	7	71°10'	122°32'	24	73?-17.5 PNV; NS-50 PNV	Aug. 10	0030	NS	1; 1	24-0	
	8	71°25'	120°35'	50	73?-17.5 PNV; NS-50 PNV	Aug. 11	1030	NS	1; 1	50-0	
	10	71°32'	119°46'	135	73?-17.5 PNV; NS-50 PNV	Aug. 11	1445	NS	1; 1	98-0; 135-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
53-0001 (cont'd)	11	71°34'	119°11'	19	73?-17.5 PNV; NS-50 PNV	Aug. 11	1915	NS	1; 1	19-0	
	12	71°59'	120°02'	30	73?-17.5 PNV; NS-50 PNV	Aug. 15	0815	NS	1; 1	30-0	
	13	71°50'	118°55'	35	73?-17.5 PNV; NS-50 PNV	Aug. 15	1300	NS	1; 1	35-0	
	32	71°56'	119°48'	120	73?-17.5 PNV; NS-50 PNV	Aug. 25	1045	NS	1	120-0	
	35	71°31'	119°46'	145	73?-17.5 PNV; NS-50 PNV	Aug. 25	2230	NS	1; 1	98-0; 145-0	
54-0003	1?	70°45'	128°00'	53	NS PN	Aug. 09	1500	NS	1?	NS	During the cruise 36 plankton tows were taken at the 37 stations occupied; 17 of these stations are located in the study area. Therefore, a plankton tow may not have been taken at one of the stations listed here. Inquiries with Mr. R.H. Herlinveaux and Pacific Biological Station failed to reveal location of field data and samples.
	2?	71°22'	126°31'	457	NS PN	Aug. 09	2100	NS	1?	NS	
	3?	71°46'	125°14'	457	NS PN	Aug. 10	0200	NS	1?	NS	
	4?	71°54'	125°22'	33	NS PN	Aug. 10	0400	NS	1?	NS	
	5?	71°57'	125°55'	15	NS PN	Aug. 10	0500	NS	1?	NS	
	6?	74°19'	125°00'	37	NS PN	Aug. 12	2200	NS	1?	NS	
	7?	74°27'	125°01'	298	NS PN	Aug. 14	0200	NS	1?	NS	
	27?	71°58'	119°00'	50	NS PN	Sep. 06	1600	NS	1?	NS	
	28?	71°03'	123°01'	101	NS PN	Sep. 07	0200	NS	1?	NS	
	29?	71°00'	123°14'	271	NS PN	Sep. 07	0300	NS	1?	NS	
	30?	70°50'	123°32'	466	NS PN	Sep. 07	0500	NS	1?	NS	
	31?	70°38'	123°53'	472	NS PN	Sep. 07	0800	NS	1?	NS	
	32?	70°27'	124°10'	326	NS PN	Sep. 07	1000	NS	1?	NS	
	33?	74°26'	125°01'	57	NS PN	Sep. 09	1900	NS	1?	NS	
	34?	74°26'	125°01'	57	NS PN	Sep. 09	2200	NS	1?	NS	
	35?	73°58'	125°25'	38	NS PN	Sep. 10	1100	NS	1?	NS	
	36?	73°48'	124°53'	27	NS PN	Sep. 10	1300	NS	1?	NS	
55-0009	55-743	68°52'	133°28'	4	NS PN	Sep. 01-13	NS	NS	NS	NS	
58-0001	35?	70°50'	129°00'	22	NS-50 PN	Sep. 02	1100	NS	1?	NS	During the cruise 24 plankton tows were taken at the 47 stations occupied; six of these stations are located in the study area. Therefore, tows may not have been taken at the stations listed here. Inquiries at Scripps Institution of Oceanography failed to reveal location of data and samples.
	36?	71°26'	128°30'	177	NS-50 PN	Sep. 02	1600	NS	1?	NS	
	37?	71°55'	127°30'	402	NS-50 PN	Sep. 03	0000	NS	1?	NS	
	34?	70°25'	134°00'	48	NS-50 PN	Sep. 03	1400	NS	1?	NS	
	33?	69°50'	138°15'	172	NS-50 PN	Sep. 03	2100	NS	1?	NS	
	32?	69°55'	140°58'	35	NS-50 PN	Sep. 04	0600	NS	1?	NS	
60-0003	60-1014	69°36'	139°16'	4-5	NS PN	Aug. 06	NS	NS	1	Upper 5	
	60-1019	69°31.9'	138°55.3'	35	NS PN	Aug. 10	NS	NS	1	NS	
	60-1031	69°32'	138°50'	NS	NS-100 STNH	Aug. 19	NS	NS	1	11-7	
	60-1033	69°28'	138°57.5'	0-7	571-50 PNH	Aug. 23	NS	NS	2	0,7	
	60-1036	69°32'	138°57'	36	NS PN	Aug. 25	NS	NS	NS	NS	No data available; sample location unknown.
60-0067	Series IX - transect from approx. 130°W east along 75°N latitude to study area boundary	Classified	Classified	500+	SS sampler 10"w x 10"l x 38"h attached within sail of submarine; water intake directed forward; 24 net	24h period in Aug. - early Sep.	NS	60	12	122 av.	Sketch of sampler included in reference.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
60-0067 (cont'd)					receptacles (0.223 mm nitex mesh, 3/4" diam., 12" long) on carousel timed so each exposed to flow for period of 1/2, 1, 6 or 12 h; 2 sec. required to expose adjacent receptacles to flow						
61-0001	61-1013	69°58'	128°54.5'	0.3	73-30 PNH	July 10	NS	NS	1	0	
	61-1014	70°02'	128°42'	9	NS PN	July 10	NS	NS	NS	NS	No data available; sample location unknown.
	61-1015	70°11'	128°35'	12	73-30 PNV	July 10	NS	NS	1	12-0	
	61-1016	70°29.5'	128°19'	8	NS PN	July 11	NS	NS	NS	NS	No data available; sample location unknown.
	61-1017	70°23.6'	128°49'	1'	NS PN	July 11	NS	NS	NS	NS	No data available; sample location unknown.
	61-1021	69°56'	129°37'	11	NS PN	July 12	NS	NS	NS	NS	No data available; sample location unknown.
	61-1026	69°35.5'	131°02'	55	73-30 PNV	July 15	NS	NS	1	55-0	
	61-1027	69°37'	131°02'	7	NS PN	July 16	NS	NS	NS	NS	No data available; sample location unknown.
	61-1032	69°35.5'	131°10'	30	73-30 PNV	July 18	NS	NS	1	30-0	
	61-1035	69°34.2'	131°22.5'	24	73-30 PNV	July 19	NS	NS	1	24-0	
	61-1049	69°24.5'	132°08'	16	73-30 PNV	July 30	NS	NS	1	16-0	
	61-1066	69°21.8'	131°15.5'	18	73-30 PNV	Aug. 07	NS	NS	1	18-0	
	61-1071	69°24.7'	130°55.5'	20	73-30 PNV	Aug. 08	NS	NS	1	20-0	
	61-1076	69°47.5'	130°18'	9	73-30 PNV	Aug. 13	NS	NS	1	9-0	
	61-1082	70°13'	132°36'	28	73-30 PNV	Aug. 19	NS	NS	1	28-0	
	61-1090	69°24.4'	132°58.9'	22	NS PN	Aug. 23	NS	NS	NS	NS	No data available; sample location unknown.
61-0079	61-1401	68°59'	137°24'	2	NS PN	July 06	NS	NS	NS	NS	No data available; sample location unknown.
	61-1401	68°59'	137°24'	2	NS PN	July 09-31	NS	NS	NS	NS	No data available; sample location unknown.
	61-1401	68°59'	137°24'	2	NS PN	Aug. 01-10	NS	NS	NS	NS	No data available; sample location unknown.
62-0001	62-005	69°37'	130°19'	8	73-30 PNV	July 21	0100	NS	1	8-0	
	62-009	69°48'	130°13'	9	73-30 PNV	July 25	2000	NS	1	9-0	
	62-022	69°25'	132°08'	30	233-50 PNV	Aug. 09	0130, 0140	NS	2	30-0	
	62-027	69°14'	132°27'	7	233-50 PNV	Aug. 10	1800	NS	1	7-0	
	62-030	69°48'	130°19'	10	NS PN	Aug. 12	NS	NS	NS	NS	No data available; sample location unknown.
	62-031	69°48	130°18'	10	NS PN	Aug. 12	NS	NS	NS	NS	No data available; sample location unknown.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
62-0001 (cont'd)	62-032	69°48'	130°19'	10	233-50 PNV	Aug. 12	2100	NS	1	10-0	
	62-033	69°55'	129°08'	24	233-50 PNV	Aug. 13	0010	NS	1	24-0	
	62-036	70°31'	128°19'	6	233-50 PNV	Aug. 15	2145, 2315	NS	3	2-0, 4-0, 6-0	
	62-041	70°31'	128°19'	3	233-50 PNH	Aug. 20	1915	NS	1	0.5	
	62-042	70°11.4'	127°03'	68	233-50 PNH	Aug. 21	1945	NS	1	0.5	
	62-043	70°06.6'	125°43'	100	233-50 PNV	Aug. 22	0330	NS	1	100-0	
63-0001	63-002L7	70°08.36'	124°41.3'	4.9	NS PN	July 05	NS	NS	NS	NS	
	63-028L65	70°11.3'	124°50'	7-45	NS PN	Aug. 24	NS	NS	NS	NS	
64-0001	64-001CL3	70°08.49'	124°39.48'	9	NS PN	July 05	NS	NS	NS	NS	
69-0066	56	71°19.5'	137°39.5'	1784	NS PNH?; NS PNV	Aug.	NS	NS	NS	0; NS	Gear used and depths sampled are not clearly reported; additional information indicated that surface plankton hauls may have been taken and vertical tows were perhaps to a depth of 20 m.
	57	70°18.5'	133°55'	39	NS PNV	Aug.	NS	NS	NS	20?	
	58	70°05.5'	136°38'	34	NS PNH?; NS PNV	Aug.	NS	NS	NS	0; 20?	
	59	70°01'	139°00'	184	NS PNH?; NS PNV	Aug.	NS	NS	NS	0; NS	
70-0002	345	70°27.5'	138°57'	740	202-5 PNV	Aug. 27	0753	NS	1	≤200-0	
	346	70°19'	138°47.5'	539	202-5 PNH; 202-5 PNV	Aug. 27	1010	NS	1; 1	≤5-0; ≤200-0	
	348	70°06.5'	138°31.5'	295	202-5 PNH; 202-5 PNV	Aug. 27	1315	NS	1; 1	≤5-0; ≤200-0	
	350	69°50.5'	138°18'	190	202-5 PNH; 202-5 PNV	Aug. 27	1602	NS	1; 1	≤5-0; ≤200-0	
	351	69°56'	138°54.0'	200	202-5 PNH; 202-5 PNV	Aug. 27	1806	NS	1; 1	≤5-0; ≤200-0	
	353	70°29.7'	139°00.4'	610	202-5 PNH; 202-5 PNV	Aug. 28	0651	NS	1; 1	≤5-0; ≤200-0	
	354	70°22'	139°05.5'	NS	202-5 PNH; 202-5 PNV	Aug. 28	0821	NS	1; 1	≤5-0; ≤200-0	
	355	70°15.5'	139°12.3'	451	202-5 PNH; 202-5 PNV	Aug. 28	1026	NS	1; 1	≤5-0; ≤200-0	
	356	70°08.4'	139°15.9'	211	202-5 PNH; 202-5 PNV	Aug. 28	NS	NS	1; 1	≤5-0; ≤200-0	
	357	69°56'?	139°29'?	44	202-5 PNH; 202-5 PNV	Aug. 28	1446	NS	1; 1	≤5-0; ≤200-0	Lat., long. reported incorrectly; mapped position here.
	358	69°42'?	139°42.1'	25	202-5 PNV	Aug. 28	NS	NS	1	≤200-0	Lat. reported incorrectly; mapped position here.
	359	71°31.1'	138°11'	2011	202-5 PNH; 202-5 PNV	Aug. 28	1712	NS	1; 1	≤5-0; ≤200-0	
	360	70°59.8'	138°16.7'	1657	202-5 PNH; 202-5 PNV	Aug. 29	0556	NS	1; 1	≤5-0; ≤200-0	
	361	70°37'	139°29'	1455	202-5 PNH; 202-5 PNV	Aug. 29	0945	NS	1; 1	≤5-0; ≤200-0	
	362	70°22'	139°42'	537	202-5 PNH; 202-5 PNV	Aug. 30	1405	NS	1; 1	≤5-0; ≤200-0	
	363	70°10.1'	139°52.6'	62	202-5 PNH; 202-5 PNV	Aug. 30	1701	NS	1; 1	≤5-0; ≤200-0	
	364	69°59.28'	140°15.76'	51	202-5 PNH; 202-5 PNV	Aug. 30	1907	NS	1; 1	≤5-0; ≤200-0	
	365	69°43'	140°37'	26	202-5 PNH; 202-5 PNV	Aug. 30	2176?	NS	1; 1	≤5-0; ≤200-0	Vilks advises that correct time is probably 2116.
	366	69°11'	137°57	33	202-5 PNH; 202-5 PNV	Aug. 31	1520	NS	1; 1	≤5-0; ≤200-0	
	367	69°22.1'	138°04.8'	42	202-5 PNH; 202-5 PNV	Aug. 31	1640	NS	1; 1	≤5-0; ≤200-0	
	368	69°33'	138°11.8'	100	202-5 PNH; 202-5 PNV	Aug. 31	1758	NS	1; 1	≤5-0; ≤200-0	
	369	70°30.5'	138°19.5'	801	202-5 PNH; 202-5 PNV	Sep. 01	0930	NS	1; 1	≤5-0; ≤200-0	
	370	70°19.8'	138°11'	421	202-5 PNH; 202-5 PNV	Sep. 01	1205	NS	1; 1	≤5-0; ≤200-0	
	371	70°10.5'	137°39.5'	240	202-5 PNH; 202-5 PNV	Sep. 01	1345	NS	1; 1	≤5-0; ≤200-0	
	372	70°01'	137°57'	98	202-5 PNH; 202-5 PNV	Sep. 01	1521	NS	1; 1	≤5-0; ≤200-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
70-0002 (cont'd)	373	69°47'	137°32'	60	202-S PNH; 202-S PNV	Sep. 01	1700	NS	1; 1	≤5-0; ≤200-0	
	374	69°36'	137°20'	44	202-S PNH; 202-S PNV	Sep. 01	1817	NS	1; 1	≤5-0; ≤200-0	
	375	69°27'	137°10'	24	202-S PNH; 202-S PNV	Sep. 01	1929	NS	1; 1	≤5-0; ≤200-0	
	376	70°29.8'	137°49'	866	202-S PNH; 202-S PNV	Sep. 02	0750	NS	1; 1	≤5-0; ≤200-0	
	377	70°21.5'	137°33'	322	202-S PNH; 202-S PNV	Sep. 02	1030	NS	1; 1	≤5-0; ≤200-0	
	378	70°08.2'	137°15.8'	47	202-S PNH; 202-S PNV	Sep. 02	1359	NS	1; 1	≤5-0; ≤200-0	
	379	69°58'	137°00'	38	202-S PNH; 202-S PNV	Sep. 02	1658	NS	1; 1	≤5-0; ≤200-0	
	380	69°51'	136°48'	27	202-S PNH; 202-S PNV	Sep. 02	NS	NS	1; 1	≤5-0; ≤200-0	
	381	69°44.7'	136°37.3'	18	202-S PNH; 202-S PNV	Sep. 02	1835	NS	1; 1	≤5-0; ≤200-0	
	383	70°45.7'	137°04'	1390	202-S PNH; 202-S PNV	Sep. 04	1047	NS	1; 1	≤5-0; ≤200-0	
	384	70°32'	136°40'	700	202-S PNH; 202-S PNV	Sep. 04	1334	NS	1; 1	≤5-0; ≤200-0	
	385	70°18'	136°15'	57	202-S PNH; 202-S PNV	Sep. 04	1617	NS	1; 1	≤5-0; ≤200-0	
	386	70°10.6'	135°54.5'	46	202-S PNH; 202-S PNV	Sep. 04	1731	NS	1; 1	≤5-0; ≤200-0	
	387	70°00.6'	135°39.1'	28	202-S PNH; 202-S PNV	Sep. 04	1858	NS	1; 1	≤5-0; ≤200-0	
	388	70°50.85'	136°17.92'	864	202-S PNH; 202-S PNV	Sep. 05	1024	NS	1; 1	≤5-0; ≤200-0	
	389	70°42.5'	135°52'	495	202-S PNH; 202-S PNV	Sep. 05	1302	NS	1; 1	≤5-0; ≤200-0	
	390	70°37.6'	135°49.4'	87	202-S PNH; 202-S PNV	Sep. 05	1543	NS	1; 1	≤5-0; ≤200-0	
	391	70°26'	135°27'	62	202-S PNV	Sep. 05	1717	NS	1	≤200-0	
	392	70°17.6'	135°11'	55	202-S PNV	Sep. 05	1830	NS	1	≤200-0	
	394	70°41.3'	134°41.5'	58	202-S PNH; 202-S PNV	Sep. 06	0835	NS	1; 1	≤5-0; ≤200-0	
	395	70°26.5'	134°17.5'	62	202-S PNH; 202-S PNV	Sep. 06	1034	NS	1; 1	≤5-0; ≤200-0	
	396	70°17'	134°00'	45	202-S PNH; 202-S PNV	Sep. 06	1215	NS	1; 1	≤5-0; ≤200-0	
	397	71°31.98'	135°57.48'	NS	202-S PNH; 202-S PNV	Sep. 07	NS	NS	1; 1	≤5-0; ≤200-0	
	398	70°57.4'	135°03.4'	457	202-S PNH; 202-S PNV	Sep. 07	1531	NS	1; 1	≤5-0; ≤200-0	
	399	70°52.35'	134°57'	146	202-S PNH; 202-S PNV	Sep. 07	1744	NS	1; 1	≤5-0; ≤200-0	
	400	70°46.5'	134°50'	73	202-S PNH; 202-S PNV	Sep. 07	1909	NS	1; 1	≤5-0; ≤200-0	
	407	71°12'	134°22.5'	850	202-S PNH; 202-S PNV	Sep. 09	0824	NS	1; 1	≤5-0; ≤200-0	
	408	71°01'	134°07'	277	202-S PNH; 202-S PNV	Sep. 09	1109	NS	1; 1	≤5-0; ≤200-0	
	409	70°56'	133°59.5'	85	202-S PNH; 202-S PNV	Sep. 09	NS	NS	1; 1	≤5-0; ≤200-0	
	410	70°47'	133°47'	70	202-S PNH; 202-S PNV	Sep. 09	1408	NS	1; 1	≤5-0; ≤200-0	
	411	70°38'	133°29'	62	202-S PNH; 202-S PNV	Sep. 09	1525	NS	1; 1	≤5-0; ≤200-0	
	412	70°24'	133°09'	42	202-S PNH; 202-S PNV	Sep. 09	NS	NS	1; 1	≤5-0; ≤200-0	
	413	71°18.5'	133°23.5'	699	202-S PNH; 202-S PNV	Sep. 10	0906	NS	1; 1	≤5-0; ≤200-0	
	414	71°09.5'	133°07'	346	202-S PNH; 202-S PNV	Sep. 10	1100	NS	1; 1	≤5-0; ≤200-0	
	415	71°02.8'	132°59.5'	80	202-S PNH; 202-S PNV	Sep. 10	1223	NS	1; 1	≤5-0; ≤200-0	
	416	70°56.2'	132°47'	50	202-S PNH; 202-S PNV	Sep. 10	1345	NS	1; 1	≤5-0; ≤200-0	
	418	70°45.2'	132°27.6'	32	202-S PNH; 202-S PNV	Sep. 10	1818	NS	1; 1	≤5-0; ≤200-0	
	419	70°31.8'	132°10'	63	202-S PNH; 202-S PNV	Sep. 10	1917	NS	1; 1	≤5-0; ≤200-0	
	420	70°52'	128°33'	36	202-S PNH; 202-S PNV	Sep. 12	0842	NS	1; 1	≤5-0; ≤200-0	
	421	70°41'	128°19'	29	202-S PNH; 202-S PNV	Sep. 12	1012	NS	1; 1	≤5-0; ≤200-0	
	423	70°38.7'	129°39.4'	22	202-S PNH; 202-S PNV	Sep. 12	1514	NS	1; 1	≤5-0; ≤200-0	
	424	71°25.7'	132°06'	580	202-S PNH; 202-S PNV	Sep. 13	0917	NS	1; 1	≤5-0; ≤200-0	
	425	71°14.74'	131°54.76'	97	202-S PNH; 202-S PNV	Sep. 13	1150	NS	1; 1	≤5-0; ≤200-0	
	426	71°03.5'	131°42.7'	62	202-S PNV	Sep. 13	1320	NS	1	≤200-0	
	427	70°56.5'	131°24.7'	54	202-S PNV	Sep. 13	1437	NS	1	≤200-0	
	428	70°41.3'	130°52.1'	32	202-S PNV	Sep. 13	1619	NS	1	≤200-0	
	429	70°31.8'	130°41.6'	25	202-S PNV	Sep. 13	1735	NS	1	≤200-0	
	430	70°22.4'	130°31'	18	202-S PNV	Sep. 13	1843	NS	1	≤200-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
70-0002	431	71°26.75'	130°53.87'	300	202-5 PNV	Sep. 14	0843	NS	1	≤200-0	
(cont'd)	432	71°16.6'	130°37.6'	62	202-5 PNH; 202-5 PNV	Sep. 14	1100	NS	1; 1	≤5-0; ≤200-0	
	433	71°07'	130°17.8'	44	202-5 PNH; 202-5 PNV	Sep. 14	1221	NS	1; 1	≤5-0; ≤200-0	
	434	70°56.8'	130°03.6'	32	202-5 PNH; 202-5 PNV	Sep. 14	1406	NS	1; 1	≤5-0; ≤200-0	
	435	70°50'	120°52'	29	202-5 PNH; 202-5 PNV	Sep. 14	1531	NS	1; 1	≤5-0; ≤200-0	
	436	71°25.2'	129°27.3'	69	202-5 PNH; 202-5 PNV	Sep. 15	0907	NS	1; 1	≤5-0; ≤200-0	
	437	71°17.5'	129°10.6'	475	202-5 PNH; 202-5 PNV	Sep. 15	1157	NS	1; 1	≤5-0; ≤200-0	
	438	71°07.9'	128°59'	45	202-5 PNH; 202-5 PNV	Sep. 15	1310	NS	1; 1	≤5-0; ≤200-0	
	439	71°01'	128°49.5'	40	202-5 PNH; 202-5 PNV	Sep. 15	NS	NS	1; 1	≤5-0; ≤200-0	
	442	71°34.3'	129°41.8'	225	202-5 PNH; 202-5 PNV	Sep. 20	1210	NS	1; 1	≤5-0; ≤200-0	
	445	71°02'	125°44'	409	202-5 PNH; 202-5 PNV	Sep. 23	1037	NS	1; 1	≤5-0; ≤200-0	
	446	70°49.8'	123°30.5'	523	202-5 PNH; 202-5 PNV	Sep. 23	2202	NS	1; 1	≤5-0; ≤200-0	
	447	71°20'	119°24'	142	202-5 PNH; 202-5 PNV	Sep. 24	NS	NS	1; 1	≤5-0; ≤200-0	
	448	71°59'	119°30'	128	202-5 PNH; 202-5 PNV	Sep. 24	1233	NS	1; 1	≤5-0; ≤200-0	
70-0067	1	69°24'?	133°36'	NS	202-10 MNH	July 10	NS	NS	1	2	
	2	69°23.5'?	133°41'?	NS	202-10 MNH	July 10	NS	NS	1	2.5	
	3	69°24'?	133°48'?	NS	202-10 MNH	July 10	NS	NS	1	1.5	
	4	69°25.25'?	133°50'?	NS	726-43 PN	July 10	NS	NS	1	1.5	
	5	69°27'?	133°49'?	NS	726-43 PN	July 10	NS	NS	1	1.5	
	6	69°28.75'?	133°44.5'?	NS	726-43 PN	July 10	NS	NS	1	1.5	
	7	69°31'?	133°43'?	NS	202-10 MNH	July 10	NS	NS	1	2.5	
	8	69°34.75'?	133°42'?	NS	202-10 MNH	July 10	NS	NS	1	3	
	9	69°39.5'?	133°43'?	4.5	726-43 PN	July 10	NS	NS	1	3.5	
	23 - Kidluit Bay	NS	NS	NS	726-43 PN	July 11	NS	NS	2	2.5	
	24 - Kidluit Bay	NS	NS	NS	726-43 PN	July 11	NS	NS	1	2	
	25 - Kidluit Bay	NS	NS	NS	726-43 PN	July 11	NS	NS	1	2	
	26 - Kidluit Bay	NS	NS	NS	726-43 PN	July 11	NS	NS	1	1.5	
	27 - Kidluit Bay	NS	NS	NS	726-43 PN	July 11	NS	NS	1	3	
	28 - Kidluit Bay	NS	NS	NS	726-43 PN	July 12	NS	NS	2	1.5	
	29 - Kidluit Bay	NS	NS	NS	726-43 PN	July 12	NS	NS	1	1.5	
	30 - Kidluit Bay	NS	NS	NS	726-43 PN	July 12	NS	NS	1	1.5	
	31	69°26.05'?	132°58.5'?	NS	726-66 PN	July 14	NS	NS	2	4.5	
	32	69°25.95'?	132°57.9'?	NS	726-66 PN	July 14	NS	NS	2	1.5	
	33	69°25.92'?	132°57.5'?	NS	726-66 PN	July 14	NS	NS	2	subsurface	
	34	69°25.78'?	132°56.75'?	NS	726-66 PN	July 14	NS	NS	2	subsurface	
	35	69°26.45'?	132°57.6'?	NS	726-66 PN	July 14	NS	NS	2	1.5	
	36	69°26.33'?	132°57.8'?	NS	726-66 PN	July 15	NS	NS	2	3.5	
	37	69°26 .5'?	132°57.9'?	NS	726-66 PNH	July 15	NS	NS	2	1.5	
	38	69°26.67'?	132°58.23'?	NS	726-66 PN	July 15	NS	NS	2	4.5	
	39	69°26.9'?	132°57.8'?	NS	726-66 PN	July 15	NS	NS	4	4.5	
	40	69°26.98'?	132°57.3'?	NS	726-66 PN	July 15	NS	NS	4	subsurface	
	41	69°26.74'?	132°57.15'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	42	69°27.35'?	132°57.65'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	43	69°27.78'?	132°57.4'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	44	69°27.88'?	132°57'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	45	69°27.12'?	132°59.1'?	NS	726-66 PN	July 15	NS	NS	2	4.5	
	46	69°27.3'?	132°59.4'?	NS	726-66 PN	July 15	NS	NS	2	1.5	

TABLE 8 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
70-0067 (cont'd)	47	69°27.4'?	132°59.15'?	NS	726-66 PN	July 15	NS	NS	2	3	
	48	69°27.05'?	133°00.15'?	NS	726-66 PN	July 15	NS	NS	2	1.5	
	49	69°27.55'?	132°59.55'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	50	69°27.42'?	133°03'?	NS	726-66 PN	July 15	NS	NS	2	subsurface	
	51	69°27.35'?	133°01.03'?	NS	726-66 PN	July 16	NS	NS	2	subsurface	
	52	69°27.34'?	133°01.65'?	NS	726-66 PN	July 16	NS	NS	4	3	
	53	69°27.13'?	133°01.45'?	NS	726-66 PN	July 16	NS	NS	2	1.5	
	54	69°26.93'?	133°01.3'?	NS	726-66 PN	July 16	NS	NS	3?	1.5	
	55	69°27'?	133°01.7'?	NS	726-66 PN	July 16	NS	NS	2	4.5	
	56	69°26.85'?	133°01.7'?	NS	726-66 PN	July 16	NS	NS	2	3	
	57	69°26.83'?	133°01.1'?	NS	726-66 PN	July 17	NS	NS	2?	4.5	
	58	69°26.77'?	133°00.1'?	NS	726-66 PN	July 17	NS	NS	2?	4.5	
	59	69°26.72'?	132°59.5'?	NS	726-66 PN	July 17	NS	NS	2?	4.5	
	60	69°26'.56'	132°59.3'?	NS	726-66 PN	July 17	NS	NS	2	6	
	61	69°26.35'?	132°59.2'?	NS	726-66 PN	July 17	NS	NS	2	6	
	62	69°26.5'?	132°57.1'?	NS	726-66 PN	July 17	NS	NS	2?	4.5	
	63	69°26.3'?	132°56.9'?	NS	726-66 PN	July 17	NS	NS	2?	4.5	
	64	69°27.23'?	132°59.9'?	NS	726-66 PN	July 17	NS	NS	2?	1.5	
	65	69°27.09'?	133°00.6'?	NS	726-66 PN	July 17	NS	NS	2?	1.5	
	66	69°27.15'?	132°58.25'?	NS	726-66 PN	July 17	NS	NS	2	subsurface	
	67	69°27.4'?	132°58.7'?	NS	726-66 PN	July 17	NS	NS	2	subsurface	
	68	69°25.45'?	132°59.2'?	NS	726-66 PN	July 18	NS	NS	1?	3	
	69	69°25.16'?	132°59.7'?	NS	726-66 PN	July 18	NS	NS	1?	3	
	70	69°25.25'?	133°00.42'?	NS	726-66 PN	July 18	NS	NS	1?	3	
	71	69°25.03'?	132°59.95'?	NS	726-66 PN	July 18	NS	NS	1?	3	
	72	69°25'?	133°00.5'?	NS	726-66 PN	July 18	NS	NS	1?	subsurface	
	73	69°25.84'?	132°57.24'?	NS	726-66 PN	July 18	NS	NS	1?	subsurface	
	74	69°27.35'?	132°59.55'?	NS	726-66 PN	July 18	NS	NS	1?	subsurface	
	75	69°26.9'?- 69°26.75'?	133°00.2'?- 132°59.28'?	NS	202-10 MNH	July 19	NS	NS	1?	4.5	
	76	69°26.85'?- 69°27.15'?	132°59.12'?- 132°58.65'?	NS	202-10 MNH	July 19	NS	NS	1?	4.5	
	77	69°27.53'?- 69°27.9'?	132°59'?- 132°59.6'?	NS	202-10 MNH	July 19	NS	NS	1?	4.5	
	78	69°27.8'?- 69°27.6'?	132°59.75'?- 133°00.7'?	NS	202-10 MNH	July 19	NS	NS	1	3	
	79	69°27.67'?- 69°28.1'?	133°01.9'?- 133°02.45'?	NS	202-10 MNH	July 27	NS	NS	2?	3	
	80	69°28.13'?- 69°27.8'?	133°02.6'?- 133°03.5'?	NS	202-10 MNH	July 27	NS	NS	2?	3	
	81	69°27.73'?- 69°27.28'?	133°03.6'?- 133°04.02'?	NS	202-10 MNH	July 27	NS	NS	2?	3	
	82	69°27.23'?- 69°27.35'?	133°04.02'?- 133°03'?	NS	202-10 MNH	July 27	NS	NS	2?	3	
	83	69°27.18'?- 69°26.7'?	133°03.2'?- 133°03.75'?	NS	202-10 MNH	July 27	NS	NS	2	1.5	
	84	69°26.62'?- 69°26.36'?	133°03.75'?- 133°03.5'?	NS	202-10 MNH	July 27	NS	NS	2?	3	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
70-0067 (cont'd)	85	69°26.35'?- 69°26.50'?	133°03.4'?- 133°03.15'?		202-10 MNH	July 27	NS	NS	2?	3	
	86	69°27.09'?	133°00.6'?		726-66 PN	July 27	NS	NS	2?	subsurface	
	87	69°27.23'?	132°59.9'?		726-66 PN	July 27	NS	NS	2?	subsurface	
	88	69°27.35'?	132°59.55'?		726-66 PN	July 27	NS	NS	2?	subsurface	
	89	69°27.23'?	132°59.9'?		726-66 PN	July 27	NS	NS	2?	subsurface	
	90	69°26.50'? 69°27.05'?	132°58.85'? 132°58.9'?		202-10 MNH	July 29	NS	NS	2?	4.5	
	91	69°27.18'?- 69°27.5'?	132°58.75'?- 132°58.9'?		202-10 MNH	July 29	NS	NS	2?	4.5	
	92	69°27.68'?- 69°28.05'?	132°59.4'?- 133°00'?		202-10 MNH	July 29	NS	NS	2	4.5	
	93	69°28.14'?- 69°27.60'?	133°00.4'?- 133°01.3'?		202-10 MNH	July 29	NS	NS	3?	4.5	
	94	69°27.21'?- 69°26.9'?	133°01.72'?- 133°01.3'?		202-10 MNH	July 29	NS	NS	2	4.5	
71-0001	71-502	69°49.3'	132°41.5'	NS	73-30 PNV; 233-30 PNV	July 19	0130	NS	1; 1	8-0	
	71-503	69°58.4'	132°57'	NS	73-30 PNV; 233-30 PNV	July 19	0629	NS	1; 1	17-0	
	71-504	70°16'	131°40'	NS	73-30 PNV; 233-30 PNV	July 19	1515	NS	1; 1	36-0	
	71-505	70°13.2'	131°06'	NS	73-30 PNV; 233-30 PNV	July 19	2100	NS	1; 1	15-0	
71-0008	71-508	69°34.8'	131°18'	NS	73-30 PNV; 233-30 PNV	Aug. 28	2030	NS	1; 1	20-0	
	71-510	69°36.5'	131°04'	NS	73-30 PNV; 233-30 PNV	Aug. 26	2120, 2140	NS	2; 2	30-0, 50-0	
	71-508	69°34.8'	131°18'	NS	73-30 PNV	Dec. 15	2100	NS	1	18-0	
72-0001	72-01	70°44.7'	129°53.8'	NS	202-C PNV	Mar. 21	2055	NS	1	≤200	C net: Hauled through 12" hole drilled through ice or in partly frozen lead where thinner ice cut by saw. To pass net through ice, an umbrella-type suspension arrangement designed. One end of each of three rods was fastened to the net; the opposite ends were attached to a free-moving sleeve on a vertical rod. By pulling the sleeve up along the vertical rod, the net is collapsed; by letting it down, the mouth of the net is spread and kept tight in the form of a triangle. S net: Hauled through open water in lead.
	72-02	70°52.7'	130°04.2'	NS	75-C PNV; 202-S PNV	Mar. 21	2215	NS	1; 1	≤200	
	72-03	71°00.9'	130°24'	NS	75-C PNV; 202-C PNV	Mar. 22	1705	NS	1; 1	≤200	
	72-06	71°33.2'	131°20.1'	NS	75-C PNV; 202-C PNV	Mar. 24	1930	NS	1; 1	≤200	
	72-04	71°05.9'	130°53.5'	NS	202-C PNV	Mar. 25	0015	NS	1	≤200	
	72-05	71°20.9'	130°58.5'	NS	202-S PNV	Mar. 25	1655	NS	1	≤200	
	72-12	70°58.3'	135°12.3'	NS	75-C PNV; 202-C PNV	Mar. 27	1730	NS	1; 1	≤200	
	72-11	70°50.9'	135°00.9'	NS	75-C PNV; 202-C PNV	Mar. 27	2010	NS	1; 1	≤200	
	72-10	70°44.8'	136°50.8'	NS	202-C PNV	Mar. 27	2200	NS	1	≤200	
	72-09	70°40.9'	136°44.8'	NS	202-C PNV	Mar. 27	2330	NS	1	≤200	
	72-13	69°45.3'	140°29.5'	NS	202-C PNV	Mar. 31	1735	NS	1	≤200	
	72-14	69°59.9'	140°10.4'	NS	75-C PNV; 202-C PNV	Mar. 31	1910	NS	1; 1	≤200	
	72-15	70°11.0'	139°59'	NS	202-C PNV	Mar. 31	2130	NS	1	≤200	
	72-16	70°22.7'	139°46.3'	NS	75-C PNV; 202-S PNV	Apr. 01	1620	NS	1; 1	≤200	
	72-17	70°37.3'	139°20.6'	NS	202-C PNV	Apr. 01	1905	NS	1	≤200	
	72-18	71°00.4'	138°23.1'	NS	75-C PNV	Apr. 01	2150	NS	1	≤200	
	72-24	70°37.4'	138°27.2'	NS	75-C PNV	Apr. 02	0001	NS	1	≤200	
	72-23	70°24'	138°33.4'	NS	75-C PNV; 202-C PNV	Apr. 02	1640	NS	1; 1	≤200	
	72-22	70°09.1'	138°31.3'	NS	75-C PNV; 202-C PNV	Apr. 02	1830	NS	1; 1	≤200	
	72-21	70°00.1'	138°29.7'	NS	202-C PNV	Apr. 02	2025	NS	1	≤200	
	72-20	69°48.5'	138°26.4'	NS	202-C PNV	Apr. 02	2220	NS	1	≤200	
	72-19	69°36.4'	138°12.9'	NS	202-C PNV	Apr. 03	0015	NS	1	≤200	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
72-0001 (cont'd)	72-26	70°41.4'	137°29.1'	NS	75-C PNV; 202-C PNV	Apr. 03	1635	NS	1; 1	≤200	
	72-27	71°05.8'	137°47.7'	NS	202-C PNV	Apr. 03	1845	NS	1	≤200	
	72-28	70°28.5'	136°30.6'	NS	202-C PNV	Apr. 03	2115	NS	1	≤200	
	72-25	70°26.4'	137°27.7'	NS	202-C PNV	Apr. 03	2310	NS	1	≤200	
	72-08	70°26'	134°20.8'	NS	75-C PNV; 202-S PNV	Apr. 05	2110	NS	1; 1	≤200	
	72-07	70°25.1'	133°28.6'	NS	75-C PNV; 202-C PNV	Apr. 05	2255	NS	1; 1	≤200	
	72-36	71°19.7'	131°59'	NS	202-S PNV	Apr. 07	1730	NS	1	≤200	
	72-35	71°27.2'	132°05.1'	NS	75-C PNV; 202-S PNV	Apr. 07	1925	NS	1; 1	≤200	
	72-33	71°17.2'	133°03.6'	NS	202-S PNV	Apr. 07	2135	NS	1	≤200	
	72-34	71°07.5'	132°54.9'	NS	202-C PNV	Apr. 07	2305	NS	1	≤200	
	72-32	71°03'	133°58.8'	NS	75-C PNV; 202-S PNV	Apr. 09	1635	NS	1; 1	≤200	
	72-31	71°12.2'	134°16.5'	NS	75-C PNV; 202-S PNV	Apr. 09	1830	NS	1; 1	≤200	
	72-29	70°52.4'	135°53.5'	NS	202-S PNV	Apr. 09	2040	NS	1	≤200	
	72-30	70°40.7'	135°43.6'	NS	202-S PNV	Apr. 09	2215	NS	1	≤200	
72-0004	72-508	69°34.8'	131°18'	NS	S corer	Mar. 17	NS	NS	1	NA	Ice thickness - 157 cm; top and bottom 10 cm analysed.
	72-508	69°34.8'	131°18'	NS	S corer	May 18	NS	NS	1	NA	Ice thickness - 191 cm; sections analysed include (top) 0-10 cm, 35-45 cm, 70-80 cm, 105-115 cm, 140-150 cm and 181-191 cm (bottom).
	72-508	69°34.8'	131°18'	NS	73-30 PNV	May 19	0040	NS	1	20-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	July 09	2110	NS	1	20-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	July 22	0750	NS	1	20-0	
	72-510	69°36.5'	131°04'	NS	73-30 PNV	July 24	0015	NS	3	10-0, 30-0, 50-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	July 28	0605	NS	1	18-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 04	2145	NS	1	18-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 11	1918	NS	1	20-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 18	1825	NS	1	18-0	
	72-513	69°22'	131°00'	NS	73-30 PNV	Aug. 21	1965	NS	1	28-0	
	72-514	69°27'	131°06'	NS	73-30 PNV	Aug. 21	2130	NS	1	30-0	
	72-515	69°32'	131°11'	NS	73-30 PNV	Aug. 22	0020	NS	1	25-0	
	72-519	69°27'	130°55'	NS	73-30 PNV	Aug. 23	2020	NS	2	10-0, 40-0	
	72-518	69°25'	130°47'	NS	73-30 PNV	Aug. 23	2320	NS	1	13-0	
	72-517	69°32'	130°40'	NS	73-30 PNO	Aug. 24	0120	NS	1	5-0	
	72-516	69°32'	130°55'	NS	73-30 PNV	Aug. 24	0250	NS	1	5-0	
	72-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 25	1645	NS	1	20-0	
	72-512	NS	NS	NS	73-30 PNV	Aug. 25	1900	NS	1	30-0	
	72-520	69°30'	131°39'	NS	73-30 PNV	Aug. 25	1950	NS	2	10-0, 40-0	
	72-521	69°26'	131°53'	NS	73-30 PNV	Aug. 25	2400	NS	1	6-0	
	72-522	69°20'	132°05'	NS	73-30 PNV	Aug. 26	0215	NS	2	10-0, 22-0	
	72-523	69°17'	132°14'	NS	73-30 PNV	Aug. 26	0355	NS	1	8-0	
	72-524	69°14'	132°21'	NS	73-30 PNV	Aug. 26	1450	NS	1	5-0	
	72-525	69°09'	132°29'	NS	73-30 PNV	Aug. 26	1825	NS	1	8-0	
	72-510	69°36.5'	131°04'	NS	73-30 PNV	Aug. 28	NS	NS	3	10-0, 30-0, 50-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
72-0006 (cont'd)	72-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 29	1600	NS	1	20-0	
	72-515A	69°33'	131°12'	NS	73-30 PNV	Aug. 29	1840	NS	1	39-0	
	72-508	69°34.8'	131°18'	NS	S corer	Nov. 24	NS	NS	1	NA	Ice thickness - 50 cm; top and bottom 10 cm analysed.
	72-508	69°34.8'	131°18'	NS	73-30 PNV	Nov. 25	2220	NS	1	20-0	
72-0007	A.1	69°37.2'	134°40.6'	NS	12-20 PNH	Aug. 15?	NS	10	1	0	For all stations: report states mid-August collecting period; specific dates estimated from concurrent studies.
	10b	69°32.1'	134°26.9'	NS	12-20 PNH	Aug. 16?	NS	10	1	0	
	6c	69°39.5'	134°43.5'	NS	12-20 PNH	Aug. 17?	NS	10	1	0	
	10 mi. offshore from A.1	69°46'?	134°51'?	NS	12-20 PNH	Aug. 17?	NS	10	1	0	Lat., long. estimated from mapped positions in report.
73-0001	1	69°37.2'	134°40'	3?	73?-12 WNV	Mar. 20?	NS	NS	1	3.1-0	Hauls taken through holes in ice. Report states March collecting period; specific dates and station depths estimated from concurrent studies.
	6c	69°39.5'	134°43.5'	5?	73?-12 WNV	Mar. 20?	NS	NS	1	4.3-0	
	5S	69°37.5'	134°41'	4?	73?-12 WNV	Mar. 21?	NS	NS	1	3.4-0	
	5d	69°36.9'	134°35'	3?	73?-12 WNV	Mar. 21?	NS	NS	1	2.4-0	
	7b	69°35.7'	134°33.2'	3?	73?-12 WNV	Mar. 21?	NS	NS	1	2.4-0	
73-0002	526	69°23.7'	132°59.6'	NS	NBS; 73-30 PNV	July 20	NS; 1730	NS	2; 1	1,5; 5-0	
	527	69°30'	133°15'	5	NBS; VD; 73-30 PNV	July 20	NS; NS; 2155	NS	2; 3; 1	1,5; 0,3,5; 4-0	
	528	69°50'	132°22'	NS	NBS; 73-30 PNV	July 22	NS; 2150	NS	2; 1	1,5; 6-0	
	529	70°01'	131°26'	12	NBS; VD; 73-30 PNV	July 23	NS; NS; 0200	NS	3; 6; 1	1,5,10; 0,1,3,5,7,10; 10-0	
	530	70°11'	130°50'	9	NBS; VD; 73-30 PNV	July 23	NS; NS; 1605	NS	3; 6; 1	1,5,8; 0,1,3,5,7,9; 8-0	
	531	70°23'	130°01'	NS	NBS; 73-30 PNV	July 23	NS; 2030	NS	4; 1	1,5,10,14; 14-0	
	532	70°43'	130°14'	NS	NBS; 73-30 PN; 571-50 PNH	July 24	NS; 0110; 0155	NS	5; 1; 1	1,5,10,20,35; 34-0; 2-0	
	533	70°56'	130°14'	NS	NBS; 73-30 PNV	July 24	NS; 0505	NS	5; 1	1,5,10,20,42; 40-0	
	534	69°43'	133°06'	NS	NBS; 73-30 PNV	July 24	NS; 2345	NS	2; 1	1,5; 6-0	
	535	69°40'	133°53'	NS	NBS; 73-30 PNV	July 25	NS; 2355	NS	2; 1	1,5; 5-0	
	536	69°50'	134°30'	NS	NBS; 73-30 PNV	July 26	NS; 0335	NS	2; 1	1,5; 8-0	
	537	69°48'	135°17'	NS	NBS; 73-30 PNV	July 26	NS; 0615	NS	3; 1	1,5,9; 8-0	
	538	69°33'	136°00'	5	NBS; VD; 73-30 PNV; 571-50 PNH	July 26	NS; NS; 1925; 1955	NS; NS; 10	2; 3; 1; 1	1,5; 0,3,5; 5-0; 1-0	
	539	69°17'	136°34'	NS	NBS	July 26	NS	NS	2	1,3	
	540	69°02'	137°12'	4	NBS; VD; 73-30 PNV	July 27	NS; NS; 0250	NS	2; 3; 1	1,3; 0,3,4; 4-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
73-0002 (cont'd)	541	69°14'	137°54'	NS	NBS; 73-30 PNV; 571-50 PNV	July 27	NS; 1920, 1925; 1945	NS	5; 2; 1	1,5,10,20,33; 30-0,10-0; 30-0	
	542	69°32'	138°18'	NS	NBS; 73-30 PNV; 571-50 PNV	July 28	NS; 0035, 0040,0045, 0050; 0055	NS	7; 4; 1	1,5,10,20,50, 75,93; 90-0, 75-0; 50-0, 10-0; 90-0	
73-0003A	73-508	69°34.8'	131°18'	NS	NBS; 5 corer	Feb. 23	NS	NS	1; 1	10; NA	Ice thickness - 141 cm; sections analysed include (top) 0-10 cm, 35-45 cm, 70-80 cm, 105-115 cm and 131-141 cm (bottom).
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	Mar. 24	NS; NS; 1800	NS	1; 2	10; 10-0,20-0	
	73-508	69°34.8'	131°18'	NS	5 corer	May 19	NS	NS	1	NA	Ice thickness - 169 cm; sections analysed include (top) 0-10 cm, 35-45 cm, 70-80 cm, 105-115 cm and 159-169 cm (bottom).
	73-515A	69°33'	131°12'	NS	73-30 PNV	May 20	2030	NS	1	40-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	May 20	2045	NS	1	10-0	
	73-508	69°34.8'	131°18'	NS	NBS	May 20	NS	NS	1	10	
	73-508	69°34.8'	131°18'	NS	73-30 PNV	May 21	NS; 0005	NS	2	10-0,20-0	
	73-508	69°34.8'	131°18'	20	NBS; 73-30 PNV	June 21	NS; 1905	NS	1; 1	10; 20-0	
	73-508	69°34.8'	131°18'	NS	73-30 PNV	June 28	1650	NS	1	20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	June 29	0015	NS	1	40-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	July 05	2055	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	July 07	NS; 2130	NS	1; 1	10; 20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	July 11	2030	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	73-30 PNV	July 12	1745	NS	1	20-0	
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	July 19	NS; 2003	NS	1; 1	10; 20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	July 20	0130	NS	1	40-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	July 27	2030	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	73-30 PNV	July 28	0045	NS	1	20-0	
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	Aug. 01	NS; 1800	NS	1; 1	10; 20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	Aug. 03	2220	NS	1	40-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	Aug. 13	1450	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	73-30 PNV	Aug. 13	1925	NS	1	20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	Aug. 21	1705	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	Aug. 21	NS; 2245	NS	1; 1	10; 20-0	
	73-515	69°32'	131°11'	NS	73-30 PNV	Sep. 30	2015	NS	1	40-0	
	73-508	69°34.8'	131°18'	NS	NBS; 73-30 PNV	Oct. 01	NS; NS; 1840	NS	1; 2	10; 10-0,20-0	
73-0003B	L1	70°07.5'	129°12.2'	NS	73-30 PNV	Aug. 05?	NS	NS	4	5-0, 10-0, 15-0, 20-0	Report states two weeks in August as collecting period;

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
73-0003B (cont'd)	L 2	69°55.2'	129°31'	NS	73-30 PNV	Aug. 05?	NS	NS	4	5-0, 10-0, 15-0, 20-0	specific dates estimated from concurrent studies.
	L 3	69°50.7'	129°51.5'	NS	73-30 PNV	Aug. 05?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	L 4	69°45.8'	130°13.2'	NS	73-30 PNV	Aug. 05?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	L 5	69°39'	130°30'	NS	73-30 PNV	Aug. 05?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	L 6	69°31'	130°56'	NS	73-30 PNV	Aug. 06?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 1	69°31'	131°24'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 2	69°31'	131°35'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 3	69°24.5'	131°57'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 4	69°21'	132°05'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 5	69°17.5'	132°12'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 6	69°14'	132°19'	NS	73-30 PNV	Aug. 12?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 7	69°13.5'	132°31'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 8	69°05'	132°50'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 9	68°55.5'	132°50'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 10	68°51'	133°20'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 11	68°46'	133°21'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
	E 12	68°41.5'	132°57'	NS	73-30 PNV	Aug. 17?	NS	NS	4	5-0, 10-0, 15-0, 20-0	
74-0001	101	69°27.8'	135°22.2'	NS	737-12 WNV	Mar. 11?	NS	NS	1	1.8-0	Hauls taken through holes in ice. Report states March-April collecting period; specific dates estimated from concurrent studies.
	203	69°32.9'	135°24.7'	NS	737-12 WNV	Mar. 22?	NS	NS	1	3.7-0	
	206	69°33.7'	135°25.1'	NS	737-12 WNV	Mar. 24?	NS	NS	1	4-0	
	207	69°34'	135°24.7'	NS	737-12 WNV	Mar. 24?	NS	NS	1	4.3-0	
	209	69°36.4'	135°24.1'	NS	737-12 WNV	Mar. 26?	NS	NS	1	5.8-0	
	210	69°39.1'	135°22.7'	NS	737-12 WNV	Mar. 26?	NS	NS	1	7-0	
	212	69°39.3'	135°21.7'	NS	737-12 WNV	Mar. 27?	NS	NS	1	7.3-0	
	301	69°42.5'	135°25'	NS	737-12 WNV	Mar. 28?	NS	NS	1	7.6-0	
	302	69°35.8'	135°26.6'	NS	737-12 WNV	Mar. 31?	NS	NS	1	7.6-0	
	404	69°48'	134°00'	NS	737-12 WNV	Apr. 09?	NS	NS	1	5.8-0	
	502	69°51'	134°03.5'	NS	737-12 WNV	Apr. 09?	NS	NS	1	7.6-0	
	413	69°50'	133°54.5'	NS	737-12 WNV	Apr. 11?	NS	NS	1	4.6-0	
	416	69°49.8'	133°38.5'	NS	737-12 WNV	Apr. 12?	NS	NS	1	7.6-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0001 (cont'd)	419	69°49.9'	133°39'	NS	73?-12 WNV	Apr. 12?	NS	NS	1	7.6-0	
	801	69°36.5'	134°39.9'	NS	73?-12 WNV	Apr. 14?	NS	NS	1	2.4-0	
	802	69°36.5'	134°41.9'	NS	73?-12 WNV	Apr. 14?	NS	NS	1	2.7-0	
	803	69°36.3'	134°41'	NS	73?-12 WNV	Apr. 14?	NS	NS	1	2.1-0	
	999	69°46.6'	132°49'	NS	73?-12 WNV	Apr. 18?	NS	NS	1	2.1-0	
74-0003	23	69°18.6'	135°37.6'	2	72-50 PNH	Aug. 03	2000	1.6	1	1	
	21	69°27.2'	135°51.6'	2	72-50 PNH	Aug. 04	2110	1.5	1	1	
	22	69°25'	135°52.3'	1.7	PVC; 72-50 PNH	Aug. 04	NS; 2145	NS; 1.5	1; 1	0; 1	
	24	69°38.6'	135°32.2'	5.1	72-50 PNH	Aug. 05	1230	1.5	1	1	
	8	69°32.9'	135°56.9'	4.7	72-50 PNH	Aug. 05	1500	1.5	1	1	
	10	69°32.2'	135°44.3'	2.7	72-50 PNH	Aug. 05	1645	1.5	1	1	
	17	69°40'	135°12.8'	4.5	72-50 PNH	Aug. 05	1705	1.5	1	1	
	19?	69°30.6'	135°37.4'	1.9	72-50 PNH	Aug. 05	1930	1.5	NS	1	Listed in station summary but not in data table. May be error in date (see Aug. 6).
	29?	69°28.8'	135°47.8'	6	72-50 PNH	Aug. 06	1630	1.5	NS	NS	Listed in station summary but not in data table or on plankton station map.
	28?	69°30.9'	135°44.7'	6.4	72-50 PNH	Aug. 06	1730	1.5	NS	NS	Listed in station summary but not in data table.
	15	69°31.2'	134°56.3'	1.2	72-50 PNH	Aug. 06	1930	1.8	1	1	
	25	69°26.5'	135°33.4'	1.9	72-50 PNH	Aug. 06	1930	1.5	1	1	
	19?	69°30.6'	135°37.4'	1.9	72-50 PNH	Aug. 06	NS	NS	1	1	Not listed in station summary but does appear in data table. May be error in date (see Aug. 5).
	21?	69°27.2'	135°51.6'	2	72-50 PNH	Aug. 06	NS	NS	1	1	Not listed in station summary but does appear in data table.
	22?	69°25'	135°52.3'	1.7	PVC; 72-50 PNH	Aug. 06	NS	NS	1; 1	0; 1	Not listed in station summary but does appear in data table.
	13	69°35.9'	134°48.7'	2	72-50 PNH	Aug. 07	1400	1.5	1	1	
	14	69°34.1'	134°37.7'	1.3	72-50 PNH	Aug. 07	1750	1.5	1	1	
	22	69°25'	135°52.3'	1.7	72-50 PNH	Aug. 10	1645	1.5	1	1	
	21a?	69°27.2'?	135°51.6'?	2?	72-50 PNH	Aug. 10	1845	1.5	1	1	Listed in station summary as 21a but appears in data table as 21.
	24	69°38.6'	135°32.2'	5.1	72-50 PNH	Aug. 10	2230	1.5	1	1	
	17?	69°40'	135°12.8'	4.5	72-50 PNH	Aug. 11	NS	NS	1	1	Not listed in station summary but does appear in data table. May be error in date (see Aug. 12).
	16	69°37'	135°10.4'	2.8	72-50 PNH	Aug. 12	1200	1.5	1	1	
	17?	69°40'	135°12.8'	4.5	72-50 PNH	Aug. 12	1300	1.5	NS	NS	Listed in station summary but not in data table. May be error in date (see Aug. 11).

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0003 (cont'd)	12	69°42'	134°37.1'	3.9	72-50 PNH	Aug. 12	1630	1.5	1	1	
	26	69°26.5'	135°33.4'	1.9	72-50 PNH	Aug. 12	2000	1.5	1	1	
	27	69°30.2'	135°43.3'	1.6	PVC	Aug. 12	NS	NS	1	0	
	2	69°48.7'	133°57.4'	6	72-50 PNH	Aug. 13	0030	1.5	1	1	
	11	69°49.2'	134°36.5'	6	72-50 PNH	Aug. 13	1750	1.5	1	1	
	18	69°33.8'	135°23.5'	1.8	72-50 PNH	Aug. 14	1100	1.5	1	1	
	10?	69°32.2'	135°44.3'	2.7	72-50 PNH	Aug. 14	1330	1.5	NS	NS	Listed in station summary but not in data table.
	23	69°18.6'	135°37.6'	2	72-50 PNH	Aug. 14	1620	1.5	1	1	
	24	69°38.6'	135°32.2'	5.1	PVC	Aug. 24	NS	NS	1	1	
	28	69°30.9'	135°44.7'	6.4	72-50 PNH	Aug. 28	0930	1.5	1	1	
	29?	69°28.8'	135°47.8'	6	72-50 PNH	Aug. 28	1005	1.5	NS	NS	Listed in station summary but not in data table or on plankton station map.
	8	69°32.9'	135°56.9'	4.7	72-50 PNH	Aug. 28	1130	1.5	1	1	
	25	69°26.5'	135°33.4'	1.9	72-50 PNH	Aug. 28	1320	1.5	1	1	
	27?	69°30.2'	135°43.3'	1.6	72-50 PNH	Aug. 28	NS	NS	1	1	Not listed in station summary but does appear in data table.
	19	69°30.6'	135°37.4'	1.9	72-50 PNH	Aug. 29	1145	1.5	1	1	
	10	69°32.2'	135°44.3'	2.7	72-50 PNH	Aug. 29	1215	1.5	1	1	
	21	69°27.2'	135°51.6'	2	72-50 PNH	Aug. 29	1630	1.5	1	1	
	22	69°25'	135°52.3'	1.7	72-50 PNH	Aug. 30	1505	1.5	1	1	
	5	69°52.7'	135°04.7'	13	72-50 PNH	Sep. 01	1025	1.5	1	1	
	17	69°40'	135°12.8'	4.5	PVC; 72-50 PNH	Sep. 07	NS; 1700	NS; 1.5	1; 1	0; 1	
	24	69°38.6'	135°32.2'	5.1	72-50 PNH	Sep. 07	2000	1.5	1	1	
	16	69°37'	135°10.4'	2.8	PVC; 72-50 PNH	Sep. 08	NS; 1100	NS; 1.5	1; 1	0; 1	
	11	69°49.2'	134°36.5'	6	PVC; 72-50 PNH	Sep. 08	NS; 2115	NS; 1.5	1; 1	0; 1	
	26	69°26.5'	135°33.4'	1.9	PVC; 72-50 PNH	Sep. 09	NS; 1400	NS; 1.5	1; 1	0; 1	
	12	69°42'	134°37.1'	3.9	PVC; 72-50 PNH	Sep. 09	NS; 1915	NS; 1.5	1; 1	0; 1	
	18	69°33.8'	135°23.5'	1.8	PVC; 72-50 PNH	Sep. 10	NS; 0930	NS; 1.5	1; 1	0; 1	
74-0007A	74-544	70°33.4'	131°42.8'	41	Phleger corer; Petersen grab; NBS; VD; 233-30 PNV	Aug. 27	NS; NS; NS; NS; 1600	NS	1; 1; 5; 6; 1	NA; NA; 1,5, 10,20,35; 0, 1,3,5,7,10; 35-0	Listed in Knowles 1975 as Station 554.
	74-545	70°23'	131°42'	37	Phleger corer; Petersen grab; NBS; 233-30 PNV	Aug. 27	NS; NS; NS; 1900	NS	1; 1; 5; 1	NA; NA; 1,5, 10,20,35; 35-0	
	74-546	69°56'	133°27'	21	Petersen grab; NBS; 233-30 PNV	Aug. 28	NS; NS; 0325	NS	1; 4; 1	NA; 1,5,10, 18; 18-0	
	74-547	70°18'	135°10.2'	55	Phleger corer; Petersen grab; NBS; 73-30 PNV	Aug. 29	NS; NS; NS; 0545	NS	1; 1; 6; 1	NA; NA; 1,5, 10,20,30,50; 50-0	
	74-548	70°08.1'	135°34.3'	44	Phleger corer; Petersen grab; NBS; 73-30 PNV	Aug. 29	NS; NS; NS; 0550	NS	1; 1; 6; 1	NA; NA; 1,5, 10,20,30,40; 40-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0007A (cont'd)	74-549	69°56.2'	135°47.8'	20	Phleger corer; Petersen grab; NBS; 73-30 PNV	Aug. 30	NS; NS; NS; 1430	NS	1; 1; 4; 1	NA; NA; 1,5, 10,20; 18-0	
	74-550	70°22.1'	136°36.3'	58	Phleger corer; Petersen grab; NBS; 73-30 PNV	Aug. 30	NS; NS; NS; 2345	NS	1; 1 6; 1	NA; NA; 1,5, 10,20,30,55; 55-0	
	74-551	70°06.9'	136°50.2'	40	Phleger corer; Petersen grab; NBS; 73-30 PNV	Aug. 31	NS; NS; NS; 0710	NS	1; 1; 5; 1	NA; NA; 1,5, 10,20,35; 35-0	
	74-552	69°56.2'	137°04.7'	43	Phleger corer; Petersen grab; NBS; VD; 73-30 PNV; 569-50 PND	Aug. 31	NS; NS; NS; NS; 1500; NS	NS	1; 1; 6; 4; 1; 1	NA; NA; 1,5, 10,20,30,40; 0,3,5,10; 35-0; 150-0	
	74-553	70°05.4'	139°08.2'	202	Phleger corer; Petersen grab; NBS; 73-30 PNV	Sep. 01	NS; NS; NS; 0545,0550, 0600	NS	1; 1; 8; 3	NA; NA; 1,5, 10,20,50,70, 100,190; 50-0,100-0, 190-0	
	74-554	69°47.4'	138°55.7'	97	Phleger corer; Petersen grab; NBS, 73-30 PNV	Sep. 01	NS; NS; NS; 1700,1705	NS	2; 1; 7; 2	NA; NA; 1,5, 10,20,50,70, 90; 50-0, 100-0	
	74-555	69°44.9'	139°36.7'	30	Phleger corer; Petersen grab; NBS; 73-30 PNV	Sep. 01	NS; NS; NS; 1950	NS	1; 1; 4; 1	NA; NA; 1,5, 10,26; 26-0	
	74-556	69°27'	138°48.5'	55	Phleger corer; Petersen grab; NBS; 73-30 PNV	Sep. 02	NS; NS; NS; 0120	NS	2; 1; 6; 1	NA; NA; 1,5, 10,20,30,50; 50-0	
	74-557	69°36.8'	138°21'	126	Phleger corer; Petersen grab; NBS; 73-30 PNV	Sep. 02	NS; NS; NS; 1610,1615, 1620	NS	1; 1; 7; 3	NA; NA; 1,5, 10,20,50,70, 120; 50-0, 100-0,120-0	
	74-558	69°33'	136°58'	20	Phleger corer; Petersen grab; NBS	Sep. 02	NS	NS	1; 1; 4	NA; NA; 1,5, 10,18	
	74-559	69°59.7'	135°21'	31	Phleger corer; Petersen grab; NBS; VD; 73-30 PNV	Sep. 03	NS; NS; NS; NS; 0320	NS	1; 1; 5; 4; 1	NA; NA; 1,5, 10,20,30; 0, 3,5,10; 29-0	
74-0007C	74-14 BS	70°09'	133°24'	34	NBS; VD; 73-30 PNV; 233-30 PNV	July 14	NS; NS; 2200	NS	5; 7; 2	1,5,10,20,32; 0,1,3,5,7, 10; 32-0	
	74-7 BS	70°39'	130°06'	24	NBS; VD; 73-30 PNV	July 19	NS; NS; 2300	NS	4; 7; 1	1,5,10,20; 0, 1,3,5,7,10, 20; 20-0	
	74-7 BS	70°39'	130°06'	NS	Phleger corer	July 22-30	NS	NS	1	NA	
	74-9 BS	71°22'	130°24'	NS	Phleger corer	July 22-30	NS	NS	1	NA	
	74-12 BS	71°02'	133°56'	NS	Phleger corer	July 22-30	NS	NS	1	NA	
	74-14 BS	70°09'	133°24'	NS	Phleger corer	July 22-30	NS	NS	4	NA	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0007C (cont'd)	74-9 BS	71°22'	130°24'	11	NBS; VD; 73-30 PNV	July 25	NS; NS; 2350	NS	7; 6; 1	1,5,10,20,50, 75,110; 0,1, 3,5,7,10; 100-0	
	74-9 BS	71°22'	130°24'	11	73-30 PNV; 233-30 PNV	July 26	0015,0035; 0007,0025, 0040	NS	2; 3	20-0,50-0; 20-0,50-0, 100-0	
	74-12 BS	71°02'	133°56'	267	NBS; 73-30 PNV	July 29	NS; 2310, 2325,2335, 2338,2342, 2348	NS	9; 6	1,5,10,20,50, 75,100,200, 260; 5-0, 10-0,20-0, 50-0,100-0, 200-0	
	74-29 BS	70°17'	136°26'	52	Phleger corer; Petersen grab; NBS; VD; 73-30 PNV	Aug. 02	NS; NS; NS; NS; 2130	NS	2; 1; 6; 8; 4	NA; NA; 1,5, 10,20,30,50; 0,1,3,5,7, 10,20,30; 5-0,10-0, 20-0,50-0	
	74-30 BS	69°55'	136°12'	NS	Phleger corer; Petersen grab	Aug. 02	NS	NS	2; 1	NA; NA	
	74-30 BS	69°55'	136°12'	25	NBS; 73-30 PNV	Aug. 03	NS; 0130	NS	4; 3	1,5,10,24; 5-0,10-0, 20-0	
	74-22 BS	69°46'	138°52'	NS	Phleger corer	Aug. 12	NS	NS	2	NA	
	74-22 BS	69°46'	138°52'	79	NBS; VD; 73-30 PNV	Aug. 13	NS; NS; 0130	NS	6; 8; 5	1,5,10,20,50, 75; 0,1,3,5, 7,10,20,30; 5-0,10-0, 20-0,50-0, 75-0	
	74-23 BS	70°01'	138°56'	NS	Phleger corer; Petersen grab	Aug. 15-16	NS	NS	1; 1	NA	
	74-23 BS	70°01'	138°56'	226	NBS; 73-30 PNV	Aug. 16	NS; 2200, 2220	NS	8; 6	1,5,10,20,50, 75,100,220; 10-0,20-0, 30-0, 50-0, 100-0,200-0	
74-0008	12	69°51.19'	137°41.1'	81	NS N	Aug. 18	NS	60	1	0	Tow speed = 4 knots
	39	69°18.1'	138°14.16'	35	NS N	Aug. 24	NS	60	1	0	
74-0010A	74-508	69°34.8'	131°18'	NS	Petersen grab	Feb. 27	NS	NS	4	NA	
	74-515	69°32'	131°11'	NS	S corer	Mar. 01	NS	NS	1	NA	Ice thickness - 144 cm; bottom 5 cm section analysed.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0010A (cont'd)	74-508	69°34.8'	131°18'	NS	S corer	Mar. 02	NS	NS	5	NA	Ice thickness - 143 cm; sections analysed include (top) 0-10 cm, 35-45 cm, 70-80 cm, 105-115 cm and 133-143 cm (bottom).
	74-515	69°32'	131°11'	NS	73-30 PNV	Mar. 02	0200	NS	1	40-0	
	74-508	69°34.8'	131°18'	NS	73-30 PNV	Mar. 03	0200	NS	1	15-0	
	74-508	69°34.8'	131°18'	NS	S corer	May 24	NS	NS	7	NA	Ice thickness - 203 cm; sections analysed include (top) 0-10 cm, 35-70 cm, 70-105 cm, 105-140 cm, 140-175 cm, 175-193 cm and 193-203 cm (bottom).
	74-508	69°34.8'	131°18'	NS	Petersen grab	May 24	NS	NS	2	NA	
	74-515	69°32'	131°11'	NS	S corer	May 25	NS	NS	1	NA	Ice thickness - 186 cm; bottom 5 cm section analysed.
	74-515	69°32'	131°11'	NS	73-30 PNV	May 26	0110	NS	1	27-0	
	74-508	69°34.8'	131°18'	NS	73-30 PNV	May 26	0200	NS	1	19-0	
74-0010C	508	69°34.8'	131°18'	NS	VD	July 13	NS	NS	7	0,1,3,5,7,10,20	
	510	69°36.5'	131°04'	NS	VD	July 16	NS	NS	6	0,1,3,5,7,10	
	515	69°32'	131°11'	NS	Petersen grab	July 22	NS	NS	1	NA	
	508	69°34.8'	131°18'	NS	Phleger corer; Petersen grab	July 27	NS	NS	1; 1	NA	
	515	69°32'	131°11'	NS	Phleger corer; Petersen grab; VD	July 27	NS	NS	2; 1; 8	NA; NA; 0,1,3,5,7,10,20,30	
	515	69°32'	131°11'	NS	Petersen grab	July 30	NS	NS	1	NA	
	560	69°44'	130°32'	NS	VD	July 31	NS	NS	5	0,1,3,5,7	
	508	69°34.8'	131°18'	NS	Phleger corer	Aug. 03	NS	NS	2	NA	
	515	69°32'	131°11'	NS	Phleger corer; Petersen grab; VD	Aug. 03	NS	NS	2; 1; 8	NA; NA; 0,1,3,5,7,10,20,30	
	520	69°30'	131°39'	NS	Phleger corer; Petersen grab; VD	Aug. 07	NS	NS	1; 1; 8	NA; NA; 0,1,3,5,7,10,20,30	
	523	69°17'	132°14'	NS	Phleger corer; Petersen grab	Aug. 07	NS	NS	1; 1	NA	
	510	69°36.5'	131°04'	NS	Phleger corer; Petersen grab	Aug. 08	NS	NS	1; 1	NA	
	550-Clupea Harbour	NS	NS	NS	Phleger corer; Petersen grab	Aug. 08	NS	NS	1; 1	NA	
	515	69°32'	131°11'	NS	VD	Aug. 10	NS	NS	8	0,1,3,5,7,10,20,30	
	508	69°34.8'	131°18'	NS	Phleger corer; Petersen grab	Aug. 15	NS	NS	1; 1	NA	
	515	69°32'	131°11'	NS	Petersen grab	Aug. 15-19	NS	NS	1	NA	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0010C (cont'd)	515	69°32'	131°11'	NS	VD	Aug. 16	NS	NS	8	0,1,3,5,7,10, 20,30	
	515	69°32'	131°11'	NS	Phleger corer; Petersen grab	Aug. 19	NS	NS	4; 1	NA	
	508	69°34.8'	131°18'	NS	Phleger corer; Petersen grab	Aug. 21	NS	NS	4; 1	NA	
74-0011	A	69°33.5'?	139°34'?	2?	505-60 FNH	July 08	NS	NS	3	0	Stn. A: net kept 1 m behind boat and towed 120 m along-shore. Stns. B,C: net kept 17.5 m behind boat and towed for premeasured distance of approx. 500 m; for subsurface tows weights were attached to leading steel hoop to keep net at desired depth. Sketch of net included in reference. All lats., longs. estimated from mapped positions in report.
	B	69°33.7'?	139°34'?	2?	505-60 FNH	July 08	NS	NS	6	0,1	
	B	69°33.7'?	139°34'?	2?	505-60 FNH	July 29	NS	NS	3	0	
	A	69°33.5'?	139°34'?	2?	505-60 FNH	Aug. 13	NS	NS	3	0	
	B	69°33.7'?	139°34'?	2?	505-60 FNH	Aug. 13	NS	NS	3	0	
	C	69°35'?	139°30'?	10?	505-60 FNH	Aug. 25	NS	NS	9	0,1.5,5	
	A	69°33.5'?	139°34'?	2?	505-60 FNH	Aug. 26	NS	NS	3	0	
	B	69°33.7'?	139°34'?	2?	505-60 FNH	Aug. 26	NS	NS	3	0	
	B	69°33.7'?	139°34'?	2?	505-60 FNH	Sep. 09	NS	NS	6	0,1	
74-0019	1	69°34.6'?	135°23.5'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	Hauls made through 14" diameter ice holes. All lats., longs. estimated from mapped positions in report. Specific sampling dates for each station not available.
	3	69°34.3'?	135°23.5'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	4	69°34.25'?	135°23.5'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	6	69°34.1'?	135°23.5'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	11	69°34.17'?	135°24.5'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	12	69°34.17'?	135°24.2'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	13	69°34.17'?	135°23.9'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	14	69°34.17'?	135°23.4'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
	17	69°34.17'?	135°22.8'?	NS	100-30 WNV	Mar. 05-07	NS	NS	1	NS	
74-0027	1 (mouth of bay)	70°02'	124°54'	NS	VD; NS PPN; 60-25 and 240-50 PNH and PNV	Aug. 13	NS	NS	1; 1; NS	4; 1-2; NS	
	2 (mid-bay)	70°02'	124°54'	NS	VD; NS PPN; 60-25 and 240-50 PNH and PNV	Aug. 13	NS	NS	1; 1; NS	4; 1-2; NS	
	3 (near oil spill site)	70°02'	124°54'	NS	VD; NS PPN; 60-25 and 240-50 PNH and PNV	Aug. 13	NS	NS	1; 1; NS	4; 1-2; NS	
74-0119	74-004	69°45'	133°12'	8.6	NS PN	Apr. 01	NS	NS	NS	NS	For all stations: no data available; sample location unknown.
	74-S-001	69°33.55'	132°59.6'	2	NS PN	July 22	NS	NS	NS	NS	
	74-004	69°26.1'	133°00.3'	NS	NS PN	July 28-29	NS	NS	NS	NS	
	74-005	69°25.9'	132°58.4'	NS	NS PN	July 30	NS	NS	NS	NS	
	74-003	69°41'	132°48.67'	4.3	NS PN	July 31	NS	NS	NS	NS	
	74-S-001	69°33.55'	132°59.6'	1.5	NS PN	Aug. 05	NS	NS	NS	NS	
	74-001	69°42'	137°28'	35-48	NS PN	Aug. 11	NS	NS	NS	NS	
	74-002	69°45'	137°08'	30	NS PN	Aug. 12	NS	NS	NS	NS	
	74-003	70°07'	134°38'	30-35	NS PN	Aug. 14	NS	NS	NS	NS	
	74-004	70°12'	134°20'	30	NS PN	Aug. 14	NS	NS	NS	NS	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
74-0119	74-5-007	69°43.57'	132°33.67'	1	NS PN	Aug. 15	NS	NS	NS	NS	
(cont'd)	74-006	69°38'	137°40'	60	NS PN	Aug. 16	NS	NS	NS	NS	
	74-007	69°52'	137°40'	72	NS PN	Aug. 18	NS	NS	NS	NS	
	74-006	69°25.5'	132°59.6'	NS	NS PN	Aug. 21	NS	NS	NS	NS	
	74-007	69°25'	132°58.4'	NS	NS PN	Aug. 21	NS	NS	NS	NS	
	74-008	69°24.8'	132°58.6'	NS	NS PN	Aug. 21	NS	NS	NS	NS	
	74-009	69°24.3'	132°59.6'	NS	NS PN	Aug. 21	NS	NS	NS	NS	
	74-009	69°21'	138°37'	6	NS PN	Aug. 21	NS	NS	NS	NS	
	74-010	69°30'	138°40'	16	NS PN	Aug. 22	NS	NS	NS	NS	
	74-011	69°27'	138°25'	70	NS PN	Aug. 22	NS	NS	NS	NS	
	74-012	69°27'	138°00'	65	NS PN	Aug. 22	NS	NS	NS	NS	
	74-013	69°27'	137°20'	50	NS PN	Aug. 22	NS	NS	NS	NS	
	74-014	69°29'	137°29'	NS	NS PN	Aug. 22	NS	NS	NS	NS	
	74-015	69°30'	137°20'	35	NS PN	Aug. 23	NS	NS	NS	NS	
	74-016	69°30'	136°45'	NS	NS PN	Aug. 23	NS	NS	NS	NS	
	74-018	69°17'	137°10'	NS	NS PN	Aug. 23	NS	NS	NS	NS	
	74-019	69°17'	137°33'	35	NS PN	Aug. 23	NS	NS	NS	NS	
	74-020	69°16'	138°52'	39	NS PN	Aug. 23	NS	NS	NS	NS	
	74-021	69°18'	138°10'	35	NS PN	Aug. 23	NS	NS	NS	NS	
	74-A009	69°26.6'	132°59'	NS	NS PN	Aug. 24-29	NS	NS	NS	NS	
	74-023	69°49'	135°00'	11	NS PN	Aug. 25	NS	NS	NS	NS	
	74-024	69°53'	135°00'	NS	NS PN	Aug. 25	NS	NS	NS	NS	
	74-026	70°01'	133°00'	23	NS PN	Aug. 26	NS	NS	NS	NS	
	74-027	69°58'	132°05'	18	NS PN	Aug. 26	NS	NS	NS	NS	
	74-028	69°57'	132°00'	22	NS PN	Aug. 26	NS	NS	NS	NS	
	74-029	70°12'	131°50'	24	NS PN	Aug. 26	NS	NS	NS	NS	
	74-030	70°27'	131°51'	35	NS PN	Aug. 26	NS	NS	NS	NS	
	74-006	69°38.07'	132°50.9'	2.2	NS PN	Sep. 06	NS	NS	NS	NS	
75-0004	16	69°37.1'	135°10.5'	NS	72-50 PNH	July 10	NS	1.5	1	1	For all stations: non-quantitative zooplankton samples were also collected (73?-12 WNH, 73?-12 WNV) at selected (but not specified) stations.
	12	69°42.2'	134°35.8'	NS	72-50 PNH	July 11	NS	1.5	1	1	
	68	69°35'?	134°08'?	NS	72-50 PNH	July 12	NS	1.5	1	1	Lat., long. estimated from mapped position in report.
	11	69°49.1'	134°36.3'	NS	PVC; 72-50 PNH	July 13	NS	NS; 1.5	1; 1	0; 1	
	18	69°33.8'?	135°23'?	NS	72-50 PNH	July 15	NS	1.5	1	1	Lat., long. estimated from mapped position in report.
	8	69°32.7'	135°57'	NS	72-50 PNH	July 16	NS	1.5	1	1	
	9	69°37.4'	135°54.2'	NS	72-50 PNH	July 16	NS	1.5	1	1	
	11	69°49.1'	134°36.3'	NS	72-50 PNH	July 16	NS	1.5	1	1	
	68	69°35'?	134°08'?	NS	72-50 PNH	July 16	NS	1.5	1	1	Lat., long. estimated from mapped position in report.
	2	69°48.8'	133°58.2'	NS	72-50 PNH	July 17	NS	1.5	1	1	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0004 (cont'd)	26	69°44.7'	134°21.3'	NS	PVC	July 18	NS	NS	1	0	
	82	69°16.5'?	135°17.1'?	NS	PVC	July 20	NS	NS	1	0	Lat., long. estimated from mapped position in report.
	63	69°24.4'?	135°46'?	NS	72-50 PNH	July 26	NS	1.5	1	1	Lat., long. estimated from mapped position in report.
	82	69°16.5'?	135°17.1'?	NS	72-50 PNH	July 26	NS	1.5	1	1	Lat., long. estimated from mapped position in report.
	8	69°32.7'	135°57'	NS	PVC; 72-50 PNH	July 28	NS	NS; 1.5	1; 1	0; 1	
	7	69°46'	135°57.8'	NS	PVC	July 29	NS	NS	1	0	
	12	69°42.2'	134°35.8'	NS	PVC; 72-50 PNH	July 30	NS	NS; 1.5	1; 1	0; 1	
	16	69°37.1'	135°10.5'	NS	PVC; 72-50 PNH	July 30	NS	NS; 1.5	1; 1	0; 1	
	1	69°54.9'	134°17'	NS	PVC; 72-50 PNH	July 31	NS	NS; 1.5	1; 1	0; 1	
	11	69°49.1'	134°36.3'	NS	PVC; 72-50 PNH	July 31	NS	NS; 1.5	1; 1	0; 1	
	2	69°48.8'	133°58.2'	NS	PVC: 72-50 PNH	Aug. 01	NS	NS; 1.5	1; 1	0; 1	
	73	69°40.4'	133°20.3'	NS	72-50 PNH	Aug. 01	NS	1.5	1	1	
	22	69°25'	135°49.7'	NS	72-50 PNH	Aug. 06	NS	1.5	1	1	
	21	69°27.5'	135°49.6'	NS	72-50 PNH	Aug. 10	NS	1.5	1	1	
	22	69°25'	135°49.7'	NS	PVC; 72-50 PNH	Aug. 13	NS	NS; 1.5	1; 1	0; 1	
	18	69°33.8'?	135°23'?	NS	PVC	Aug. 15	NS	NS	1	0	Lat., long. estimated from mapped position in report.
	8	69°32.7'	135°57'	NS	PVC	Aug. 16	NS	NS	1	0	
	71	69°18.5'	136°09'	NS	PVC	Aug. 18	NS	NS	1	0	
75-0006	1	70°15'	139°04.27'	453	73?-NS MNH	Aug. 05	NS	60	1	various	
	2	70°40.74'	134°42.91'	55	73?-NS MNH	Aug. 05	NS	60	1	various	
	3	70°00.36'	135°28.24'	33	73?-NS MNH	Aug. 05	NS	60	1	various	
	4	70°53.97'	132°28.23'	64	73?-NS MNH	Aug. 07	NS	60	1	various	
	5	71°18.85'	130°36.51'	72	73?-NS MNH	Aug. 08	NS	60	1	various	
	6	70°09.16'	132°14.58'	30	73?-NS MNH	Aug. 08	NS	60	1	various	
	7	69°33.5'	138°55.5'	9	73?-NS MNH	Aug. 09	NS	60	1	various	
	8	69°34.74'	137°29.39'	20	73?-NS MNH	Aug. 12	NS	60	1	various	
	9	70°05.42'	137°36.56'	83	73?-NS MNH	Aug. 13	NS	60	1	various	
	11	69°51.75'	136°00.33'	17	73?-NS MNH	Aug. 13	NS	60	1	various	
	13	70°27.49'	134°53.14'	50	73?-NS MNH	Aug. 13	NS	60	1	various	
	15	70°01.71'	134°03.07'	20	73?-NS MNH	Aug. 14	NS	60	1	various	
	16	70°20.92'	133°56.51'	48	73?-NS MNH	Aug. 14	NS	60	1	various	
	17	70°39.48'	133°60.49'	64	73?-NS MNH	Aug. 14	NS	60	1	various	
	20	70°00.31'	133°01.06'	14	73?-NS MNH	Aug. 15	NS	60	1	various	
	21	70°30.02'	133°00.52'	38	73?-NS MNH	Aug. 15	NS	60	1	various	
	22	70°54.78'	133°22.03'	70	73?-NS MNH	Aug. 15	NS	60	1	various	
	25	70°00.65'	132°00.3'	15	73?-NS MNH	Aug. 16	NS	60	1	various	
	26	70°29.37'	132°02.21'	35	73?-NS MNH	Aug. 16	NS	60	1	various	
	27	70°59.64'	132°03.26'	65	73?-NS MNH	Aug. 16	NS	60	1	various	
	30	70°19.56'	131°02.33'	18	73?-NS MNH	Aug. 17	NS	60	1	various	
	31	70°50.27'	131°09.43'	50	73?-NS MNH	Aug. 17	NS	60	1	various	
	32	71°10.51'	131°15.43'	80	73?-NS MNH	Aug. 17	NS	60	1	various	
	34	70°22.88'	130°19.38'	18	73?-NS MNH	Aug. 18	NS	60	1	various	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0006 (cont'd)	35	70°56.5'	129°59.45'	35	73?-NS MNH	Aug. 18	NS	60	1	various	
	36	71°22.74'	130°12.19'	73	73?-NS MNH	Aug. 18	NS	60	1	various	
	37	69°48.99'	133°20.75'	10	73?-NS MNH	Aug. 19	NS	60	1	various	
	38	69°53.66'	134°57.84'	14	73?-NS MNH	Aug. 19	NS	60	1	various	
	39	70°14.38'	134°58.24'	45	73?-NS MNH	Aug. 20	NS	60	1	various	
	40	70°26.32'	136°08.45'	58	73?-NS MNH	Aug. 20	NS	60	1	various	
	41	70°19.29'	136°40.05'	60	73?-NS MNH	Aug. 20	NS	60	1	various	
	42	70°00.56'	136°33.4'	30	73?-NS MNH	Aug. 20	NS	60	1	various	
	43	69°42.69'	136°33.35'	15	73?-NS MNH	Aug. 20	NS	60	1	various	
	44	69°37.76'	137°19.52'	48	73?-NS MNH	Aug. 21	NS	60	1	various	
	45	69°44.7'	138°07.09'	155	73?-NS MNH	Aug. 21	NS	60	1	various	
	46	69°25'	138°02'	65	73?-NS MNH	Aug. 21	NS	60	1	various	
	47	69°13.5'	137°54'	27	73?-NS MNH	Aug. 21	NS	60	1	various	
	48	70°10'	133°25'	28	73?-NS MNH	Aug. 23	NS	60	1	various	
75-0009	75-302	70°54'	134°11'	NS	IKT	Aug. 04	NS	NS	1	NS	
	75-303	71°17'	130°56'	NS	IKT	Aug. 04	NS	NS	1	NS	
	75-304	70°55'	129°36'	NS	IKT	Aug. 05	NS	NS	1	NS	
	75-305	70°15'	131°55'	NS	IKT	Aug. 06	NS	NS	1	NS	
	75-306	70°18'	133°35'	NS	IKT	Aug. 06	NS	NS	1	NS	
	75-308	70°06'	136°25'	NS	IKT	Aug. 07	NS	NS	1	NS	
	75-315	70°06'	135°18'	NS	IKT	Aug. 11	NS	NS	1	NS	
	75-318	70°11'	132°58'	NS	IKT	Aug. 18	NS	NS	1	NS	
	75-319	70°13'	134°39'	NS	IKT	Aug. 20	NS	NS	1	NS	
	75-320	70°39'	135°38'	NS	IKT	Aug. 21	NS	NS	1	NS	
	75-321	69°20'	138°13'	NS	IKT	Aug. 25	NS	NS	1	NS	
	75-322	69°28'	138°31'	NS	IKT	Aug. 29	NS	NS	1	NS	
75-0010A	75-508	69°34.8'	131°18'	NS	73-30 PNV	Feb. 27	NS	NS	1	20-0	
75-0010B	515	69°32'	131°11'	NS	VD	June 15	NS	NS	1	1	
	515	69°32'	131°11'	NS	VD	June 26	NS	NS	1	1	
	515	69°32'	131°11'	NS	VD	July 02	NS	NS	1	1	
	515	69°32'	131°11'	NS	VD	July 09	NS	NS	1	1	
	515	69°32'	131°11'	NS	VD; Petersen grab	July 15	NS	NS	1; 1	1; NA	
	515	69°32'	131°11'	NS	VD	July 21	NS	NS	1	1	
	515	69°32'	131°11'	NS	Petersen grab	July 26	NS	NS	1	NA	
	507	69°34.5'	131°15'	NS	VD	Aug. 13	NS	NS	1	1	
	507	69°34.5'	131°15'	NS	VD	Aug. 14	NS	NS	1	1	
	507	69°34.5'	131°15'	NS	VD	Aug. 15	NS	NS	1	1	
	515	69°32'	131°11'	NS	VD	Aug. 16	NS	NS	1	1	
75-0012A	75-565	70°08'	132°37'	NS	73-30 PNV	June 17	2330	NS	1	30-0	
	75-566	70°06'	138°56'	NS	VD; 73-30 PNV	July 06	NS; 0240	NS	3; 1	1,10,20; 315-0	
	75-567	70°49'	136°22'	NS	VD; 73-30 PNV	July 13	NS; 0350	NS	3; 1	1,10,20; 700-0	
	75-566	70°06'	138°56'	NS	Petersen grab	July 15	NS	NS	1	NA	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0012A (cont'd)	75-568	70°14'	139°04'	NS	VD	July 18	NS	NS	3	1,10,20	
	75-568	70°14'	139°04'	NS	Petersen grab	July 19	NS	NS	1	NA	
	75-565	70°08'	132°37'	NS	Petersen grab	July 25	NS	NS	1	NA	
75-0012B	601	69°23.3'	133°47'	NS	73-30 PNH	July 11	2250	NS	1	0	
	602	69°18.6'	133°54.5'	NS	73-30 PNV	July 12	0100	NS	1	8-0	
	603	69°15.6'	134°10'	NS	73-30 PNV	July 12	0300	NS	1	7-0	
	604	69°32.3'	133°52.5'	NS	1000-100 PNH	July 13	1905	10	1	3	
	606	69°31.8'	133°57'	NS	73-30 PNV	July 14	0005	NS	1	14-0	
	610	69°33.5'	134°05'	NS	73-30 PNV	July 15	0225	NS	1	17-0	
	612	69°35.9'	133°58.5'	NS	73-30 PNV	July 17	1845	NS	1	5.5-0	
	613	69°38.5'	133°55'	NS	73-30 PNV	July 17	2015	NS	1	2.5-0	
75-0012D	75-562	69°35'	133°05'	NS	VD	May 09	NS	NS	1	1	
	75-563	69°27'	133°01'	NS	VD	May 09	NS	NS	1	1	
	75-564	69°40'	132°40'	NS	VD	May 09	NS	NS	1	1	
75-0012E	75-569	70°14'	139°04'	NS	NBS	Aug. 04	NS	NS	9	1,5,10,20,50, 70,100,300, 430	
	75-570	70°42'	134°45'	NS	NBS; 1000-100 PNH	Aug. 06	NS; 0015	NS; 15	5; 1	1,5,10,20,50; 5-10	
	75-571	70°02'	135°34'	NS	NBS	Aug. 06	NS	NS	5	1,5,10,20,30	
	75-572	70°56'	132°33'	NS	NBS; VD; 1000-100 PNH	Aug. 07	NS; NS; 2100	NS; NS; 10	6; 5; 1	1,5,10,20,50, 60; 1,3,5, 10,20; 5-10	
	75-573	71°22'	130°24'	NS	NBS	Aug. 07	NS	NS	6	1,5,10,20,50, 66	
	75-574	70°06'?	132°24'?	NS	VD	Aug. 08	NS	NS	5	1,3,5,10,20	Lat., long. estimated from mapped position in report.
	75-575	69°33'?	139°00'?	NS	VD	Aug. 09	NS	NS	5	0,1,3,5,7	Lat., long. estimated from mapped position in report.
75-0026	Control 1	70°00'	125°00'	NS	VD	Feb. 07	NS	NS	NS	midwater	
	Control 2	70°00'	125°00'	NS	VD	Feb. 07	NS	NS	NS	midwater	
	Oiled 1	70°00'	125°00'	NS	VD	Feb. 07	NS	NS	NS	midwater	
	Oiled 2	70°00'	125°00'	NS	VD	Feb. 07	NS	NS	NS	midwater	
	Control	70°00'	125°00'	NS	VD	June 04	NS	NS	5	2.4,3.5,4.5, 5,6.5	
	Oiled	70°00'	125°00'	NS	VD	June 04	NS	NS	5	2.4,3.5,4.5, 5,6.5	
	Control 1	70°00'	125°00'	NS	VD	July 18	NS	NS	3	1,4,8	
	Control 2	70°00'	125°00'	NS	VD	July 18	NS	NS	3	1,4,8	
	Oiled 1	70°00'	125°00'	NS	VD	July 18	NS	NS	3	1,4,8	
	Oiled 2	70°00'	125°00'	NS	VD	July 18	NS	NS	3	1,4,8	
	Control 1	70°00'	125°00'	NS	VD	July 19	NS	NS	3	1,4,8	
	Control 2	70°00'	125°00'	NS	VD	July 19	NS	NS	3	1,4,8	
	Oiled 1	70°00'	125°00'	NS	VD	July 19	NS	NS	3	1,4,8	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0026	Oiled 2	70°00'	125°00'	NS	VD	July 19	NS	NS	3	1,4,8	
(cont'd)	Control 1	70°00'	125°00'	NS	60-25 PNV and.or 240-50 PNV	July 20	NS	NS	NS	NS	Horizontal tows were also taken with these nets but no data are reported.
	Control 2	70°00'	125°00'	NS	60-25 PNV and.or 240-50 PNV	July 20	NS	NS	NS	NS	Horizontal tows were also taken with these nets but no data are reported.
	Oiled 1	70°00'	125°00'	NS	60-25 PNV and.or 240-50 PNV	July 20	NS	NS	NS	NS	Horizontal tows were also taken with these nets but no data are reported.
	Oiled 2	70°00'	125°00'	NS	60-25 PNV and.or 240-50 PNV	July 20	NS	NS	NS	NS	Horizontal tows were also taken with these nets but no data are reported.
75-0028	DH1(I)	70°02'	124°54'	NS	VD	May 13	1230, 1250	NS	2	4.5,6.5	VD samples were recovered through holes drilled in ice. PB samples were collected from melt ponds.
	DH1(I)	70°02'	124°54'	NS	VD	May 16	1315	NS	2	2.5,4.5	
	DH1(I)	70°02'	124°54'	NS	VD	May 26	0920, 0930	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	May 26	1010, 1020	NS	2	2.5,5	
	BH7(C)	70°02'	124°54'	NS	VD	May 26	1050, 1125	NS	2	2.5,5	
	BH7	70°02'	124°54'	NS	VD	May 28	NS	NS	1	2.5	
	DH2	70°02'	124°54'	NS	VD	May 28	NS	NS	1	2.5	
	DH1(I)	70°02'	124°54'	NS	VD	May 29	0920, 0925	NS	2	2.5,5	
	BH7(C)	70°02'	124°54'	NS	VD	May 29	0945, 0950	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	May 29	1005, 1010	NS	2	2.5,5	
	BH6(C)	70°02'	124°54'	NS	VD	May 29	1030, 1040	NS	2	2.5,5	
	BH7(C)	70°02'	124°54'	NS	VD	May 30	1530	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	May 30	1555, 1600	NS	2	2.5,5	
	CH1(I)	70°02'	124°54'	NS	VD	May 30	1625, 1635	NS	2	2.5,5	
	DH1(I)	70°02'	124°54'	NS	VD	May 30	1650, 1655	NS	2	2.5,5	
	DH1(I)	70°02'	124°54'	NS	VD	June 01	1700, 1705	NS	2	2.5,5	
	CH1(I)	70°02'	124°54'	NS	VD	June 01	1800, 1805	NS	2	2.5,5	
	BH7(C)	70°02'	124°54'	NS	VD	June 01	1905, 1915	NS	2	2.5,5	
	BH6(C)	70°02'	124°54'	NS	VD	June 01	2115, 2125	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	June 01	2210	NS	2	2.5,5	
	DH1(I)	70°02'	124°54'	NS	VD	June 04	1630	NS	3	2.5,5,8	
	CH1(I)	70°02'	124°54'	NS	VD	June 04	1730	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	June 04	1830	NS	2	2.5,5	
	AMP #1	70°02'	124°54'	NS	PB	June 04	2155	NS	1	0.58	
	AMP #2	70°02'	124°54'	NS	PB	June 04	2155	NS	1	0.25	
	AMP #3	70°02'	124°54'	NS	PB	June 04	NS	NS	1	0	
	AMP #4	70°02'	124°54'	NS	PB	June 04	NS	NS	1	0	
	BH7	70°02'	124°54'	NS	VD	June 04	NS	NS	1	2.5	
	BH7(C)	70°02'	124°54'	NS	VD	June 06	0950	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	June 06	1050, 1225	NS	2	2.5,5	
	CH1(I)	70°02'	124°54'	NS	VD	June 06	1235	NS	2	2.5,5	
	DH1(I)	70°02'	124°54'	NS	VD	June 06	1500	NS	2	2.5,5	
	DH2	70°02'	124°54'	NS	VD	June 08	NS	NS	1	5	
	CH1(I)	70°02'	124°54'	NS	VD	June 09	0200	NS	2	2.5,5	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0028 (cont'd)	DH1(I)	70°02'	124°54'	NS	VD	June 09	0230	NS	2	2.5,5	
	SH2(O)	70°02'	124°54'	NS	VD	June 09	0300	NS	2	2.5,5	
	NW6(C)	70°02'	124°54'	NS	VD	June 09	0330	NS	2	2.5,5	
	SH1(O)	70°02'	124°54'	NS	VD	June 09	2255	NS	2	2.5,5	
	NW2(O)	70°02'	124°54'	NS	VD	June 10	0015	NS	2	2.5,5	
	NW4(O)	70°02'	124°54'	NS	VD	June 10	0120	NS	2	2.5,5	
	DH2(I)	70°02'	124°54'	NS	VD	June 10	0205	NS	2	2.5,5	
	BH7(C)	70°02'	124°54'	NS	VD	June 11	1725	NS	2	2.5,5	
	FH1(C)	70°02'	124°54'	NS	VD	June 11	1940	NS	2	2.5,5	
	BH2(C)	70°02'	124°54'	NS	VD	June 11	2000	NS	2	2.5,5	
	BH8(C)	70°02'	124°54'	NS	VD	June 11	2030	NS	2	2.5,5	
	DA1	70°02'	124°54'	NS	PB	June 14	1300	NS	1	0.5	
	DA2	70°02'	124°54'	NS	PB	June 14	1320	NS	1	0.125	
	UA2	70°02'	124°54'	NS	PB	June 14	1320	NS	1	0.25	
	UA1	70°02'	124°54'	NS	PB	June 14	1330	NS	1	0.325	
	DA3	70°02'	124°54'	NS	PB	June 14	1447	NS	1	0.5	
	UA3	70°02'	124°54'	NS	PB	June 14	1515	NS	1	0.375	
	UA4	70°02'	124°54'	NS	PB	June 14	1515	NS	1	0.5	
	DA4	70°02'	124°54'	NS	VD	June 14	1530	NS	1	0	
	BH7	70°02'	124°54'	NS	VD	June 16	1740	NS	2	2,5	
	CH1(I)	70°02'	124°54'	NS	VD	June 16	1815	NS	2	2,5	
	SH2(O)	70°02'	124°54'	NS	VD	June 16	1842	NS	2	2,5	
	NW6(O)	70°02'	124°54'	NS	VD	June 16	1919	NS	2	2,5	
	SH1(O)	70°02'	124°54'	NS	VD	June 16	1949	NS	2	2,5	
	IS1(I)	70°02'	124°54'	NS	VD	June 16	2000	NS	2	2,5	
	NW4(O)	70°02'	124°54'	NS	VD	June 16	2110	NS	2	2,5	
	NW2(O)	70°02'	124°54'	NS	VD	June 16	2255	NS	2	2,5	
	DH2(O)	70°02'	124°54'	NS	VD	June 16	2340	NS	2	2,5	
	BH7(C)	70°02'	124°54'	NS	VD	June 22	1600	NS	2	2,5	
	BH4(C)	70°02'	124°54'	NS	VD	June 22	1845	NS	2	2,5	
	FH1(C)	70°02'	124°54'	NS	VD	June 22	1930	NS	2	2,5	
	CH1(I)	70°02'	124°54'	NS	VD	June 22	2040	NS	2	2,5	
	SH1(O)	70°02'	124°54'	NS	VD	June 22	2125	NS	2	2,5	
	IS2(I)	70°02'	124°54'	NS	VD	June 22	2250	NS	2	2,5	
	NW2(O)	70°02'	124°54'	NS	VD	June 22	2325	NS	2	2,5	
	7(O)	70°02'	124°54'	NS	VD	June 25	0030, 0145	NS	2	2,5	
	BH7(C)	70°02'	124°54'	NS	VD	June 25	1800	NS	2	2,5	
	II(C)	70°02'	124°54'	NS	VD	June 25	1935	NS	4	0,1.5,3,5	
	CH1(I)	70°02'	124°54'	NS	VD	June 25	2030	NS	2	2,5	
	SH2(O)	70°02'	124°54'	NS	VD	June 25	2115	NS	2	2,5	
	NW2(O)	70°02'	124°54'	NS	VD	June 25	2345	NS	2	2,5	
75-0051	75-006	69°22'	133°13.7'	2.5	NS PN	July 07	NS	NS	NS	NS	For all stations: no data available; sample location unknown.
	75-005	69°22.7'	133°12.5'	2	NS PN	July 07-08	NS	NS	NS	NS	
	75-008	69°22.9'	133°14.6'	2	NS PN	July 10	NS	NS	NS	NS	
	75-007	69°23.8'	133°09.7'	2	NS PN	July 11	NS	NS	NS	NS	
	75-009	69°23.6'	133°07.1'	1.5	NS PN	July 11	NS	NS	NS	NS	
	75-012	69°21.7'	133°39.2'	5	NS PN	July 19	NS	NS	NS	NS	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
75-0051 (cont'd)	75-013	69°20.2'	133°40.8'	7	NS PN	July 19	NS	NS	NS	NS	
	75-015	69°19.9'	133°43.2'	4.5	NS PN	July 20	NS	NS	NS	NS	
	75-014	69°19.5'	133°44.4'	4.5	NS PN	July 20-21	NS	NS	NS	NS	
	75-016	69°39.3'	132°23.9'	3.5	NS PN	July 26	NS	NS	NS	NS	
	75-017	69°38.2'	132°27.8'	3.5	NS PN	July 26	NS	NS	NS	NS	
	75-018	69°38.8'	132°32.8'	3.5	NS PN	July 26	NS	NS	NS	NS	
	75-019	69°40.2'	132°42.2'	2	NS PN	July 29	NS	NS	NS	NS	
	75-020	69°41.6'	132°24.1'	2.5	NS PN	July 29	NS	NS	NS	NS	
	75-023	69°42.4'	132°09.2'	3.5	NS PN	Aug. 04	NS	NS	NS	NS	
	75-104B	69°24.1'	133°00'	3.8	NS PN	Aug. 04	NS	NS	NS	NS	
	75-025	69°45.2'	132°14.5'	3	NS PN	Aug. 05	NS	NS	NS	NS	
	75-026	69°43.8'	132°12.1'	2.5	NS PN	Aug. 05	NS	NS	NS	NS	
	75-028	69°40.9'	132°08.3'	4	NS PN	Aug. 08	NS	NS	NS	NS	
	75-029	69°43.2'	132°14'	2	NS PN	Aug. 08	NS	NS	NS	NS	
	75-030	69°54.1'	131°07.9'	4.5	NS PN	Aug. 15	NS	NS	NS	NS	
	75-031	69°56.5'	131°09.9'	6.5	NS PN	Aug. 15	NS	NS	NS	NS	
	75-032	70°00.7'	131°09.6'	7.3	NS PN	Aug. 16	NS	NS	NS	NS	
	75-033	69°58.2'	131°17.4'	5	NS PN	Aug. 16	NS	NS	NS	NS	
	75-034	69°59.9'	131°04.8'	7.5	NS PN	Aug. 17	NS	NS	NS	NS	
76-0003	Isserk a.i.s.	69°56'	134°25'	NS	PB	Aug. 14	NS	NS	1	0	a.i.s. = artificial island site
	Isserk H-47 i.s.	69°56'	134°25'	NS	156-50 WNV	Aug. 14	NS	NS	5	11.7-0	i.s. = island site
	Isserk 2.0N	69°56'	134°25'	NS	PB	Aug. 14	NS	NS	1	0	
	Isserk 1.0E	69°56'	134°25'	NS	PB	Aug. 14	NS	NS	1	0	
	Isserk H47 N2.0	69°56'	134°25'	NS	156-50 WNH	Aug. 15	NS	NS	5	0	Stn. no. perhaps should be 2.0N.
	Isserk H47 2.0N	69°56'	134°25'	NS	156-50 WNV	Aug. 15	NS	NS	5	NS	
	Isserk H47 1.0E	69°56'	134°25'	NS	156-50 WNH; 156-50 WNV	Aug. 15	NS	NS	5; 1	0; 12-0	
	Kannerk s.	70°00'	131°12'	NS	PB	Aug. 16	NS	NS	1	0	s = site
	Kannerk G-42 i.s.	70°00'	131°12'	NS	156-50 WNV	Aug. 16	NS	NS	1	11.4-0	
	Kannerk 1.5 SW	70°00'	131°12'?	NS	PB	Aug. 16	NS	NS	1	0	
	Kannerk G-42 a.i.s.	70°00'	131°13'	NS	156-50 WNH	Aug. 16	NS	NS	5	0	
	Kannerk G-42 SE	70°00'	131°13'	NS	156-50 WNV	Aug. 16	NS	NS	5	9.5-0	
	Isserk NN	69°59'	134°23'	NS	PB	Aug. 30	NS	NS	1	0	
	Isserk NN-NNE	69°59'	134°23'	NS	156-50 WNV	Aug. 30	NS	NS	5	16-0	
	Isserk NNE	69°59'	134°23'	NS	PB	Aug. 30	NS	NS	1	0	
	Isserk NNE-NN	69°59'	134°23'	NS	156-50 WNV	Aug. 30	NS	NS	5	18.5-0	
	Nipterk p.i.s.	69°46'	135°28'	NS	PB; 156-50 WNV	Sep. 01	NS	NS	1; 5	0; 10.8-0	p.i.s. = proposed island site
	Igalak p.i.s.	69°44'	135°13'	NS	PB; 156-50 WNV	Sep. 01	NS	NS	1; 5	0; 8.3-0	
	Kadluk p.i.s.	69°49'	136°10'	NS	PB; 156-50 WNV	Sep. 02	NS	NS	1; 5	0; 16.9-0	
	Pakteq p.i.s.	69°39'	136°25'	NS	PB; 156-50 WNV	Sep. 02	NS	NS	1; 5	0; 12.3-0	
	NS p.a.s. adj. to Minuk blk	69°47'	136°45'	NS	PB; 156-50 WNV	Sep. 03	NS	NS	1; 5	0; 21.5-0	
	Minuk p.i.s.	69°43'	136°28'	NS	PB; 156-50 WNV	Sep. 05	NS	NS	1; 5	0; 14.8-0	
	Ernerk p.i.s.	69°49'	133°03'	NS	PB; 156-50 WNV	Sep. 07	NS	NS	1; 5	0; 10.4-0	
76-0004	Control area	69°43.35'	132°37.64'	NS	PB; 156-10 MNH	July 17?	NS	NS; 1.5	5; 5	0; 1	All stations are within 1.5 miles of the dredge site
	Construction area	69°43.35'	132°37.64'	NS	PB; 156-10 MNH	July 17?	NS	NS; 1.5	5; 5	0; 1	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
76-0004 (cont'd)	Control stn. 1.5 NNW	69°43.35'	132°37.64'	NS	PB	Sep. 01?	NS	NS	5	0	(69°43.35' N, 132°37.64'W) Sampling dates are reported in the text as July 18 and Aug. 31 and on the phytoplankton data sheets as July 17 and Sep. 01.
	Plume stn. 0.125 NE	69°43.35'	132°37.64'	NS	PB	Sep. 01?	NS	NS	5	0	
76-0005	Control area 1.5N	69°48'	133°45.5'	NS	PB; 156-10 MNH	July 31	NS	NS; 1.5	5; 5	0; 1	All stations are within 1.5 miles of the Arnak L-30 artificial island site (69°48'N, 133°45.5'W).
	Plume area	69°48'	133°45.5'	NS	PB; 156-10 MNH	July 31	NS	NS; 1.5	5; 5	0; 1	
	Plume area	69°48'	133°45.5'	NS	PB	Aug. 05	NS	NS	5	0	
	Control area 1.5N	69°48'	133°45.5'	NS	PB	Aug. 05	NS	NS	5	0	
	Plume area 0.125 SW	69°48'	133°45.5'	NS	PB	Aug. 11	NS	NS	5	0	
	Control area 1.5N	69°48'	133°45.5'	NS	PB	Aug. 11	NS	NS	5	0	
	Control area	69°48'	133°45.5'	NS	156-10 MNH or 156-50 WNH	Aug. 16	NS	1.5	1?	1?	The Miller net was lost on Aug. 16 and replaced by the Wisconsin net. However, the report does not indicate clearly in which net the zooplankton used in the grazing experiment were collected.
	Plume area	69°48'	133°45.5'	NS	156-10 MNH or 156-50 WNH	Aug. 16	NS	1.5	1?	1?	
	Control area 1.5N	69°48'	133°45.5'	NS	PB	Sep. 06	NS	NS	5	0	
	Plume area 0.3 SE	69°48'	133°45.5'	NS	PB	Sep. 06	NS	NS	5	0	
76-0036	HB-1	68°52.62'	133°26.02'	5.5	VD; 80-12.7 BRNV	June 24	1810?	NS	2; 1	1.5,4; 5.5-0	For all stations: time zone not specified. Therefore, time sample collected is assumed to be MST and, for this inventory, has been adjusted to GMT.
	HB-2	68°53.33'	133°28.03'	17.7	VD; 80-12.7 BRNV	June 24	2215?; 2200?	NS	3; 1	1,1.5,6; 16.7-0	
	HB-2	68°53.33'	133°28.03'	10	80-12.7 BRNV	July 02	1910?	NS	1	10-0	
	HB-3	68°51.9'	133°33.72'	NS	VD	July 02	NS	NS	1	1	
	HB-3	68°51.9'	133°33.72'	13	80-12.7 BRNV	July 03	0030?	NS	1	12-0	
	HB-2	68°53.33'	133°28.03'	28.5	80-12.7 BRNV	July 31	2250?	NS	1	27.5-0	
	HB-3	68°51.9'	133°33.72'	20.5	80-12.7 BRNV	July 31	2400?	NS	1	19.5-0	
	HB-2	68°53.33'	133°28.03'	26.5	80-12.7 BRNV	Aug. 10	1845?	NS	1	25.5-0	
	HB-3	68°51.9'	133°33.72'	21	80-12.7 BRNV	Aug. 10	2115?	NS	1	20-0	
	HB-3	68°51.9'	133°33.72'	22.5	80-12.7 BRNV	Sep. 11	2215?	NS	1	22-0	
	HB-2	68°53.33'	133°28.03'	25	80-12.7 BRNV	Sep. 11	2400?	NS	1	24.5-0	
	HB-3	68°51.9'	133°33.72'	19.5	80-12.7 BRNV	Sep. 19	2100?	NS	1	18.5-0	
	HB-2	68°53.33'	133°28.03'	25.2	80-12.7 BRNV	Sep. 20	0100?	NS	1	24.5-0	
	HB-2	68°53.33'	133°28.03'	25.2	80-12.7 BRNH	Sep. 21	1800?	60	1	NS	
77-0002	6	69°43.2'	132°33.7'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 16?	NS	NS	1; 2?	0.5; NS	Specific sampling dates estimated from concurrent studies. Number of zooplankton samples collected not specified in text of report; report tables not available for review.
	8	69°42.1'	132°34.8'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 16?	NS	NS	1; 2?	0.5; NS	
	5	69°43.3'	132°36.5'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 17?	NS	NS	1; 2?	0.5; NS	
	7	69°43.1'	132°38.2'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 17?	NS	NS	1; 2?	0.5; NS	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
77-0002 (cont'd)	9	69°39'	132°40.5'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 18?	NS	NS	1; 2?	0.5; NS	
	10	69°38.9'	132°37.4'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 18?	NS	NS	1; 2?	0.5; NS	
	11	69°40'	132°28.4'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 19?	NS	NS	1; 2?	0.5; NS	
	12	69°39.2'	132°27.6'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 19?	NS	NS	1; 2?	0.5; NS	
	3	69°44.5'	132°21.1'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 21?	NS	NS	1; 2?	0.5; NS	
	4	69°46.3'	132°14'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 21?	NS	NS	1; 2?	0.5; NS	
	2	69°42'	132°16.2'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 22?	NS	NS	1; 2?	0.5; NS	
	1	69°44.3'	132°30.6'	NS	KB; NS-12 WNV and.or 392-60 FNH	July 23?	NS	NS	1; 2?	0.5; NS	
	5	69°43.3'	132°36.5'	NS	KB; 392-60 FNH	Aug. 27?	NS	NS	1; 1?	0.5; NS	
	7	69°43.1'	132°38.2'	NS	KB; 392-60 FNH	Aug. 27?	NS	NS	1; 1?	0.5; NS	
	6	69°43.2'	132°33.7'	NS	KB; 392-60 FNH	Aug. 28?	NS	NS	1; 1?	0.5; NS	
	8	69°42.1'	132°34.8'	NS	KB; 392-60 FNH	Aug. 28?	NS	NS	1; 1?	0.5; NS	
	1	69°44.3'	132°30.6'	NS	KB; 392-60 FNH	Aug. 30?	NS	NS	1; 1?	0.5; NS	
	2	69°42'	132°16.2'	NS	KB; 392-60 FNH	Aug. 30?	NS	NS	1; 1?	0.5; NS	
	3	69°44.5'	132°21.1'	NS	KB; 392-60 FNH	Aug. 30?	NS	NS	1; 1?	0.5; NS	
	4	69°46.3'	132°14'	NS	KB; 392-60 FNH	Aug. 30?	NS	NS	1; 1?	0.5; NS	
	9	69°39'	132°40.5'	NS	KB; 392-60 FNH	Aug. 31?	NS	NS	1; 1?	0.5; NS	
	10	69°38.9'	132°37.4'	NS	KB; 392-60 FNH	Aug. 31?	NS	NS	1; 1?	0.5; NS	
	11	69°40'	132°28.4'	NS	KB; 392-60 FNH	Sep. 02?	NS	NS	1; 1?	0.5; NS	
	12	69°39.2'	132°27.6'	NS	KB; 392-60 FNH	Sep. 02?	NS	NS	1; 1?	0.5; NS	
77-0003	N-3	71°15.5'- 71°17.8'	127°03.4'- 127°04.9'	NS	NS N	Aug. 08?	NS	NS	1	0	Towed 2.5 nm. Date estimated from other tow dates.
	S-1	71°13.7'- 71°18.8'	127°01.8'- 127°05.1'	NS	NS N	Aug. 08?	NS	NS	1	NS	Towed 4.6 nm. Date estimated from other tow dates.
	M-3	71°17.7'- 71°14.3'	124°29.5'- 124°32.6'	NS	75-NS MNH and.or 243-NS MNH	Aug. 08	NS	NS	1	40-50	Towed 3.7 nm
	N-1	70°28.2'- 70°31.5'	127°09.8'- 127°06'	NS	NS N	Aug. 09?	NS	NS	1	0	Towed 3.9 nm. Date estimated from other tow dates.
	M-1	70°25.9'- 70°32.3'	127°11.2'- 127°05.1'	NS	75-NS MNH and.or 243-NS MNH	Aug. 09	NS	NS	1	40-50	Towed 6.7 nm
	N-2	70°42.7'- 70°46.4'	126°58.9'- 126°58.3'	NS	NS N	Aug. 09?	NS	NS	1	0	Towed 3.6 nm. Date estimated from other tow dates.
	M-2	70°41.5'- 70°47.2'	126°58.9'- 126°57.8'	NS	75-NS MNH and.or 243-NS MNH	Aug. 09	NS	NS	1	40-50	Towed 5.5 nm
	N-4	71°17.9'- 71°15.1'	124°29.8'- 124°32.8'	NS	NS N	Aug. 12?	NS	NS	1	0	Towed 3.2 nm. Date estimated from other tow dates.
	M-4	70°39.8'- 70°31.7'	125°10.9'- 125°20.3'	NS	75-NS MNH and.or 243-NS MNH	Aug. 12	NS	NS	1	40-50	Towed 8.8 nm

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED Date	Hour (GMT)	Interval (min.)	NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
77-0003 (cont'd)	N-5	70°38.7'- 70°33.8'	125°13.3'- 125°18.6'	NS	NS N	Aug. 12	NS	NS	1	0	Towed 5.3 nm
	M-5	70°30.5'- 70°32.5'	125°34.2'- 125°49.2'	NS	75-NS MNH and.or 243-NS MNH	Aug. 12	NS	NS	1	40-50	Towed 5.4 nm
	N-6	70°30.8'- 70°32.5'	125°34.2'- 125°47'	NS	NS N	Aug. 12?	NS	NS	1	0	Towed 4.7 nm. Date estimated from other tow dates.
	M-6	70°33.6'- 70°35.1'	125°59.1'- 126°12.8'	NS	75-NS MNH and.or 243-NS MNH	Aug. 12?	NS	NS	1	40-50	Towed 4.8 nm. Date estimated from other tow dates.
	N-7	70°33.6'- 70°34.9'	125°58.9'- 126°11.2'	NS	NS N	Aug. 12	NS	NS	1	0	Towed 4.3 nm
	M-7	71°36.7'- 71°30.6'	126°27.8'- 126°27.7'	NS	75-NS MNH and.or 243-NS MNH	Aug. 13	NS	NS	1	40-50	Towed 6.8 nm
	N-8	71°36.5'- 71°32.1'	126°28.6'- 126°27.9'	NS	NS N	Aug. 13?	NS	NS	1	0	Towed 4.4 nm. Date estimated from other tow dates.
	M-8	71°26'- 71°22.1'	126°26'- 126°25.6'	NS	75-NS MNH and.or 243-NS MNH	Aug. 13	NS	NS	1	40-50	Towed 4.1 nm
	N-9	71°25.1'- 71°22.9'	126°26.8'- 126°26.5'	NS	NS N	Aug. 13?	NS	NS	1	0	Towed 2.4 nm. Date estimated from other tow dates.
	M-9	71°16.8'- 71°06.5'	126°25.4'- 126°25.8'	NS	75-NS MNH and.or 243-NS MNH	Aug. 13	NS	NS	1	40-50	Towed 10.2 nm
	N-10	71°15.7'- 71°07.4'	126°26.5'- 126°26.7'	NS	NS N	Aug. 13?	NS	NS	1	0	Towed 8.4 nm. Date estimated from other tow dates.
	M-10	71°03.2'- 70°55.5'	126°26.1'- 126°27.2'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 13	NS	NS	1	40-50	Towed 7.5 nm
	N-11	71°01.8'- 70°52.8'	126°27.3'- 126°29.4'	NS	NS N	Aug. 13?	NS	NS	1	0	Towed 9 nm. Date estimated from other tow dates.
	M-11	70°50'- 70°44'	126°29.5'- 126°32'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 13?	NS	NS	1	40-50	Towed 6.3 nm. Date estimated from other tow dates.
	N-12	70°50'- 70°44.9'	126°30.2'- 126°32.7'	NS	NS N	Aug. 13	NS	NS	1	0	Towed 5.6 nm
	M-12	70°48.9'- 70°44.5'	125°04'- 124°42.2'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 15	NS	NS	1	40-50	Towed 8.7 nm
	N-13	70°48.3'- 70°45.2'	124°58.7'- 124°42.3'	NS	NS N	Aug. 15?	NS	NS	1	0	Towed 5.8 nm. Date estimated from other tow dates.
	M-13	70°41.8'- 70°36.2'	124°29.6'- 124°04.3'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 15	NS	NS	1	40-50	Towed 10.2 nm
	N-14	70°42.2'- 70°39.8'	124°29.4'- 124°19'	NS	NS N	Aug. 16?	NS	NS	1	0	Towed 4.3 nm. Date estimated from other tow dates.
	M-14	70°26.8'- 70°22.5'	123°21.9'- 123°10.4'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 16	NS	NS	1	40-50	Towed 5.2 nm
	N-15	70°26.3'- 70°23.3'	123°22.3'- 123°11.8'	NS	NS N	NS	NS	NS	1	0	Towed 4.7 nm
	M-15	70°21.6'- 70°06.4'	123°06.7'- 122°23'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 21.2 nm
	N-16	70°21.2'- 70°06.9'	123°06.6'- 122°23.1'	NS	NS N	Aug. 21?	NS	NS	1	0	Towed 20 nm. Date estimated from other tow dates.
	M-16	69°55'- 70°02.8'	121°07.2'- 121°11.3'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 21	NS	NS	1	40-50	Towed 7.8 nm

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
77-0003 (cont'd)	N-17	69°56.4'-70°01.8'	121°08.4'-121°11.6'	NS	NS N	NS	NS	NS	1	0	Towed 5.5 nm
	M-17	70°04.9'-70°13.8'	121°13'-121°20.5'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 9.3 nm
	N-18	70°05.5'-70°11.1'	121°13.9'-121°18.7'	NS	NS N	NS	NS	NS	1	0	Towed 6 nm
	M-18	70°15.4'-70°23.6'	121°21.9'-121°30.6'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 8.7 nm
	N-19	70°15.6'-70°22.7'	121°23'-121°30.4'	NS	NS N	NS	NS	NS	1	0	Towed 7.5 nm
	M-19	70°25.4'-70°32.2'	121°33'-121°42.7'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 7.4 nm
	N-20	70°25.2'-70°31.8'	121°33.9'-121°42'	NS	NS N	NS	NS	NS	1	0	Towed 7 nm
	M-20	71°22.4'-71°11.1'	119°04.1'-119°08.4'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 11.1 nm
	N-21	71°21.1'-71°12.2'	119°05.3'-119°08.8'	NS	NS N	Aug. 29?	NS	NS	1	0	Towed 9.1 nm. Date estimated from other tow dates.
	M-21	71°08.7'-70°58.7'	119°09.8'-119°12.8'	NS	75 MNH-NS and.or 243-NS MNH	Aug. 29	NS	NS	1	40-50	Towed 9.9 nm
	N-22	71°08.6'-70°59.8'	119°10.5'-119°13.6'	NS	NS N	NS	NS	NS	1	0	Towed 9.3 nm
	M-22	70°56'-70°46.3'	119°13.7'-119°16.3'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 9.5 nm
	N-23	70°55.7'-70°47.3'	119°14.7'-119°17.3'	NS	NS N	NS	NS	NS	1	0	Towed 8.3 nm
	M-23	70°44.8'-70°34'	119°16.7'-119°18.4'	NS	75 MNH-NS and.or 243-NS MNH	NS	NS	NS	1	40-50	Towed 10 nm
	N-24	70°44.9'-70°36.1'	119°17.8'-119°19.1'	NS	NS N	Sep. 05?	NS	NS	1	0	Towed 9 nm. Date estimated from other tow dates.
	M-24	70°58.9'-70°50.1'	120°38.1'-120°33.5'	NS	75 MNH-NS and.or 243-NS MNH	Sep. 05	NS	NS	1	40-50	Towed 9 nm
	N-25	70°58.3'-70°50.6'	120°37.9'-120°35'	NS	NS N	Sep. 05?	NS	NS	1	0	Towed 7.8 nm. Date estimated from other tow dates.
	M-25	70°48.8'-70°39.7'	120°33.2'-120°28.5'	NS	75 MNH-NS and.or 243-NS MNH	Sep. 05	NS	NS	1	40-50	Towed 9.3 nm
	N-26	70°47.7'-70°40.9'	120°33.4'-120°30'	NS	NS N	Sep. 05?	NS	NS	1	0	Towed 7.5 nm. Date estimated from other tow dates.
	M-26	70°39'-70°34.7'	120°28'-120°25.9'	NS	75 MNH-NS and.or 243-NS MNH	Sep. 05	NS	NS	1	40-50	Towed 4.6 nm
	N-27	70°38.4'-70°35.5'	120°28.6'-120°27.3'	NS	NS N	Sep. 07?	NS	NS	1	0	Towed 3 nm. Date estimated from other tow dates.
	M-27	70°24.1'-70°40.4'	121°30.6'-121°52.4'	NS	75-NS MNH and.or 243-NS MNH	Sep. 07	NS	NS	1	40-50	Towed 17.8 nm
	M-28	70°41.3'-70°55.6'	121°54.5'-122°17.4'	NS	75-NS MNH and.or 243-NS MNH	Sep. 07	NS	NS	1	40-50	Towed 16.4 nm
	M-29	70°47.9'-70°57.9'	124°29.8'-123°39'	NS	75-NS MNH and.or 243-NS MNH	Sep. 09	NS	NS	1	40-50	Towed 19.5 nm

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
77-0003 (cont'd)	M-30	70°57.9'- 71°00.2'	123°38.3'- 123°21.1'	NS	75-NS MNH and.or 243-NS MNH	Sep. 10	NS	NS	1	40-50	Towed 6.2 nm
77-0009	022-1	69°56.3'?	133°20.8'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,12; B-0; 0	All lats., longs. estimated from mapped positions in report.
	022-2	69°56.5'?	133°20.5'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	022-3	69°57.3'?	133°19.5'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	022-4	69°59.9'?	133°16.2'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,15; B-0; 0	
	112-1	69°56'?	133°20.3'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	112-2	69°55.9'?	133°19.8'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	112-3	69°55.6'?	133°17.9'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	112-4	69°54.4'?	133°10.5'?	HS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	202-1	69°55.9'?	133°21.2'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	202-2	69°55.75'?	133°21.5'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	202-3	69°55.05'?	133°22.3'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,11; B-0; 0	
	202-4	69°52.2'?	133°25.5'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,9; B-0; 0	
	292-1	69°56.2'?	133°21.5'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	292-2	69°56.25'?	133°22'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 2; 1	1,5,11; B-0; 0	
	292-3	69°56.55'?	133°24'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,10; B-0; 0	
	292-4	69°57.75'?	133°32'?	NS	VD; 150-50 PNV; 571-100 PNH	July	NS	NS	3; 3; 1	1,5,13; B-0; 0	
	022-1	69°56.3'?	133°20.8'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	022-2	69°56.5'?	133°20.5'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	022-3	69°57.3'?	133°19.5'?	NS	150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 1	B-0; 0	
	022-4	69°59.9'?	133°16.2'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,15; B-0; 0	
	112-1	69°56'?	133°20.3'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	112-2	69°55.9'?	133°19.8'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; B-0; 0	
	112-3	69°55.6'?	133°17.9'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,11; B-0; 0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
77-0009 (cont'd)	112-4	69°54.4'?	133°10.5'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 2; 1	1,5,15; 8-0; 0	
	202-1	69°55.9'?	133°21.2'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; 8-0; 0	
	202-2	69°55.75'?	133°21.5'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 2	1,5,12; 8-0	
	202-3	69°55.05'?	133°22.3'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,11; 8-0; 0	
	202-4	69°52.2'?	133°25.5'?	NS	VD; 150-50 PNV	Aug.	NS	NS	3; 3	1,5,15; 8-0	
	292-1	69°56.2'?	133°21.5'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; 8-0; 0	
	292-2	69°56.25'?	133°22'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,12; 8-0; 0	
	292-3	69°56.55'?	133°24'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,11; 8-0; 0	
	292-4	69°57.75'?	133°32'?	NS	VD; 150-50 PNV; 571-100 PNH	Aug.	NS	NS	3; 3; 1	1,5,15; 8-0; 0	
78-0002	1NW	70°10.6'?	130°58.9'?	NS	NS	July 17	NS	NS	1	0-30	All lats., longs. estimated from mapped positions in report.
	1	70°23.4'?	135°49.9'?	NS	NS	July 18	1430	NS	1	0-25	
	7SE	70°10'?	130°58'?	NS	NS	July 18	1630	NS	1	0-25	
78-0018	Centre	69°32.5'	136°12.1'	NS	NS SNV	July 20	NS	NS	1	18-0	
79-0009	Nerlerk M98	70°30'	133°30'	NS	NS NB	Sep. 09	NS	NS	5	1,2,3,5,10	
79-0026	A	70°01.6'?	131°13'?	NS	NS corer	Dec. 13	NS	NS	2	NA	All lats., longs. estimated from mapped positions in report.
	B	70°01.6'?	131°13'?	NS	NS corer	Dec. 13	NS	NS	2	NA	
80-0016	A	70°01.6'?	131°13'?	NS	NS corer	Apr. 08	NS	NS	1	NA	All lats., longs. estimated from mapped positions in report.
	D	70°01.6'?	131°13'?	NS	NS corer	Apr. 08	NS	NS	1	NA	
	E	70°01.6'?	131°13'?	NS	Chainsaw (block)	Apr. 08	NS	NS	1	NA	
	D	70°01.6'?	131°13'?	NS	NS corer	June 19	NS	NS	1	NA	
80-0025	1	69°56'	134°23'	12	500-61 PNV	Aug. 14	1400	NS	1	12-0	
	2	70°00'	134°16'	17	500-61 PNV	Aug. 14	1500	NS	1	17-0	
	3	70°01'	134°04'	20	500-61 PNV	Aug. 14	1600	NS	1	20-0	
	9	69°07'	138°00'	5.5	500 MNH	Aug. 20	1900	10	6	3	
	10	69°09'	138°00'	20	500 MNH	Aug. 20	2100	10	12	3,10	
	9	69°07'	138°00'	6.8	500-61 PNV	Aug. 21	2000	NS	2	6.8-0	
	10	69°09'	138°00'	17	500-61 PNV	Aug. 21	2200	NS	2	17-0	
	4	69°56'	134°55'	17	500-61 PNV	Aug. 24	1830	NS	1	17-0	
	5	70°04'	134°19'	25.7	500-61 PNV	Aug. 26	1800	NS	1	25.7-0	
	6	69°59'	133°56'	19.3	500-61 PNV	Aug. 27	0400	NS	1	19.3-0	
	7	69°59'	134°18'	13.5	500-61 PNV	Aug. 27	0500	NS	1	13.5-0	
	8	69°56'	134°18'	10.2	500-61 PNV	Aug. 27	0600	NS	1	10.2-0	
	9	69°07'	138°00'	7.5	240-50 PNV	Sep. 06	2100	NS	1	7.5-0	
	10	69°09'	138°00'	16.3	240-50 PNV	Sep. 06	2300	NS	1	16.3-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
81-0003	1	70°00.9'?	134°19.3'?	NS	NBS	Mar. 07	NS	NS	3	3,10,16	All lats., longs. estimated from mapped positions in report.
	2	70°01.1'?	134°18.7'?	NS	NBS	Mar. 08	NS	NS	3	3,10,17	
	3	70°01'?	134°19.5'?	NS	NBS	Mar. 09	NS	NS	3	3,10,16	
	3	70°01'?	134°19.5'?	NS	NBS	May 13	NS	NS	3	2.5,10,17	
	2	70°01.1'?	134°18.7'?	NS	NBS	May 14	NS	NS	3	2.5,10,17	
	1	70°00.9'?	134°19.3'?	NS	NBS	May 16	NS	NS	3; 1	2.5,10,15	
	1	70°00.9'?	134°19.3'?	NS	NBS	Oct. 10	NS	NS	4	0,5,10,18	
	2	70°01.1'?	134°18.7'?	NS	NBS	Oct. 10	NS	NS	4	0,5,10,15	
	3	70°01'?	134°19.5'?	NS	NBS	Oct. 10	NS	NS	4	0,5,10,17	
81-0014	Camp 205, McKinley Bay	69°57.5'	131°10'	NS	253 75 PNV	July 04	NS	NS	1	8-0	Sample unanalysed; preserved in 4-10% F buffered with Borax; resides in EPS, N.W.T., warehouse.
81-0015	Issungnak 1	70°00.9'?	134°19.3'?	16	NB	Mar. 07	0930	NS	3	1.5,8,15	Lat., long. estimated from mapped position in report.
	Issungnak 2	70°01.1'?	134°18.7'?	35	NB	Mar. 08	NS	NS	3	1.5,8,16	Lat., long. estimated from mapped position in report.
	Issungnak 3	70°01'?	134°19.5'?	16	NB	Mar. 09	NS	NS	3	1.5,8,15	Lat., long. estimated from mapped position in report.
	McKinley Bay 1	70°01.6'?	131°13'?	13	NB	Apr. 21	NS	NS	5	1,3,5,8,10	Lat., long. estimated from mapped position in report.
	Issungnak 2	70°01.1'?	134°18.7'?	16.5	NB	May 14	NS	NS	3	1,8,15.5	Lat., long. estimated from mapped position in report.
	Issungnak 1	70°00.9'?	134°19.3'?	15.5	NB; 158-22.5 PNV; S corer; ice scraper	May 16	0715,1100, 1600,2115; 0715?; NS; NS	NS	9; 2; 2; 1	1,8,14.5; 15-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 180 cm; divided into 18 sections, 12 retained (0-10, 20-30, 40-50, 50-60, 60-70, 80-90, 100-110, 120-130, 140-150, 150-160, 160-170, 170-180 cm).
	McKinley Bay 1	70°01.6'?	131°13'?	14	NB; 158-22.5 PNV; S corer	May 17	NS	NS	5; 2; 2	1,3,5,8,13; 14-0; NA	Lat., long. estimated from mapped position in report; ice thickness = 165 cm; divided into 9 sections, all retained.
	Issungnak 1	70°00.9'?	134°19.3'?	16	NB; 158-22.5 PNV; S corer; ice scraper	May 29	1930; 1930; NS; NS	NS	3; 2; 6; 1	1,8,15; 15-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 145 cm; ice sections retained = 0-20, 80-100, 135-145 cm.
	Issungnak 1	70°00.9'?	134°19.3'?	16	NB; 158-22.5 PNV; S corer; ice scraper	June 16	1530; 1530; NS; NS	NS	3; 4; 6; 1	1,8,15; 4-0, 15-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 145 cm, ice sections retained = 0-20, 60-80, 125-135, 135-145 cm.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
81-0015 (cont'd)	McKinley Bay 1	70°01.6'?	131°13'?	19	NB; 158-22.5 PNV	June 20	NS	NS	8; 2	1,3,5,7,10, 12,15,18; 18.8-0	Lat., long. estimated from mapped position in report.
	Issungnak 1	70°00.9'?	134°19.3'?	19	NB; 158-22.5 PNO	July 25	2200; 2200; NS	NS	3; 4	1,7,18; 6-0, 18-0	Lat., long. estimated from mapped position in report.
	Issungnak 1	70°00.9'?	134°19.3'?	19, 18	NB	July 26	0300, 1530	NS	6	1,7,17,18	Lat., long. estimated from mapped position in report.
	Issungnak 1	70°00.9'?	134°19.3'?	18	NB	Sep. 26	2300	NS	3	1,7,17	Lat., long. estimated from mapped position in report.
	Issungnak 1	70°00.9'?	134°19.3'?	18	NB	Oct. 10	1645	NS	4	1,5,10,17	Lat., long. estimated from mapped position in report.
	Issungnak 2	70°01.1'?	134°18.7'?	16	NB	Oct. 10	1745	NS	4	1,5,10,17	Lat., long. estimated from mapped position in report.
	Issungnak 3	70°01'?	134°19.5'?	18	NB	Oct. 10	1845	NS	4	1,5,10,17	Lat., long. estimated from mapped position in report.
81-0027	N-1	69°50.7'	133°20'	14	505-60 PNH	Aug. 01	0210	5	9	0,5,10	
	L-1	69°55.6'	133°22.8'	20	505-60 PNH	Aug. 01	1650	5	9	0,5,15	
	N-4	69°36.4'	133°04.8'	5	505-60 PNH	Aug. 10	1945	5	3	0	
	N-5	70°10'	133°28.5'	40	505-60 PNH	Aug. 11	1900	5	9	0,20,30	
	N-6	70°05.2'	133°25.6'	32	505-60 PNH	Aug. 11	2225	5	9	0,15,20	
	N-7	70°00.5'	133°23.3'	27	505-60 PNH	Aug. 12	0200	5	9	0,15,20	
	L-1	69°55.6'	133°22.8'	20	505-60 PNH	Aug. 12	0700	5	9	0,5,10	
	N-1	69°50.7'	133°20'	14	505-60 PNH	Aug. 12	0720	5	6	0,5	
	N-8	70°02.8'	134°30'	20	505-60 PNH	Aug. 14	2330	5	9	0	
	N-9	70°00.3'	134°43'	26	505-60 PNH	Aug. 19	0706	5	9	0,2,15	
	N-10	70°02.6'	134°48.5'	27	505-60 PNH	Aug. 19	2255	5	9	0,5,15	
	N-11	70°08.3'	134°38.3'	30	505-60 PNH	Aug. 24	2335	5	9	0,5,15	
	N-12	69°57.3'	133°52'	17	505-60 PNH	Aug. 25	0425	5	9	0,5,15	
	N-13	69°52'	134°49'	10.5	505-60 PNH	Aug. 25	2110	5	3	10	
	L-1	69°55.6'	133°22.8'	20	505-60 PNH	Sep. 06	1923	5	9	0,5,15	
	N-2	70°02'	133°55'	23	240-50? PNV	Sep. 06	2030	NS	1	23-0	
81-0101	1	70°00.9'?	134°19.3'?	NS	NBS	July 24	NS	NS	4	0,5,10,18	All lats., longs. estimated from mapped positions in report.
	2	70°01.1'?	134°18.7'?	NS	NBS	July 24	NS	NS	4	0,5,10,18	
	3	70°01'?	134°19.5'?	NS	NBS	July 24	NS	NS	3	1,8,18	
82-0032	7	69°23.7'?	133°00.5'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	All lats., longs. estimated from mapped positions in report.
	8	69°25'?	133°00'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	
	9	69°25.85'?	132°57.02'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	
	10	69°26.22'?	132°56.8'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	
	11	69°26.85'?	132°57.13'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	
	13	69°24.62'?	132°59.35'?	NS	750-50 WNH	July 11-12	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	17	69°24.88'?	133°00.85'?	NS	750-50 WNH	July 11-12	NS	5?	2?	1-0	
	7	69°23.7'?	133°00.5'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	8	69°25'?	133°00'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
82-0032 (cont'd)	9	69°25.85'?	132°57.02'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	10	69°26.22?	132°56.8'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	11	69°26.85'?	132°57.13'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	12	69°27.12'?	133°00'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	13	69°24.62'?	132°59.35'?	NS	750-50 WNH	July 13-19	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	14	69°25.2'?	132°58.35'?	NS	750-50 WNH	July 13-19	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	15	69°26.37'?	132°58.2'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	17	69°24.88'?	133°00.85'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	18	69°25.65'?	132°55.75'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	19	69°28.02'?	132°57.1'?	NS	750-50 WNH	July 13-19	NS	5?	2?	1-0	
	7	69°23.7'?	133°00.5'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	8	69°25'?	133°00'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	9	69°25.85'?	132°57.02'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	10	69°26.22?	132°56.8'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	11	69°26.85'?	132°57.13'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	12	69°27.12'?	133°00'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	13	69°24.62'?	132°59.35'?	NS	750-50 WNH	July 20-26	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	14	69°25.2'?	132°58.35'?	NS	750-50 WNH	July 20-26	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	15	69°26.37'?	132°58.2'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	17	69°24.88'?	133°00.85'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	18	69°25.65'?	132°55.75'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	19	69°28.02'?	132°57.1'?	NS	750-50 WNH	July 20-26	NS	5?	2?	1-0	
	14	69°25.2'?	132°58.35'?	NS	750-50 WNH	July 27 - Aug. 02	NS	5?	2?	1-0	
	7	69°23.7'?	133°00.5'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	8	69°25'?	133°00'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	9	69°25.85'?	132°57.02'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	10	69°26.22?	132°56.8'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	11	69°26.85'?	132°57.13'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	12	69°27.12'?	133°00'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	14	69°25.2'?	132°58.35'?	NS	750-50 WNH	Aug. 03-09	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	15	69°26.37'?	132°58.2'?	NS	750-50 WNH	Aug. 03-09	NS	5?	2?	1-0	
	7	69°23.7'?	133°00.5'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
	8	69°25'?	133°00'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
	9	69°25.85'?	132°57.02'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
	10	69°26.22?	132°56.8'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
	11	69°26.85'?	132°57.13'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
	12	69°27.12'?	133°00'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
82-0032 (cont'd)	14	69°25.2'?	132°58.35'?	NS	750-50 WNH	Aug. 10-16	NS	5?	8?	1-0; 3.9-2.5; 7.5-4; 10.5-7.6	
	15	69°26.37'?	132°58.2'?	NS	750-50 WNH	Aug. 10-16	NS	5?	2?	1-0	
82-0109	1	70°00.9'?	134°19.3'?	NS	NBS	Apr. 15	NS	NS	4	0,5,10,15	All lats., longs. estimated from mapped positions in report.
	2	70°01.1'?	134°18.7'?	NS	NBS	Apr. 16	NS	NS	4	0,5,10,16	
	3	70°01'?	134°19.5'?	NS	NBS	Apr. 16	NS	NS	4	0,5,10,16	
82-0110	Issungnak 1	70°00.9'?	134°19.3'?	17	NB; 158-22.5 PNV; S corer; ice scraper	Feb. 19, 19, 19-20; 19	1930; 1930; NS; NS	NS	3; 2; 4; 1	1,8,16; 17-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 90-94 cm; divided into 6 sections; all retained.
	Issungnak 3	70°01'?	134°19.5'?	17	NB; 158-22.5 PNV; S corer	Feb. 21	2300; 2300; NS	NS	3; 1; 2	1,8,16; 16-0; NA	Lat., long. estimated from mapped position in report; ice thickness = 80 cm; divided into 5 sections; all retained.
	Issungnak 2	70°01.1'?	134°18.7'?	17.5	NB; 158-22.5 PNV; S corer	Feb. 22	2300; 2300; NS	NS	3; 4; 4	1,8,16; 15-0; 16-0; NA	Lat., long. estimated from mapped position in report; ice thickness = 110 cm; divided into 6 sections; all retained.
	Issungnak 1	70°00.9'?	134°19.3'?	17	S corer	Apr. 13, 15	NS	NS	4	NA	Lat., long. estimated from mapped position in report; ice thickness = 145 cm; divided into 6 sections; all retained.
	Issungnak 1	70°00.9'?	134°19.3'?	17	NB; 158-22.5 PNV	Apr. 15	2200	NS	3; 2	1,8,16; 15-0	Lat., long. estimated from mapped position in report.
	Issungnak 2	70°01.1'?	134°18.7'?	17	NB; 158-22.5 PNV	Apr. 15	2400	NS	3; 2	1,8,16; 15-0	Lat., long. estimated from mapped position in report.
	Issungnak 3	70°01'?	134°19.5'?	17	S corer	Apr. 15	NS	NS	4	NA	Lat., long. estimated from mapped position in report; ice thickness = 133 cm; divided into 6 sections; all retained.
	Issungnak 2	70°01.1'?	134°18.7'?	17	S corer	Apr. 16	NS	NS	4	NA	Lat., long. estimated from mapped position in report; ice thickness = 160-165 cm; divided into 6 sections; all retained.
	Issungnak 3	70°01'?	134°19.5'?	17	NB; 158-22.5 PNV	Apr. 16	2200	NS	3; 2	1,8,16; 15-0	Lat., long. estimated from mapped position in report.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
82-0110 (cont'd)	Issungnak 3	70°01'?	134°19.5'?	18	NB; 158-22.5 PNV; S corer; ice scraper	May 12	2230; 2230; NS; NS	NS	3; 2; 4; 1	1,8,16; 16-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 148 cm; divided into 6 sections; all retained.
	Issungnak 1	70°00.9'?	134°19.3'?	17	NB; 158-22.5 PNV; S corer; ice scraper	May 13	1830; 1830; NS; NS	NS	3; 2; 4; 1	1,8,15; 15-0; NA; NA	Lat., long. estimated from mapped position in report; ice thickness = 155 cm; divided into 6 sections; all retained.
	McKinley Bay 1	70°01.6'?	131°13'?	NS	ice scraper	May 17	NS	NS	1	NA	Lat., long. estimated from mapped position in report.
	McKinley Bay 2	70°01.6'?	131°13'?	2	NB; S corer	May 17	NS	NS	1; 4	1; NA	Lat., long. estimated from mapped position in report; ice thickness = 194 cm; divided into 6 sections; all retained.
	Issungnak 1	70°00.9'?	134°19.3'?	16.5	S corer; ice scraper	June 10	NS	NS	4; 1	NA	Lat., long. estimated from mapped position in report; ice thickness = 160 cm; divided into 6 sections; all retained.
	Issungnak 3	70°01'?	134°19.5'?	NS	S corer	June 10	NS	NS	2	NA	Lat., long. estimated from mapped position in report; ice thickness = >300 cm; ice sections retained = 0-50, 50-100, 100-150 cm.
	Issungnak 1	70°00.9'?	134°19.3'?	16.5	NB; 158-22.5 PNV	June 11	0200	NS	3; 2;	1,8,15; 15-0	Lat., long. estimated from mapped position in report.
83-0054	2	69°27.13'	133°00.2'	NS	253-75 PNV	July 12	2001	NS	1	8-0	Samples unanalysed; preserved in 4-10% F with Rose Bengal, buffered with Borax; in glass mason jars with internal labels; reside in EPS, N.W.T., warehouse.
	5	69°57.8'	131°09.8'	11	253-75 PNV	July 15	1807	NS	1	11	
83-0050	1	70°05'	134°31'	NS	PB	Aug. 21	NS	NS	1	0	Sample container broken.
	2	70°06'	134°31'	NS	PB	Aug. 21	NS	NS	1	0	
	3	70°04'	134°25'	NS	PB	Aug. 21	NS	NS	1	0	
	4	70°06'	134°27'	NS	PB	Aug. 21	NS	NS	1	0	
	5	70°04'	134°21'	NS	PB	Aug. 21	NS	NS	1	0	
	6	70°05'	134°10'	NS	PB	Aug. 21	NS	NS	1	0	
	7	70°04'	134°05'	NS	PB	Aug. 21	NS	NS	1	0	
	8	69°56'	134°17'	NS	PB	Aug. 21	NS	NS	1	0	
	9	69°47'	134°07'	NS	PB	Aug. 21	NS	NS	1	0	
	10	69°36'	133°37'	NS	PB	Aug. 21	NS	NS	1	0	Sample container broken.

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
83-0058 (cont'd)	1	70°33'	133°14'	NS	PB	Aug. 23	NS	NS	1	0	
	2	70°30'	133°13'	NS	PB	Aug. 23	NS	NS	1	0	
	3	70°25'	133°13'	NS	PB	Aug. 23	NS	NS	1	0	
	4	70°20'	133°12'	NS	PB	Aug. 23	NS	NS	1	0	
	5	70°13'	133°11'	NS	PB	Aug. 23	NS	NS	1	0	
	6	70°08'	133°09'	NS	PB	Aug. 23	NS	NS	1	0	
	7	70°03'	133°05'	NS	PB	Aug. 23	NS	NS	1	0	
	8	69°56'	133°03'	NS	PB	Aug. 23	NS	NS	1	0	
	9	69°54'	133°03'	NS	PB	Aug. 23	NS	NS	1	0	
	10	69°46'	133°09'	NS	PB	Aug. 23	NS	NS	1	0	
	11	69°39'	133°10'	NS	PB	Aug. 23	NS	NS	1	0	
	12	69°34'	133°08'	NS	PB	Aug. 23	NS	NS	1	0	
	13	69°27'	133°04'	NS	PB	Aug. 23	NS	NS	1	0	
	14	69°27'	133°03'	NS	PB	Aug. 23	NS	NS	1	0	
	1	68°58'	133°10'	NS	PB	Oct. 09	NS	NS	1	0	
	2	69°00'	133°04'	NS	PB	Oct. 09	NS	NS	1	0	
	3	69°06'	132°46'	NS	PB	Oct. 09	NS	NS	1	0	
	4	69°09'	132°33'	NS	PB	Oct. 09	NS	NS	1	0	
	5	69°16'	132°16'	NS	PB	Oct. 09	NS	NS	1	0	
	6	69°24'	131°57'	NS	PB	Oct. 09	NS	NS	1	0	
	7	69°27'	131°48'	NS	PB	Oct. 09	NS	NS	1	0	
	8	69°30'	131°33'	NS	PB	Oct. 09	NS	NS	1	0	
	9	69°33'	131°23'	NS	PB	Oct. 09	NS	NS	1	0	
	10	69°32'	130°55'	NS	PB	Oct. 09	NS	NS	1	0	
	11	69°45'	130°14'	NS	PB	Oct. 09	NS	NS	1	0	
83-0059	1	69°23.5'?	133°37.5'?	NS	VD	Aug. 23-31	NS	NS	1	0.5-1	All lats., longs. estimated from mapped positions in report. Specific sampling dates unavailable from Albright.
	2	69°25.5'?	133°20'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,2	
	3	69°29'?	133°21'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,2,3	
	4	69°38'?	133°25'?	NS	VD	Aug. 23-31	NS	NS	3	0.1,0.5-1,2	
	5	69°44'?	133°25'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,2,5	
	6	69°48.5'?	133°25'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,2,5,9	
	7	69°50.5'?	133°32.5'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,2,5,9	
	8	70°09.5'?	133°32.5'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,5,20,40	
	9	70°36'?	133°32.5'?	NS	VD	Aug. 23-31	NS	NS	5	0.5-1,5,15, 25,50	
	10	70°59'?	133°47'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,≥30	
	11	70°58'?	134°06'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,5,20,50	
	12	70°35'?	134°42.5'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,3,40	
	13	70°08.5'?	135°10'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,35	
	14	69°55'?	136°07'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,3,20	
	15	69°43'?	137°16'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,3,20, ≥30	
	16	69°31'?	138°05'?	NS	VD	Aug. 23-31	NS	NS	4	0.5-1,3,20,50	
	17	69°21'?	137°55'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,3,30	
	18	69°13'?	137°48'?	NS	VD	Aug. 23-31	NS	NS	3	0.5-1,3,≥30	
	19	69°06'?	137°27'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,3	
	20	69°27'?	136°22.5'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,3	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
83-0059 (cont'd)	21	69°40.5'?	135°55'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,3	
	22	69°49'?	135°00'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,3	
	23	69°45.5'?	134°05'?	NS	VD	Aug. 23-31	NS	NS	2	0.5-1,3	
	24	69°36.5'?	133°42.5'?	NS	VD	Aug. 23-31	NS	NS	2	0.1,0.5-1	
	25	69°23'?	133°46.5'?	NS	VD	Aug. 23-31	NS	NS	1	0.5-1	
84-0033	Mason Bay	69°30.55'	134°08.25'	NS	bucket	Aug. 01	NS	NS	1	0	
	Mason Bay	69°30.55'	134°08.25'	NS	bucket	Aug. 02	NS	NS	1	0	
	Tuk Stn 8	69°26.18'	132°58.31'	NS	bucket	Aug. 02	NS	NS	1	0	
	Mason Bay	69°30.55'	134°08.25'	NS	bucket	Aug. 03	NS	NS	1	0	
	Hutchison Bay	69°45.45'	132°04'	NS	bucket	Aug. 03	NS	NS	1	0	
	Mason Bay	69°30.55'	134°08.25'	NS	bucket	Aug. 04	NS	NS	1	0	
85-0016	J	69°20.8'?	133°44'?	NS	VD; Craib corer	Aug. 26-27	NS	NS	1; 1	1; 0-0.3	All lats., longs., estimated from mapped positions in report. Specific sampling dates unavailable from Albright.
	K	69°25'?	133°35'?	NS	VD; Craib corer	Aug. 26-27	NS	NS	1; 1	1; 0-0.3	
	L	69°29'?	133°25'?	NS	VD; Craib corer	Aug. 26-27	NS	NS	1; 1	1; 0-0.3	
	M	69°31'?	133°16'?	NS	VD; Craib corer	Aug. 26-27	NS	NS	1; 1	1; 0-0.3	
85-0017	85100	69°30.81'	133°09.5'	4	500-NS N; 500-50 BNH	July 18	NS	15; 5	2; 2	0; 2	
	85102	69°28.1'	133°01.2'	3.5	500-NS N; 500-50 BNH	July 18	NS	15; 5	1; 2	0; 2	
	Tuk Harb S	69°25.55'	132°58'	25	83-50 BNV; 500-NS N; 500-50 BNH	July 19	NS	NS; 5; 15	1; 1; 1	NS; 0; NS	
	Tuk Harb N	69°26.15'	132°58.25'	20	500-NS N; 500-50 BNH	July 20	NS	5; 15	2; 2	0; 4.5,10.5	
	Tuk Harb S	69°25.55'	132°58'	25	500-NS N; 500-50 BNH	July 20	NS	5; 15	2; 2	0; 4.5,11.5	
	85102	69°28.1'	133°02'	4.5	500-NS N; 500-50 BNH	July 21	NS	5; 15	2; 2	0; 2.6	
	85103	69°37.78'	133°11.18'	5.6	500-NS N; 500-50 BNH	July 21	NS	5; 15	2; 2	0; 3	
	85104	69°37.1'	133°02.25'	6.3	83-50 BNO; 500-NS N; 500-50 BNH	July 22	NS	NS; 5; 15	2; 2; 2	6.3-0; 0; 3	
	85105	69°36.45'	132°58.9'	2	500-NS N	July 22	NS	5	4	0	
	85106	69°20.7'	138°43.2'	4	500-NS N; 500-50 BNH	July 24	NS	5; 15	2; 2	0; 4.5-5	
	85107	69°23.6'	138°44.5'	14.5	500-NS N; 500-50 BNH	July 24	NS	5; 15	2; 3	0; 4.5,10	
	85108	69°25'	138°47.9'	53	83-50 BNV; 500-NS N; 500-50 BNO	July 24	NS	NS; 5; 15	1; 2; 2	53-0; 0; NS	
	85109	69°27.7'	138°47.9'	69	83-50 BNV; 500-NS N; 500-50 BNO	July 24	NS	NS; 5; 15	2; 2; 2	69-0; 0; NS	
	85110	69°33.2'	139°01.5'	5.6	500-NS N; 500-50 BNH	July 25	NS	5; 15	2; 2	0; 3	
	85111	69°31.58'	138°58.9'	14.5	83-50 BNV; 500-NS N; 500-50 BNH	July 25	NS	NS; 5; 15	1; 2; 2	14.5-0; 0; 5, 10	
	85112	69°31.05'	138°55.5'?	52	83-50 BNV; 500-NS N; 500-50 BNO	July 25	NS	NS; 5; 15	1; 2; 2	52-0; 0; NS	Long. questioned by Hopky.
	85113	69°28.45'	138°48.75'?	64	83-50 BNV; 500-NS N; 500-50 BNO	July 25	NS	NS; 5; 15	1; 2; 2	64-0; 0; NS	Long. questioned by Hopky.
	85114	69°16.9'	138°20.2'	5.5	500-NS N; 500-50 BNH	July 26	NS	5; 15	2; 2	0; 3	
	85115	69°16.15'	138°16.6'	15	500-NS N; 500-50 BNH	July 26	NS	5; 15	2; 2	0; 5.5,10	
	85116	69°11.95'	137°41.53'	29.3	83-50 BNV; 500-NS N; 500-50 BNO	July 26	NS	NS; 5; 15	1; 2; 2	29.3-0; 0; NS	

TABLE 3 (cont'd)

DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
85-0017 (cont'd)	85117	69°18.8'	137°11'	15.2	83-50 BNV; 500-NS N; 500-50 BNH	July 26	NS	NS; 5; 15	1; 2; 2	15.2-0; 0; 4,12	
	Tuk Harb N	69°26.15'	132°58.25'	20	83-50 BNV; 500-NS N; 500-50 BNH	Aug. 02	NS	NS; 5; 15	1; 2; 2	20-0; 0; 4,12	
	Tuk Harb S	69°25.55'	132°58'	25	83-50 BNV; 500-NS N; 500-50 BNH	Aug. 02	NS	NS; 5; 15	1; 2; 2	25-0; 0; 4,12	
	85100	69°30.8'	133°09.5'	5	500-NS N; 500-50 BNH	Aug. 03	NS	5; 15	2; 2	0; 3	
	85118	69°40.5'	132°57.5'	6	500-NS N; 500-50 BNH	Aug. 03	NS	5; 15	2; 2	0; 4	
	85119	69°36.5'	133°00'	5	500-NS N; 500-50 BNH	Aug. 03	NS	5; 15	2; 2	0; 2-3	
	85102	69°28'	133°02'	4.5	500-NS N; 500-50 BNH	Aug. 04	NS	5; 15	2; 2	0; 3	
	85120	69°39.7'	138°42.5'	100	83-50 BNV; 500-NS N; 500-50 BNO	Aug. 05	NS	NS; 5; 15	1; 2; 2	100-0; 0; NS	
	85121	69°40.5'	138°52.5'	50	83-50 BNV; 500-NS N; 500-50 BNO	Aug. 05	NS	NS; 5; 15	1; 2; 2	50-0; 0; NS	
	85122	69°38.3'	138°54.7'	30	83-50 BNV; 500-NS N; 500-50 BNO	Aug. 05	NS	NS; 5; 15	1; 2; 2	30-0; 0; NS	
	85123	69°36.2'	138°50'	5	500-NS N; 500-50 BNH	Aug. 06	NS	5; 15	2; 2	0; 4,12	
	85124	69°46.5'	138°34.8'	150	500-50 BNO	Aug. 06	NS	15	2	NS	
	Tuk Harb N	69°26.15'	132°58.25'	20	83-50 BNV; 500-NS N; 500-50 BNH	Aug. 08	NS	NS; 5; 15	1; 2; 2	20-0; 0; 4,12	
	Tuk Harb S	69°25.55'	132°58'	25	83-50 BNV; 500-NS N; 500-50 BNH	Aug. 08	NS	NS; 5; 15	1; 2; 2	25-0; 0; 4,12	
	85125	69°39'	132°54.6'	3	500-NS N	Aug. 09	NS	5	4	0	
	85126	69°40.3'	132°56.5'	6	500-NS N; 500-50 BNH	Aug. 09	NS	5; 15	2; 2	0; 4	
	85127	69°37.3'	132°59.9'	2	500-NS N	Aug. 09	NS	5	4	0	
	85128	69°37.3'	133°02'	6	500-NS N; 500-50 BNH	Aug. 09	NS	5; 15	2; 2	0; 4	
	85129	69°37.2'	133°52.5'	2.5	500-NS N	Aug. 10	NS	5	4	0	
	85130	69°41.9'	133°41'	6	500-NS N; 500-50 BNH	Aug. 10	NS	5; 15	2; 2	0; 4	
	85131	69°46.2'	133°32'	9	500-NS N; 500-50 BNH	Aug. 10	NS	5; 15	2; 2	0; 4,7	
	85132	69°46.8'	134°24.2'	4	500-NS N; 500-50 BNH	Aug. 10	NS	5; 15	2; 2	0; 3	
	85133	69°52.6'	134°35.6'	10.5	500-NS N; 500-50 BNH	Aug. 11	NS	5; 15	2; 2	0; 4,7	
	85134	69°58.1'	134°36.3'	15	83-50 BNV; 500-NS N; 500-50 BNH; 500-50 BNO	Aug. 11	NS	NS; 5; 15; 15	1; 2; 2; 1	15-0; 0; 4,10; NS	
	85135	69°39.2'	135°30.1'	5	500-NS N; 500-50 BNH	Aug. 11	NS	5; 15	2; 2	0; 4	
	85136	69°37'	135°56.4'	6	500-NS N; 500-50 BNH	Aug. 12	NS	5; 15	2; 2	0; 4	
	85137	69°19.7'	136°37.4'	15	83-50 BNV; 500-NS N; 500-50 BNH; 500-50 BNO	Aug. 12	NS	NS; 5; 15; 15	1; 2; 2; 1	15-0; 0; 4,10; NS	
	85138	69°38.9'	133°03.3'	30	83-50 BNV; 500-NS N; 500-50 BNH; 500-50 BO	Aug. 12	NS	NS; 5; 15; 15	1; 2; 1; 2	30-0; 0; 10; NS	
	Tuk Harb N	69°26.15'	132°58.25'	20	500-NS N; 500-50 BNH	Sep. 02	NS	5; 15	2; 2	0; 3,12	
	Tuk Harb S	69°25.55'	132°58'	24.5	500-NS N; 500-50 BNH	Sep. 02	NS	5; 15	2; 2	0; 3,12?	Depth questioned by Hopky.
	85102	69°28'	138°02'	3.9	500-NS N; 500-50 BNH	Sep. 02	NS	5; 15	2; 2	0; 3	
	85200	69°33.2'	139°01.6'	5	500-NS N; 500-50 BNH	Sep. 07	NS	5; 15	2; 2	0; 3.5	
	85201	69°31.6'	138°59.8'	14	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 07	NS	NS; 5; 15	1; 2; 2	14-0; 0; 4,12	
	85202	69°31'	138°55.5'	40	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 07	NS	NS; 5; 15	1; 2; 2	40-0; 0; NS	

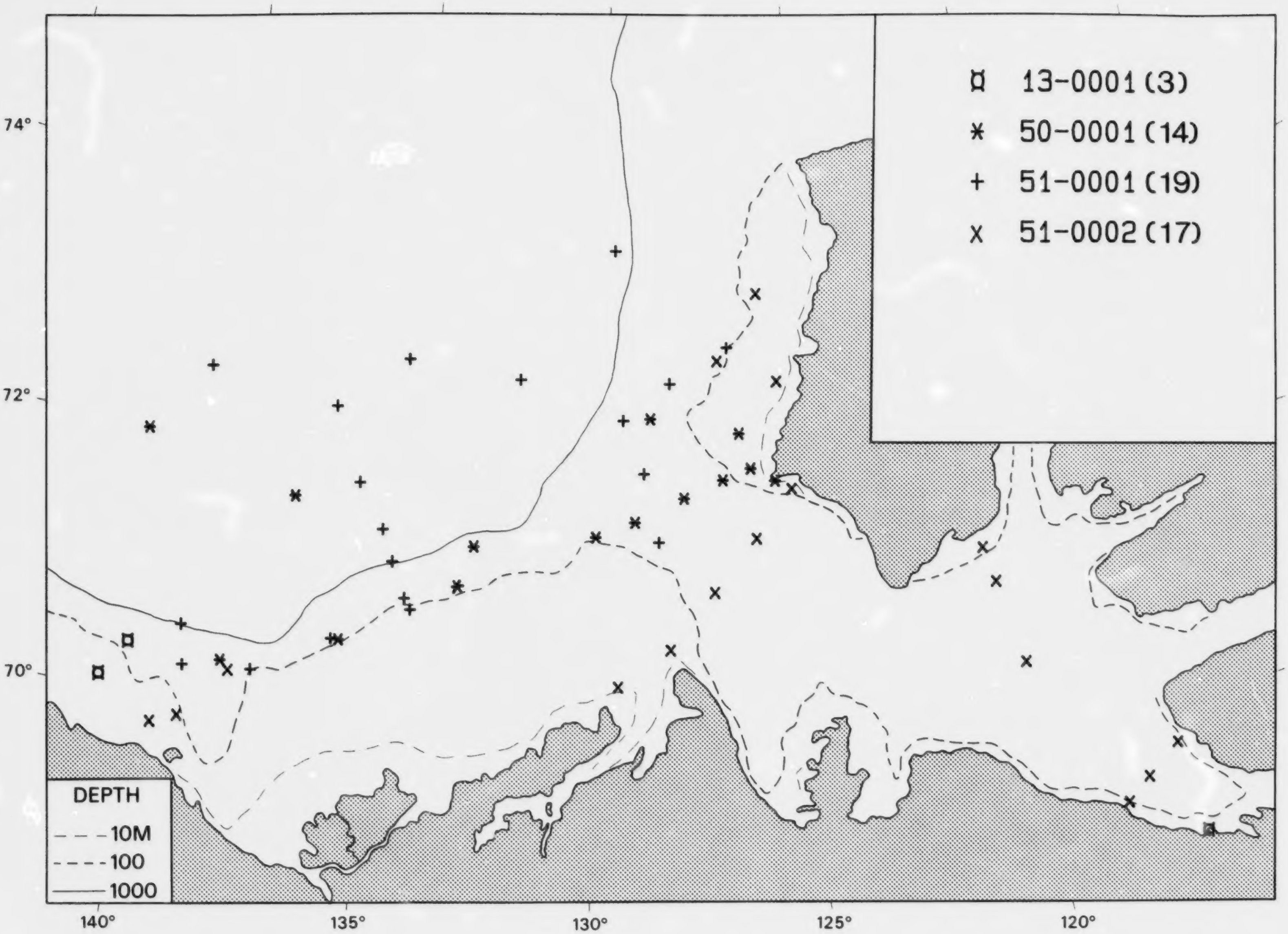
TABLE 3 (cont'd)

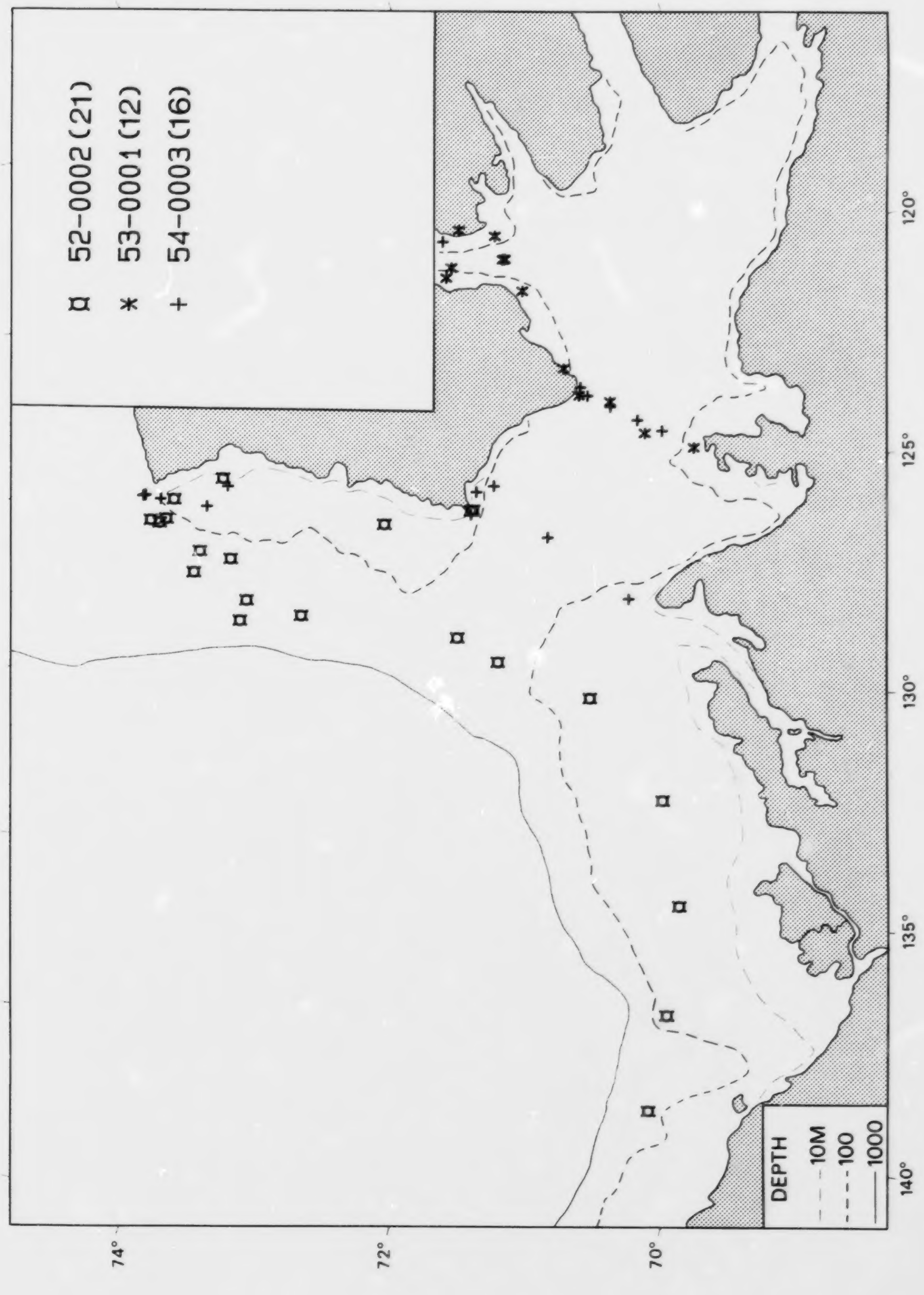
DATA SET I.D.	STN. NO./LOCATION	LATITUDE (°N)	LONGITUDE (°W)	STN. DEPTH (m)	GEAR TYPE	TIME SAMPLED			NO. SAMPLES	DEPTH SAMPLED (m)	REMARKS
						Date	Hour (GMT)	Interval (min.)			
85-0017 (cont'd)	85203	69°29.5'	138°49.1'	57	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 07	NS	NS; 5; 15	1; 2; 2	57-0; 0; NS	
	85204	69°35'	138°50'	6	500-NS N; 500-50 BNH	Sep. 08	NS	5; 15	2; 2	0; 4	
	85205	69°35.5'	138°47.4'	16.3	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 08	NS	NS; 5; 15	1; 2; 2	16.3-0; 0; 4,12	
	85206	69°37.3'	138°44.8'	50	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 08	NS	NS; 5; 15	1; 2; 2	50-0; 0; NS	
	85207	69°27.8'	138°47.8'	65	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 09	NS	NS; 5; 15	1; 2; 2	65-0; 0; NS	
	85208	69°25.2'	138°45.8'	24.5	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 09	NS	NS; 5; 15	1; 2; 2	24.5; 0; NS	
	85209	69°22.8'	138°43.4'	12	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 09	NS	NS; 5; 15	1; 2; 2	12-0; 0; 4,8	
	85210	69°20.3'	138°42.1'	4.8	500-NS N; 500-50 BNH	Sep. 09	NS	5; 15	2; 2	0; 3.8	
	85211	69°38.1'	138°35'	130	83-50 BNV; 500-NS N; 500-50 BNO	Sep. 10	NS	NS; 5; 15	1; 2; 2	130-0; 0; NS	
	85212	69°59.5'	131°10.2'	5.5	500-NS N; 500-50 BNH	Sep. 12	NS	5; 15	2; 2	0; 3.5	
	85213	70°04.6'	131°27.2'	15	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 12	NS	NS; 5; 15	1; 2; 2	15-0; 0; 4,12	
	85214	69°59.2'	132°00.9'	14.5	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 12	NS	NS; 5; 15	1; 2; 2	14.5-0; 0; 4.7,13.7	
	85215	69°47.4'	132°01.8'	5.8	500-NS N; 500-50 BNH	Sep. 12	NS	5; 15	2; 2	0; 4	
	85216	69°51.6'	132°30.5'	10.5	83-50 BNV; 500-NS N; 500-50 BNH	Sep. 12	NS	NS; 5; 15	1; 2; 2	10.5-0; 0; 4,9	
	85217	69°39.2'	132°56.2'	5.5	500-NS N; 500-50 BNH	Sep. 12	NS	5; 15	2; 2	0; 3	

9. MAPS

Station locations or transects for the data sets reviewed in this inventory are plotted on a series of maps. The main base map is a Lambert Conformal Conic projection with standard parallels of 71°N and 73°N, drawn to a scale of 1:4,680,000. When stations are too concentrated to assess readily, a larger-scale map is included. The number of stations plotted for each data set is indicated in brackets.

The maps are organized chronologically by data set number. An attempt was made to prepare a map for each data set. However, for data sets 60-0067, 77-0020, 79-0037, 81-0031 and 85-0007, this was precluded by insufficient information.


13-0001 (3)
50-0001 (14)
51-0001 (19)
51-0002 (17)
DEPTH
10M
100
1000
74°
72°
70°
140°
135°
130°
125°
120°



¤ 52-0002 (21)
✱ 53-0001 (12)
+ 54-0003 (16)
DEPTH
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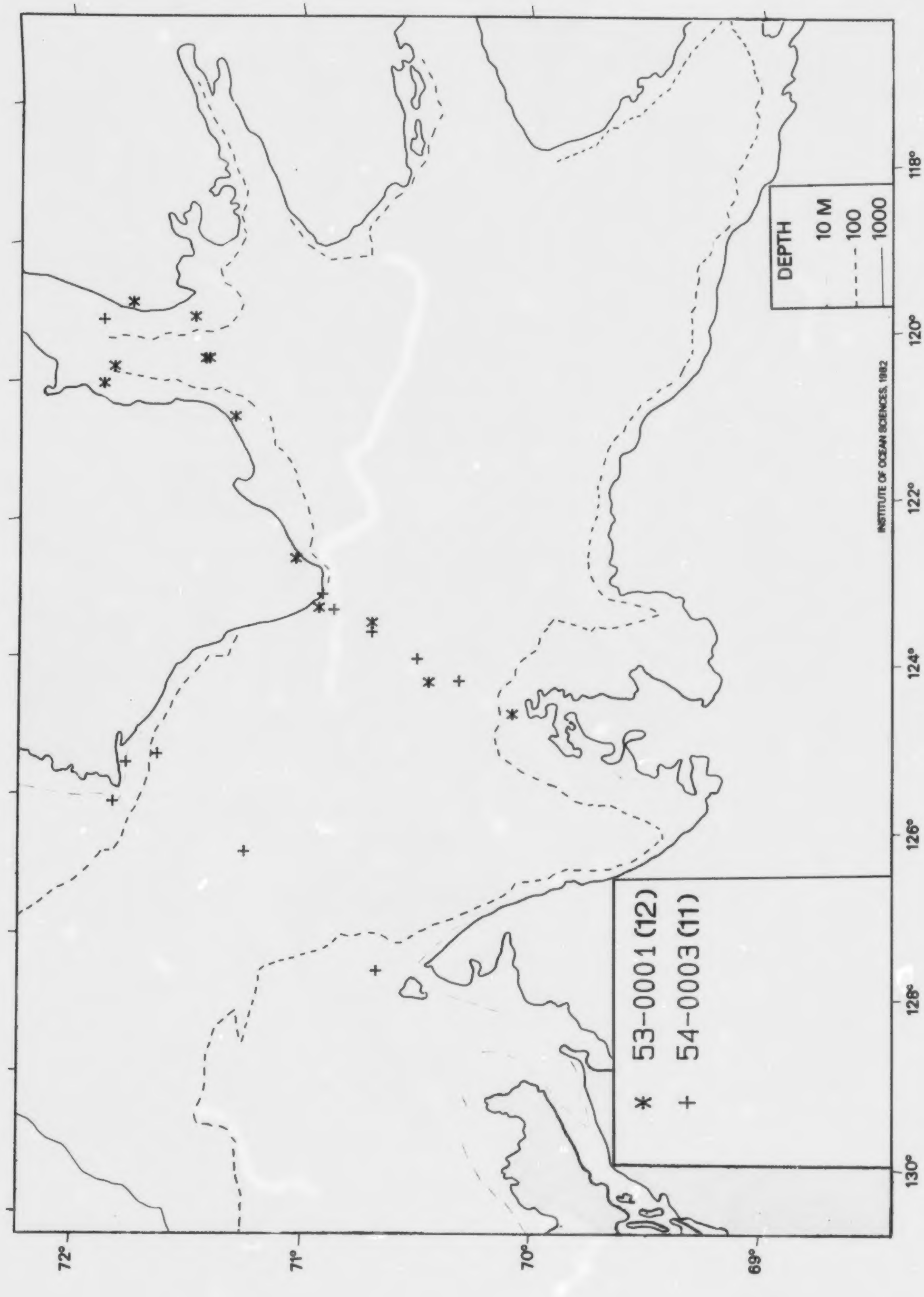
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* 53-0001 (12)
+ 54-0003 (11)

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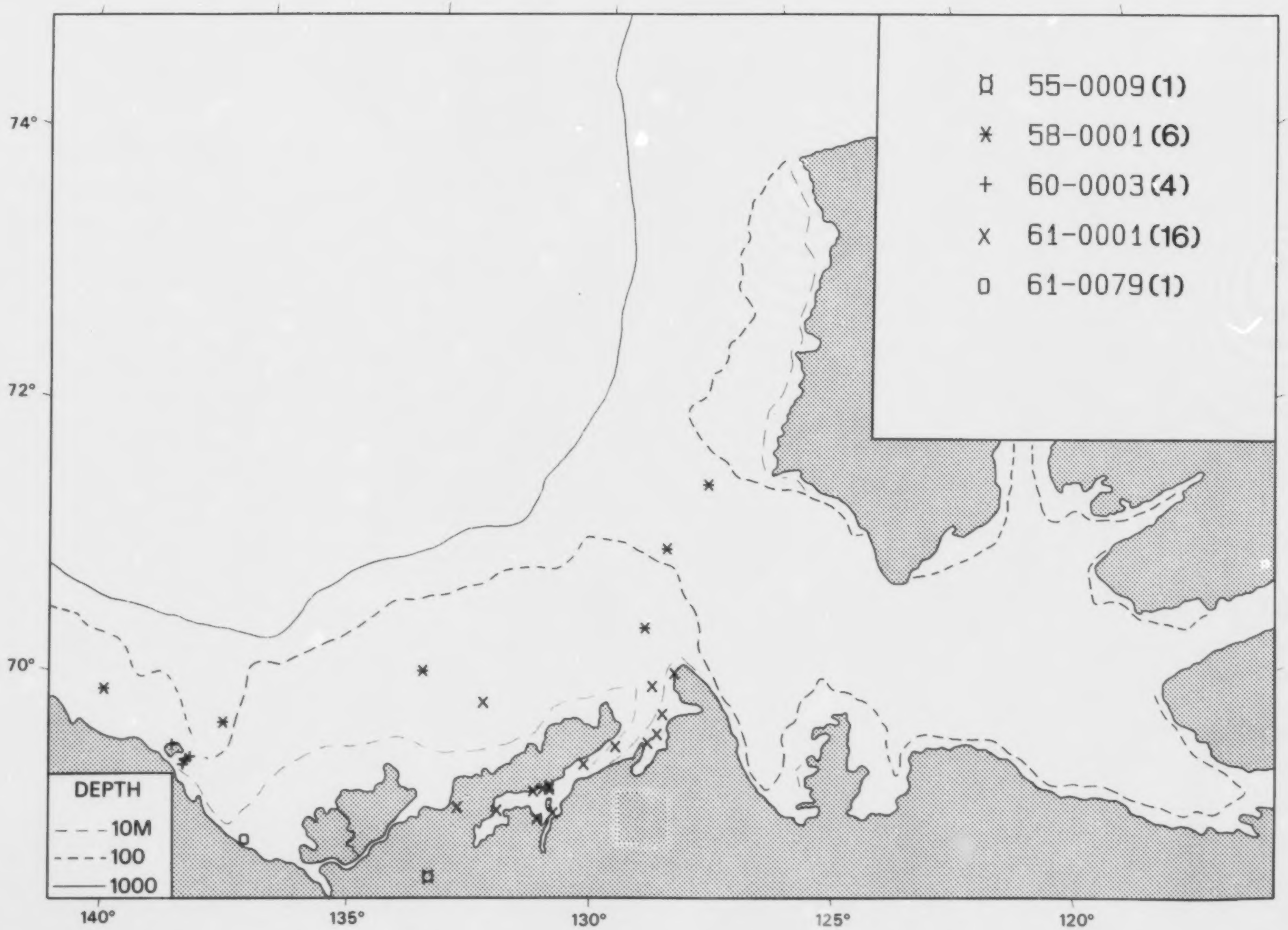
¤ 55-0009(1)
✱ 58-0001(6)
+ 60-0003(4)
x 61-0001(16)
o 61-0079(1)
DEPTH
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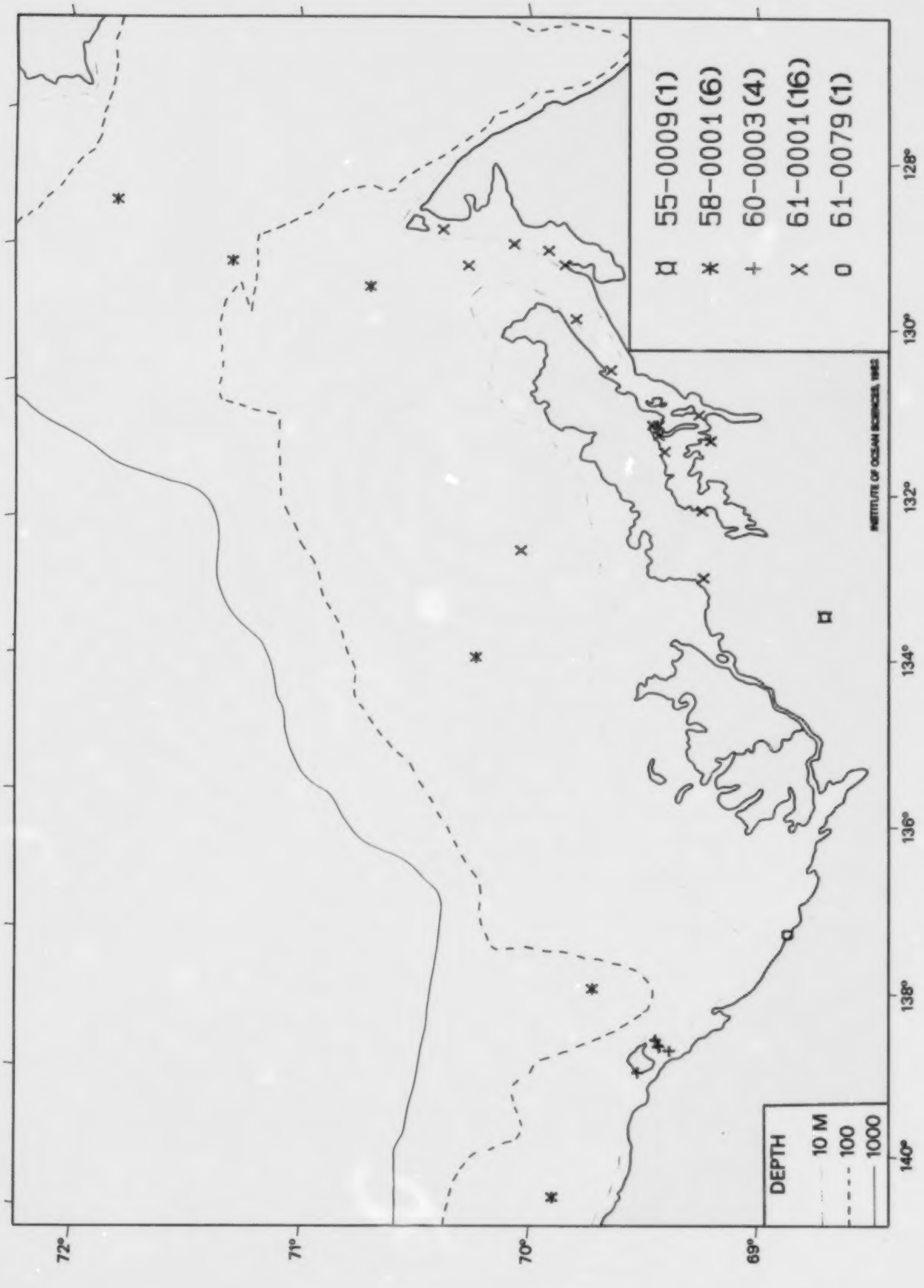
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58-0001 (6)
60-0003 (4)
61-0001 (16)
61-0079 (1)
DEPTH
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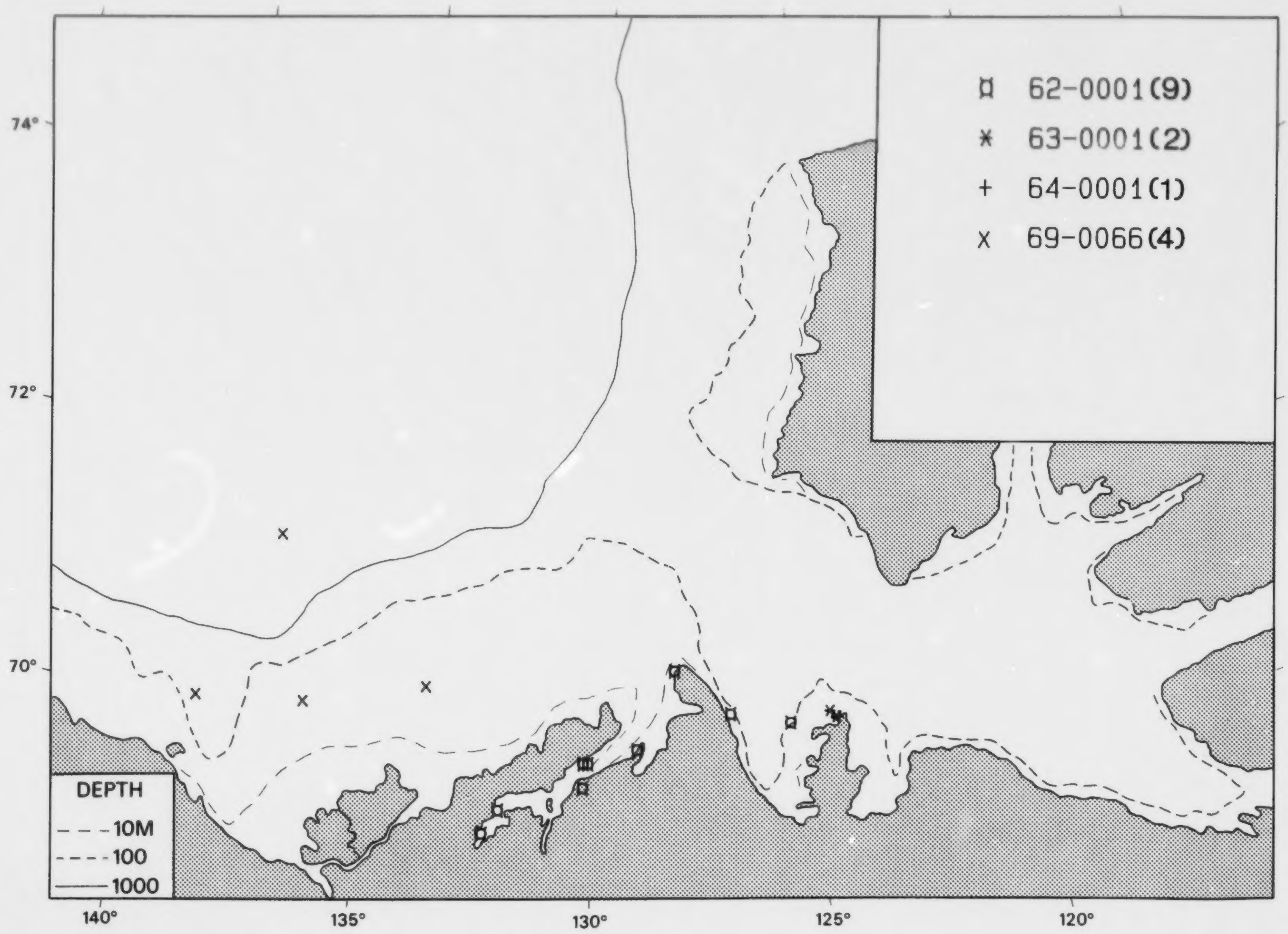
INSTITUTE OF OCEAN SCIENCES, 1982



¤ 62-0001(9)
✳ 63-0001(2)
+ 64-0001(1)
x 69-0066(4)
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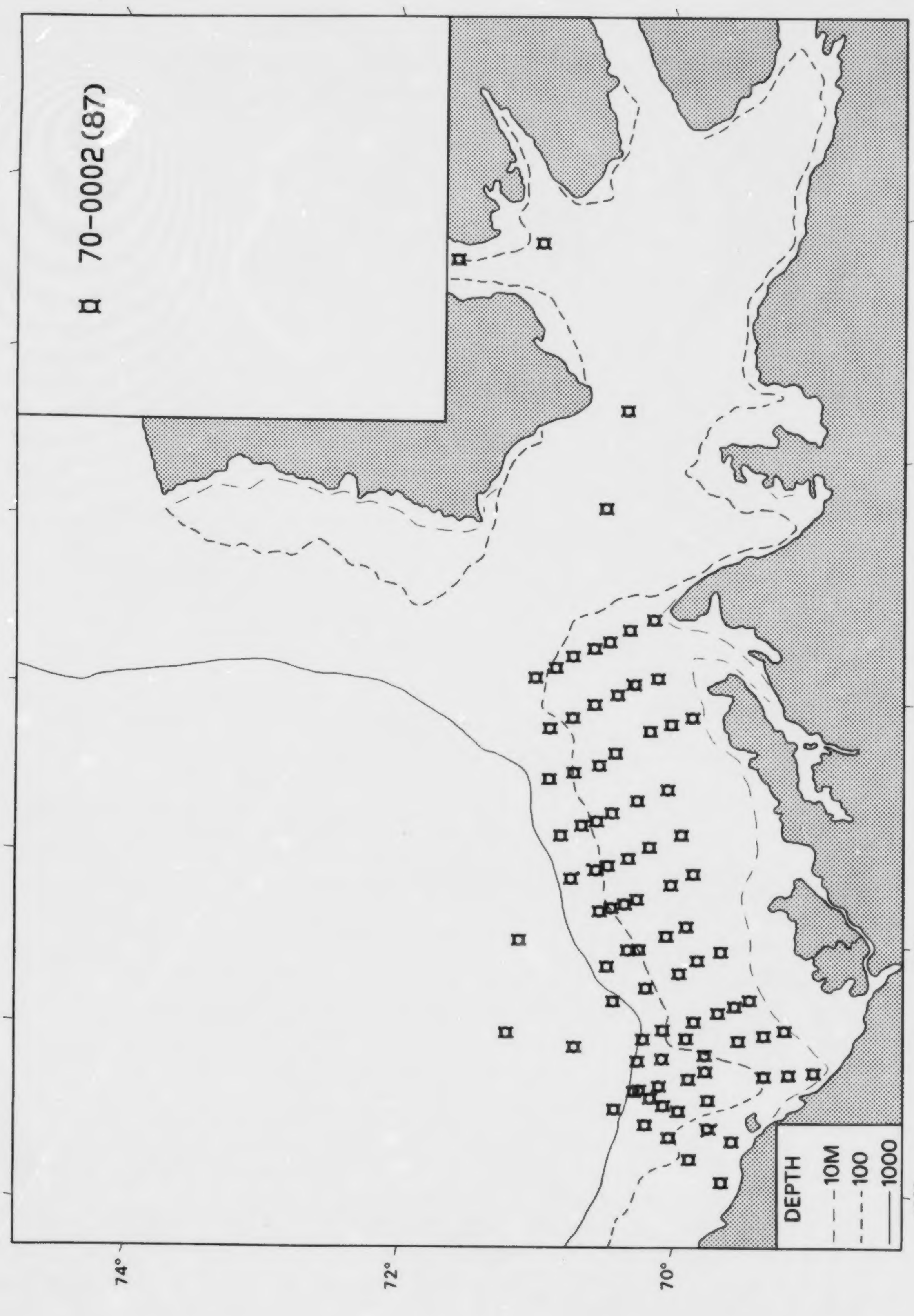
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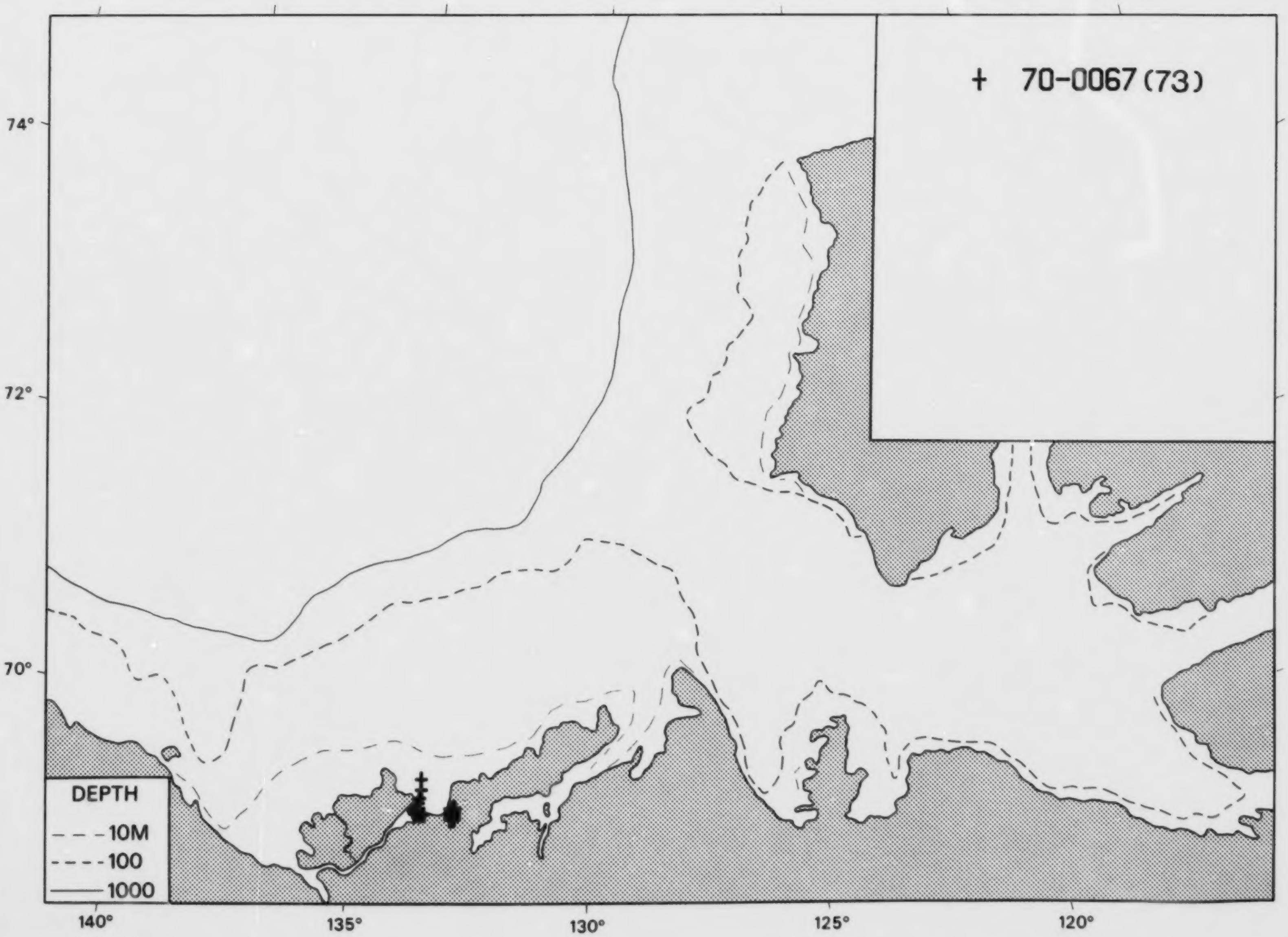
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DEPTH
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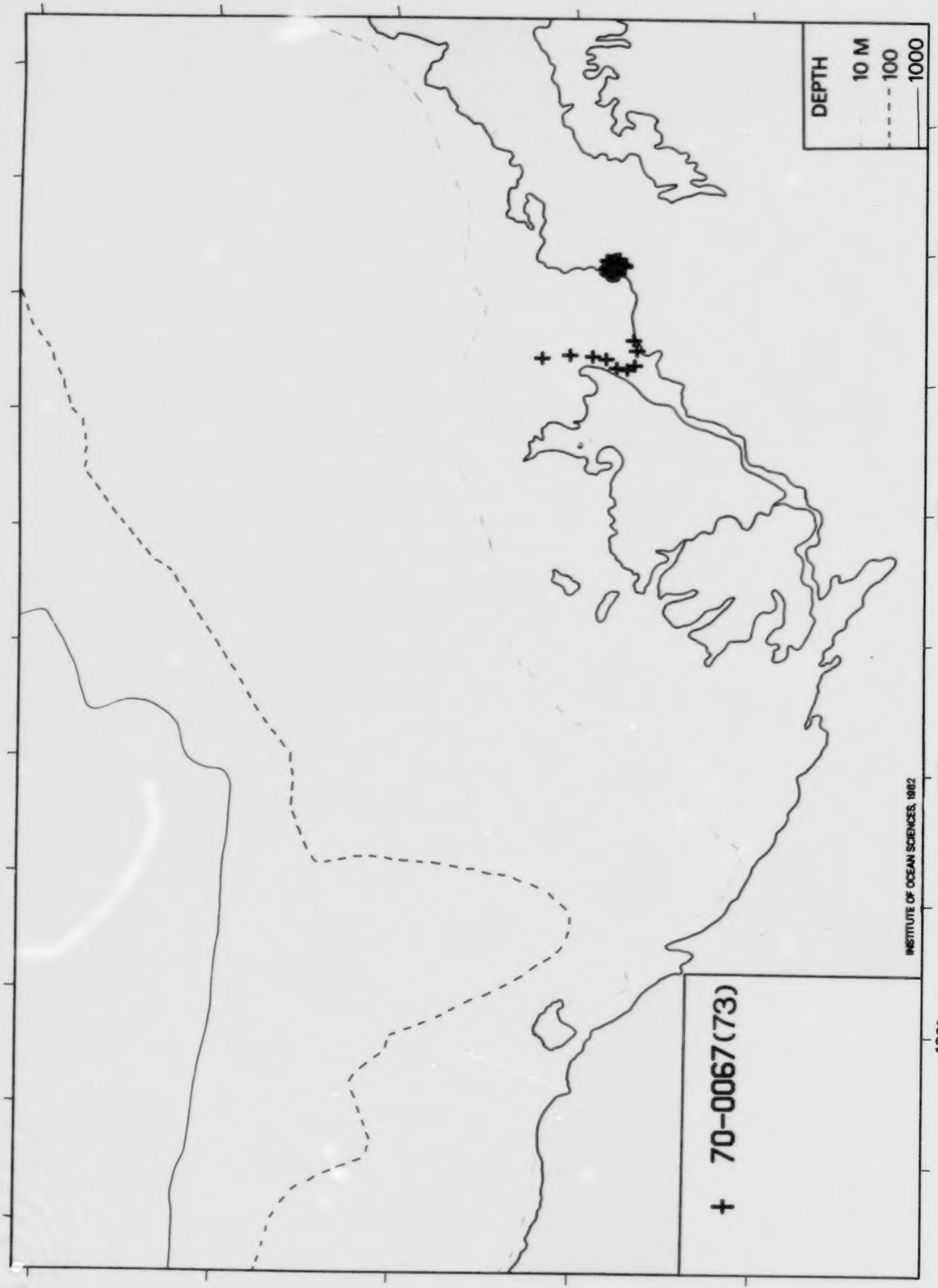
+ 70-0067(73)
DEPTH
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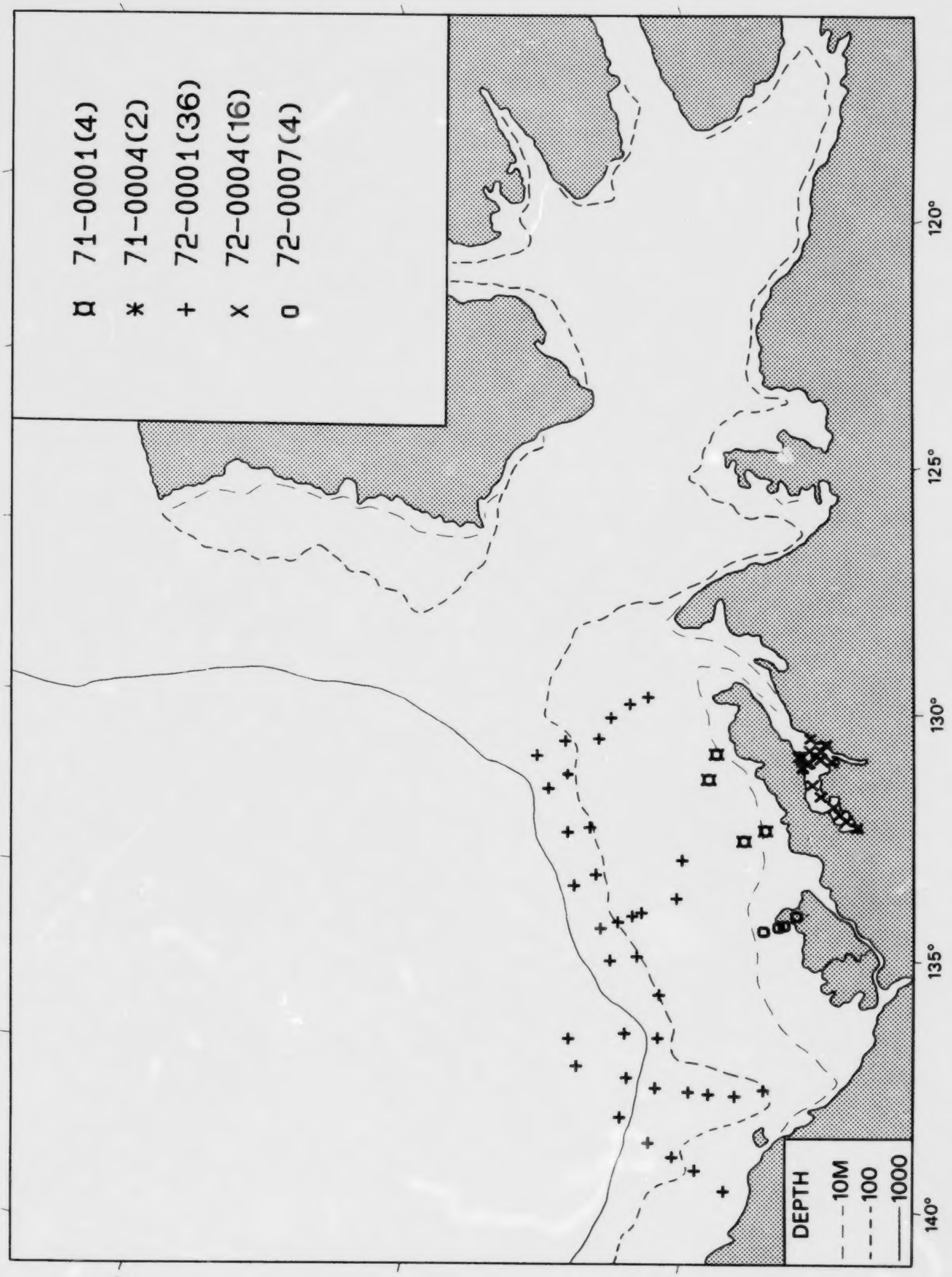
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 71-0001(4)
* 71-0004(2)
+ 72-0001(36)
x 72-0004(16)
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DEPTH
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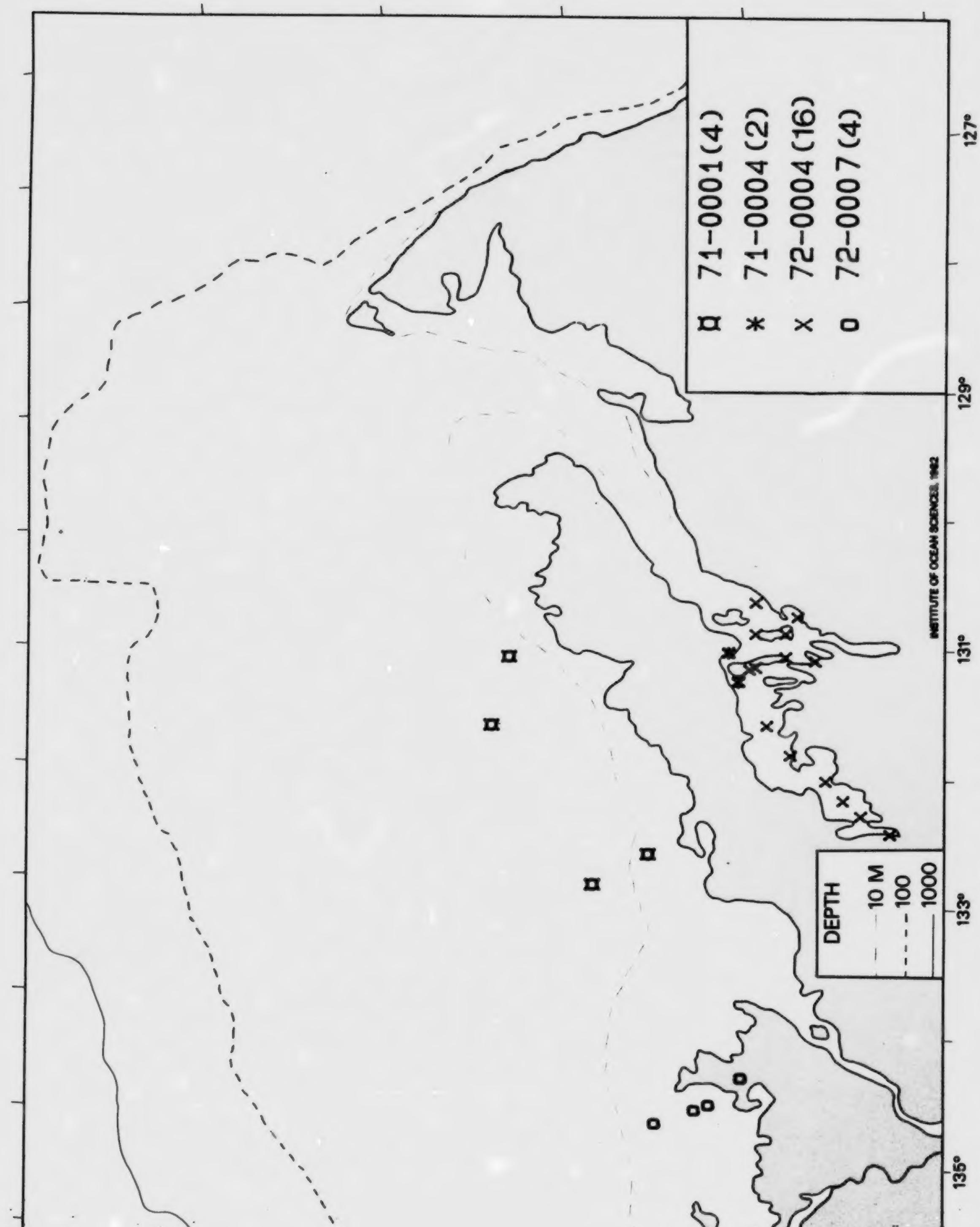
71-0001 (4)
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72-0004 (16)
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DEPTH
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INSTITUTE OF OCEAN SCIENCES, 1982
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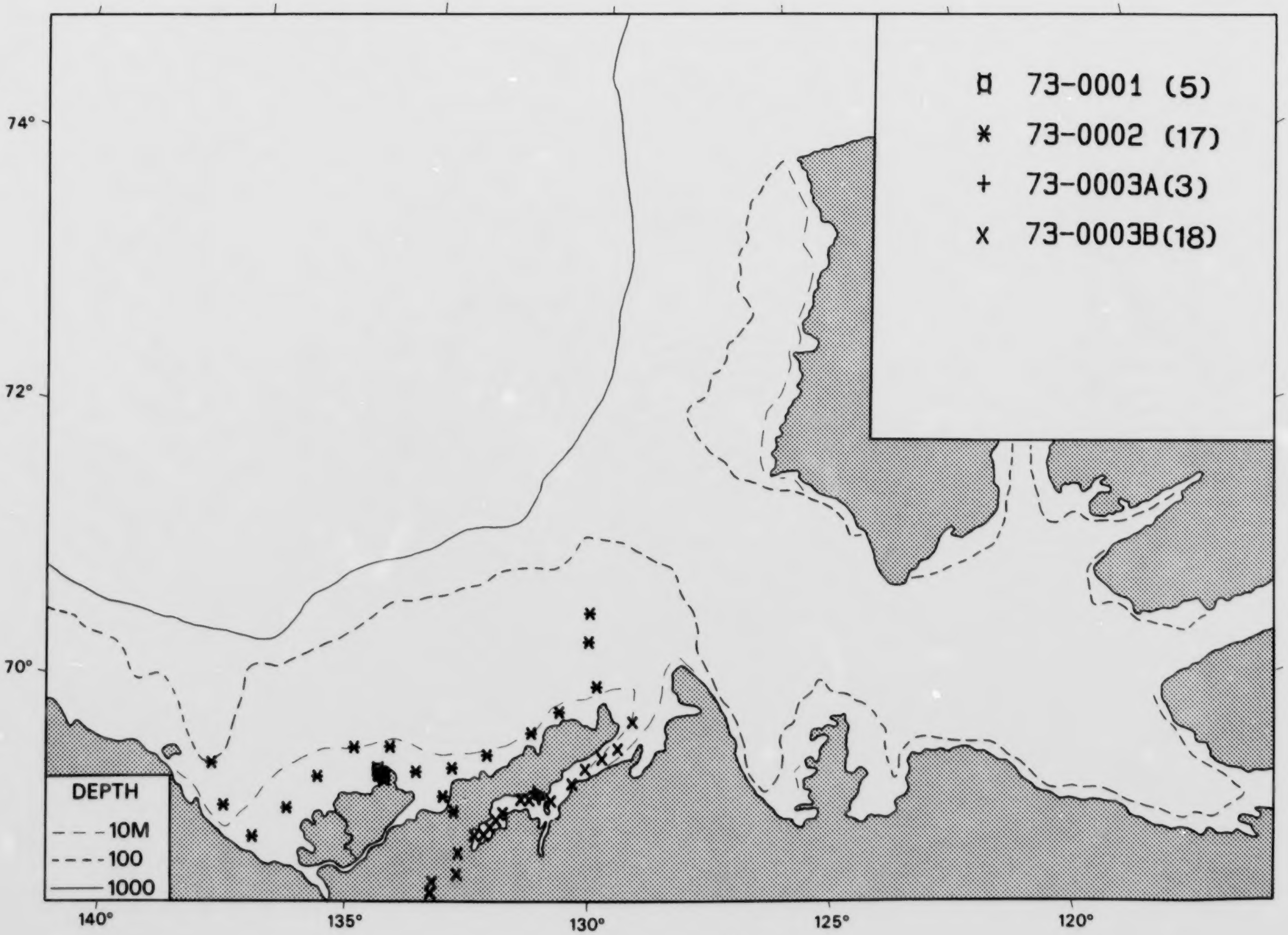
¤ 73-0001 (5)
✱ 73-0002 (17)
+ 73-0003A(3)
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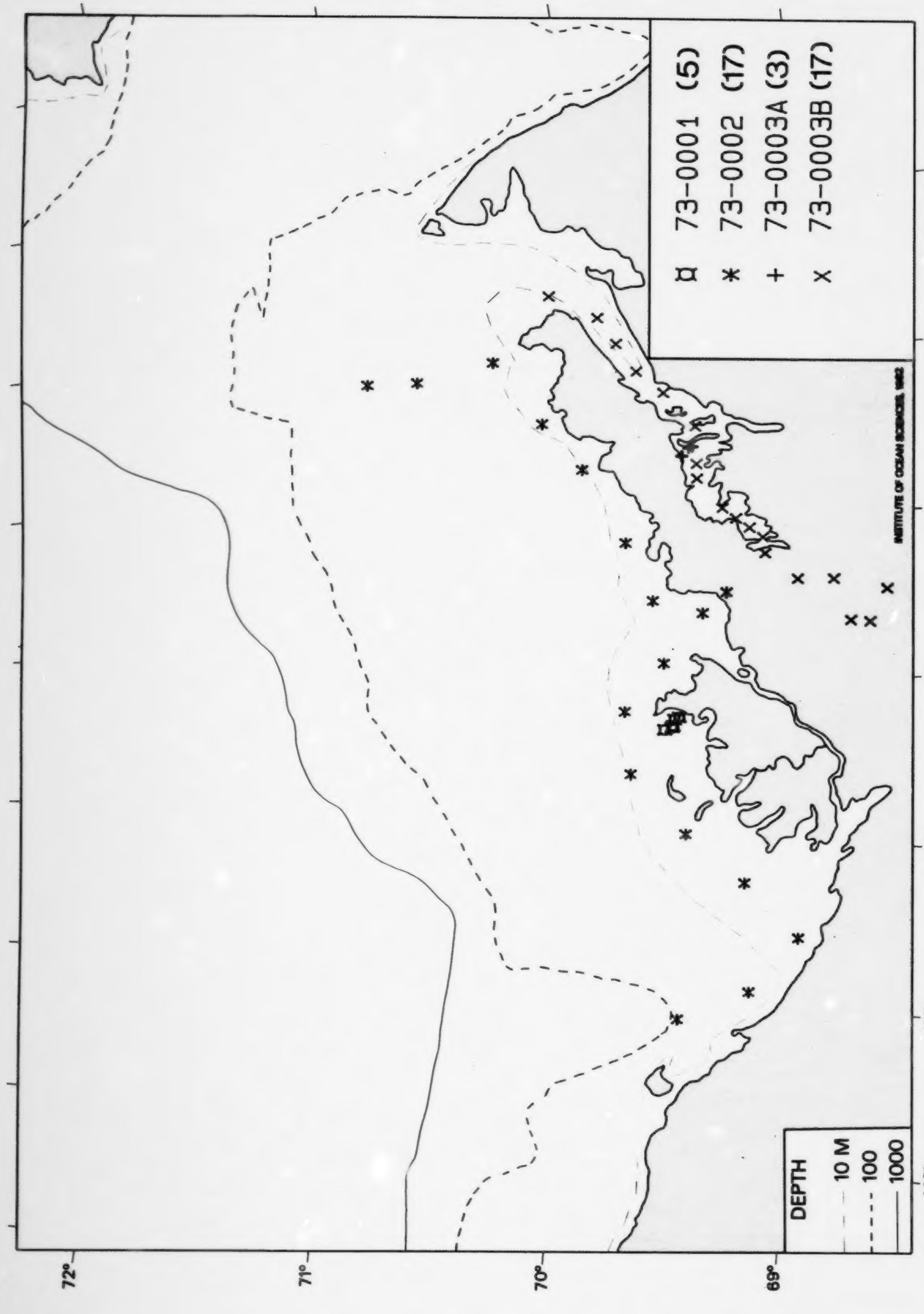
73-0001 (5)
73-0002 (17)
73-0003A (3)
73-0003B (17)
DEPTH
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INSTITUTE OF OCEAN SCIENCES, 1982
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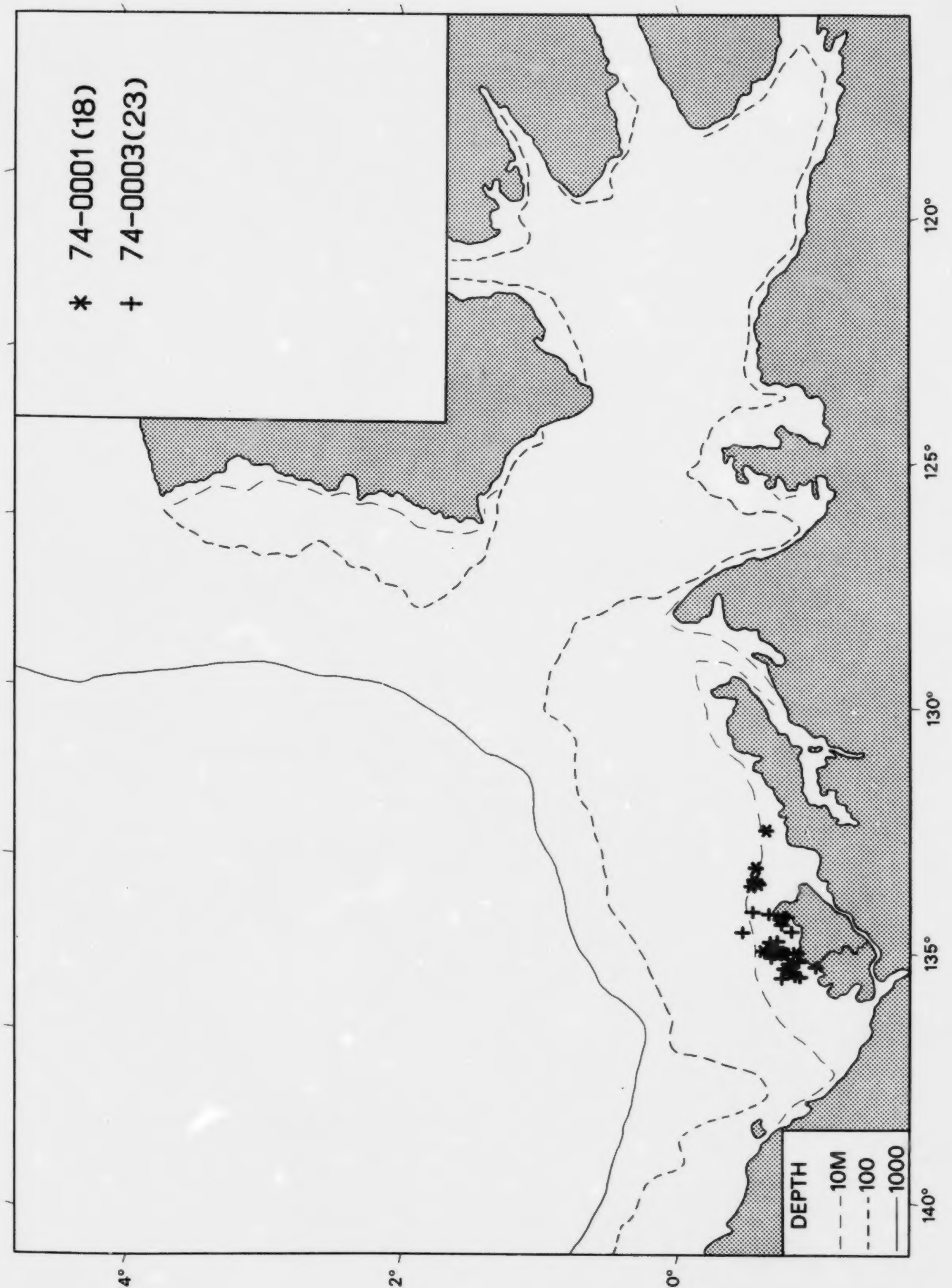

* 74-0001 (18)
+ 74-0003 (23)
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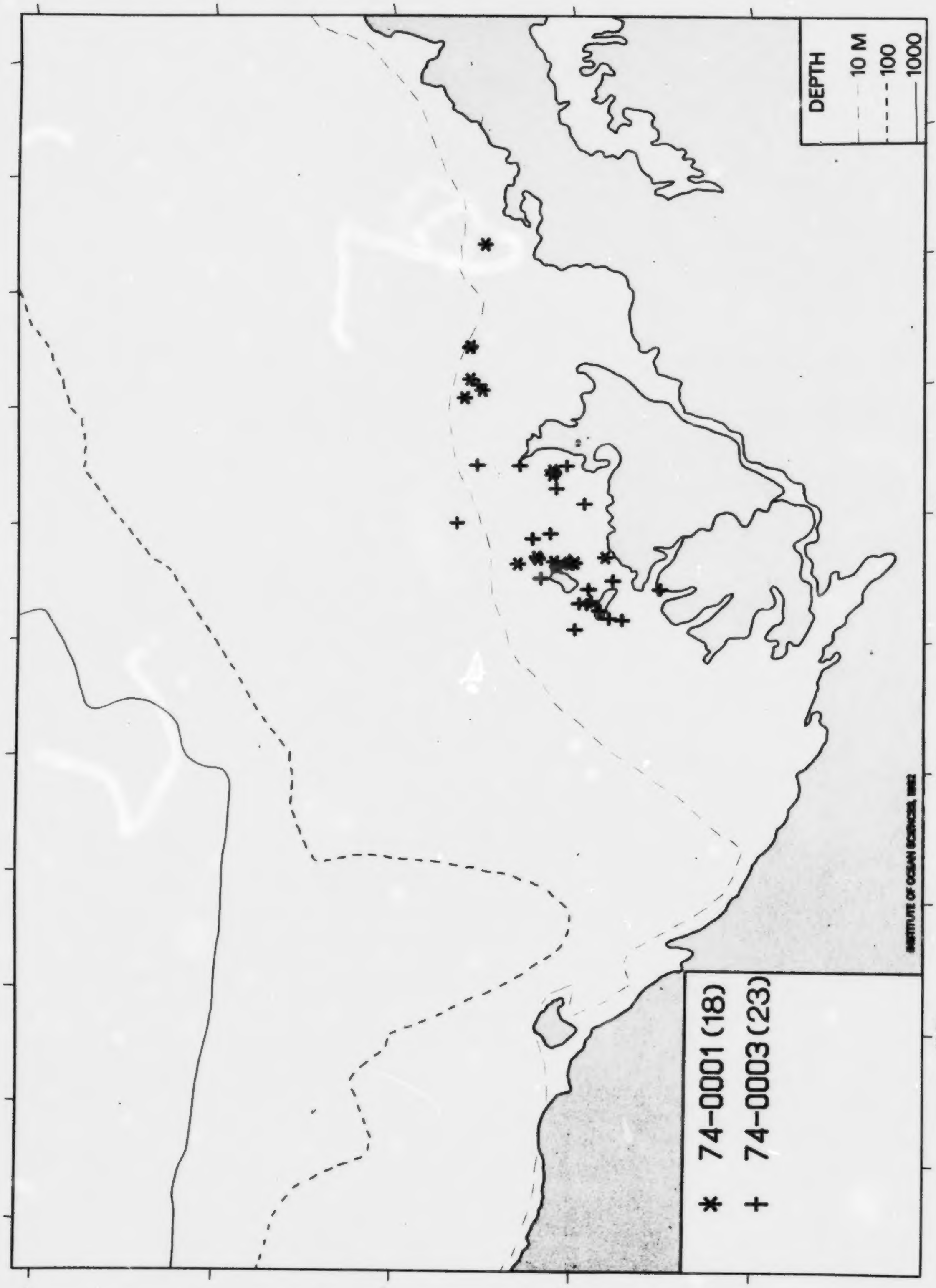
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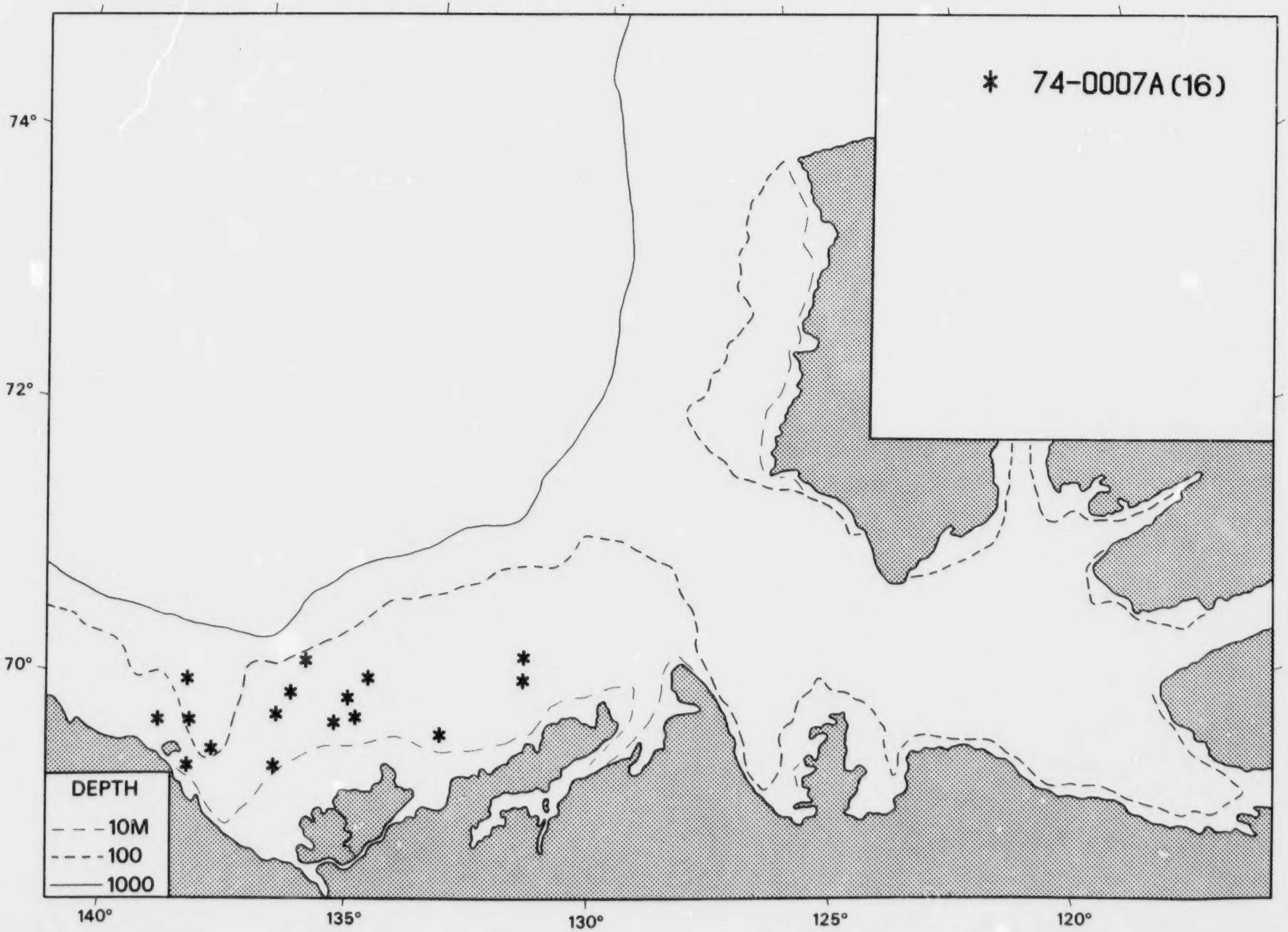
* 74-0001 (18)
+ 74-0003 (23)
INSTITUTE OF OCEAN SCIENCES,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DEPTH
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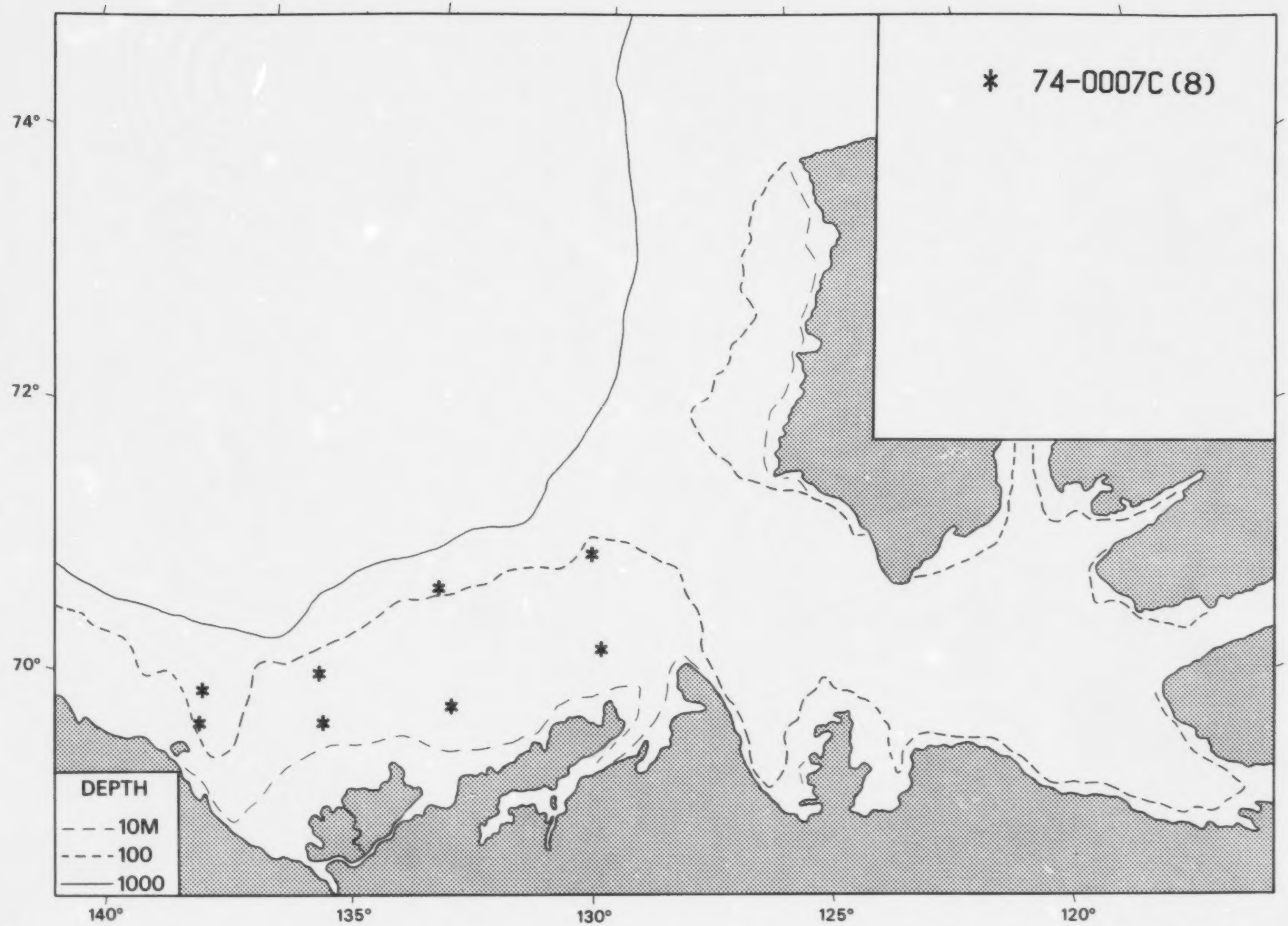
* 74-0007C (8)
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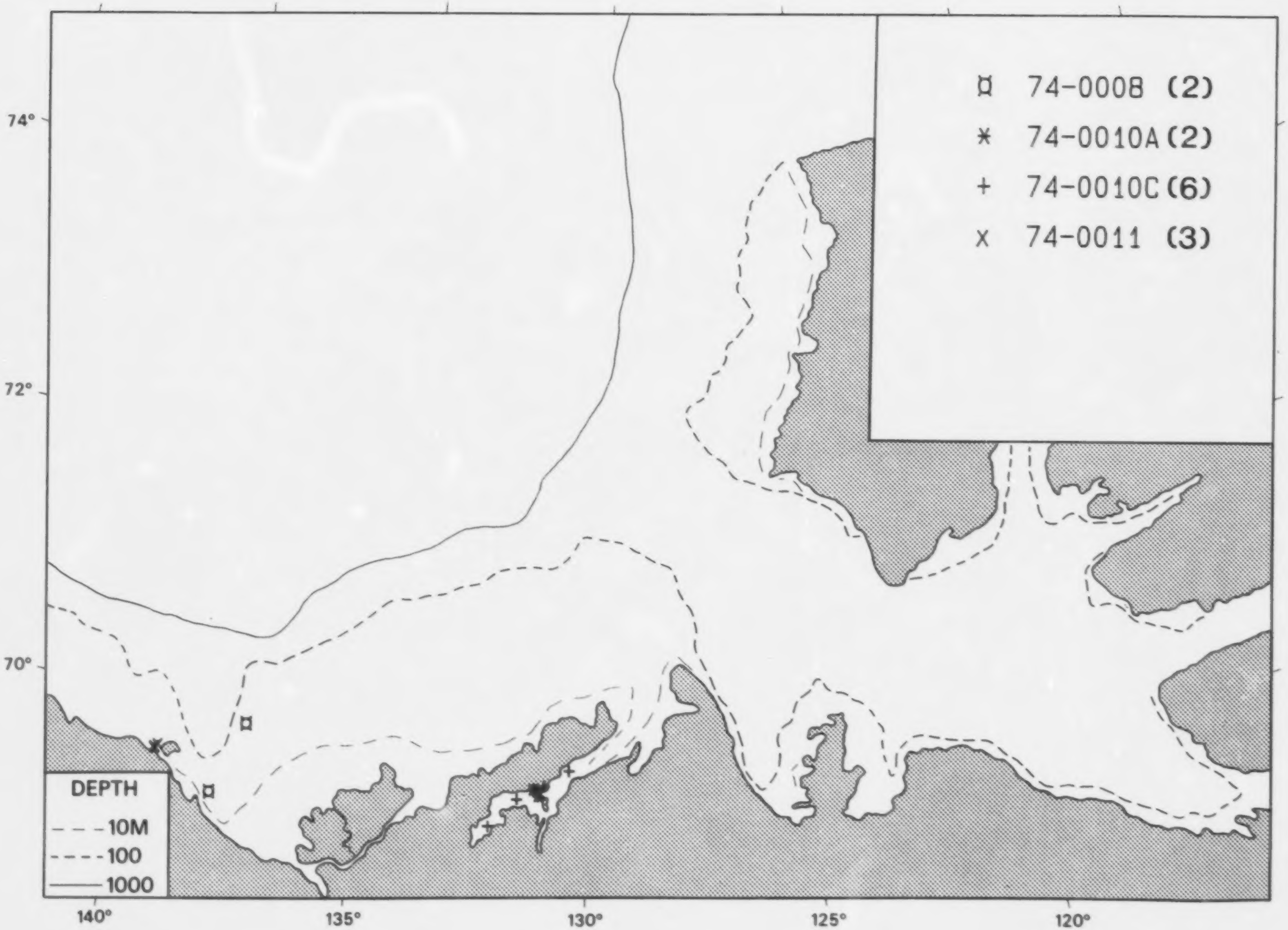
¤ 74-0008 (2)
✱ 74-0010A (2)
+ 74-0010C (6)
x 74-0011 (3)
DEPTH
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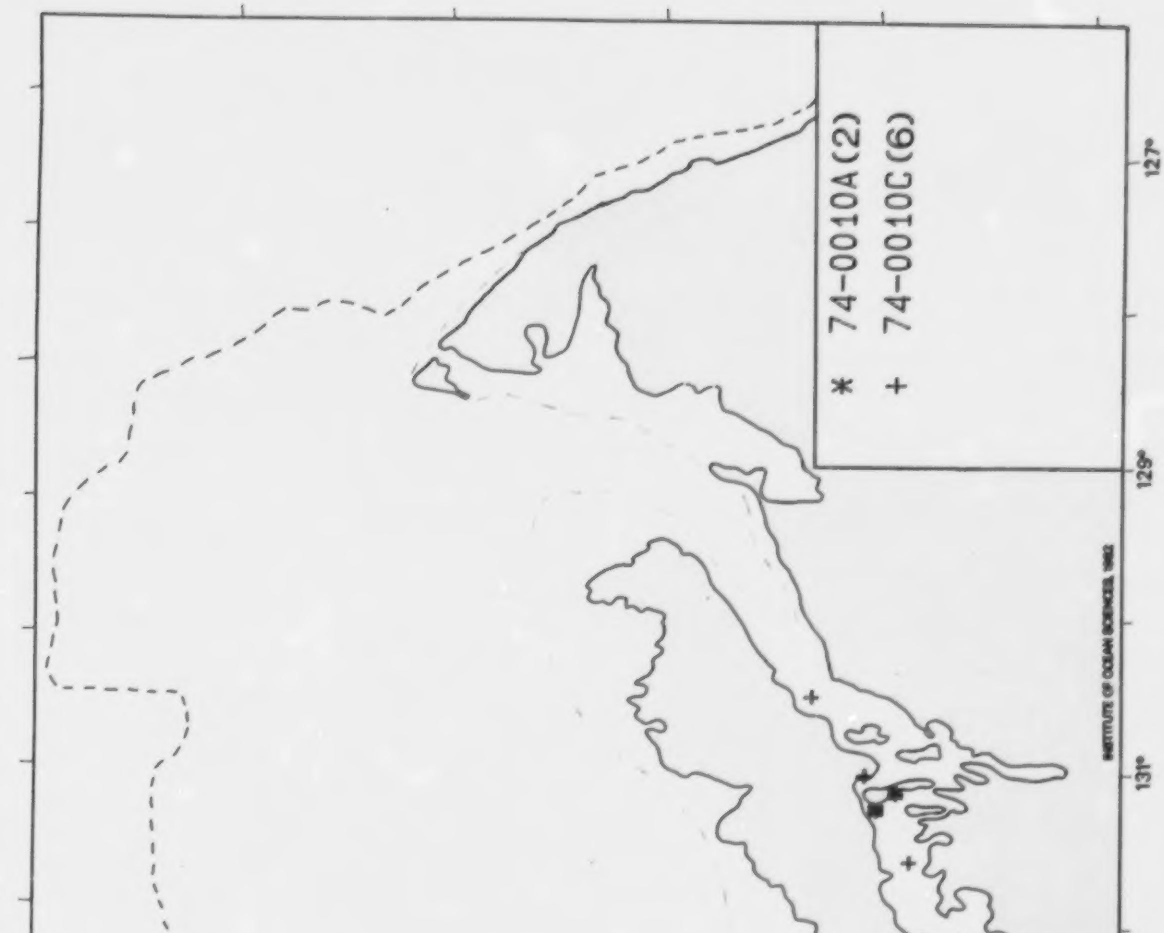
* 74-0010A(2)
+ 74-0010C(6)
DEPTH
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INSTITUTE OF OCEAN SCIENCES 1982
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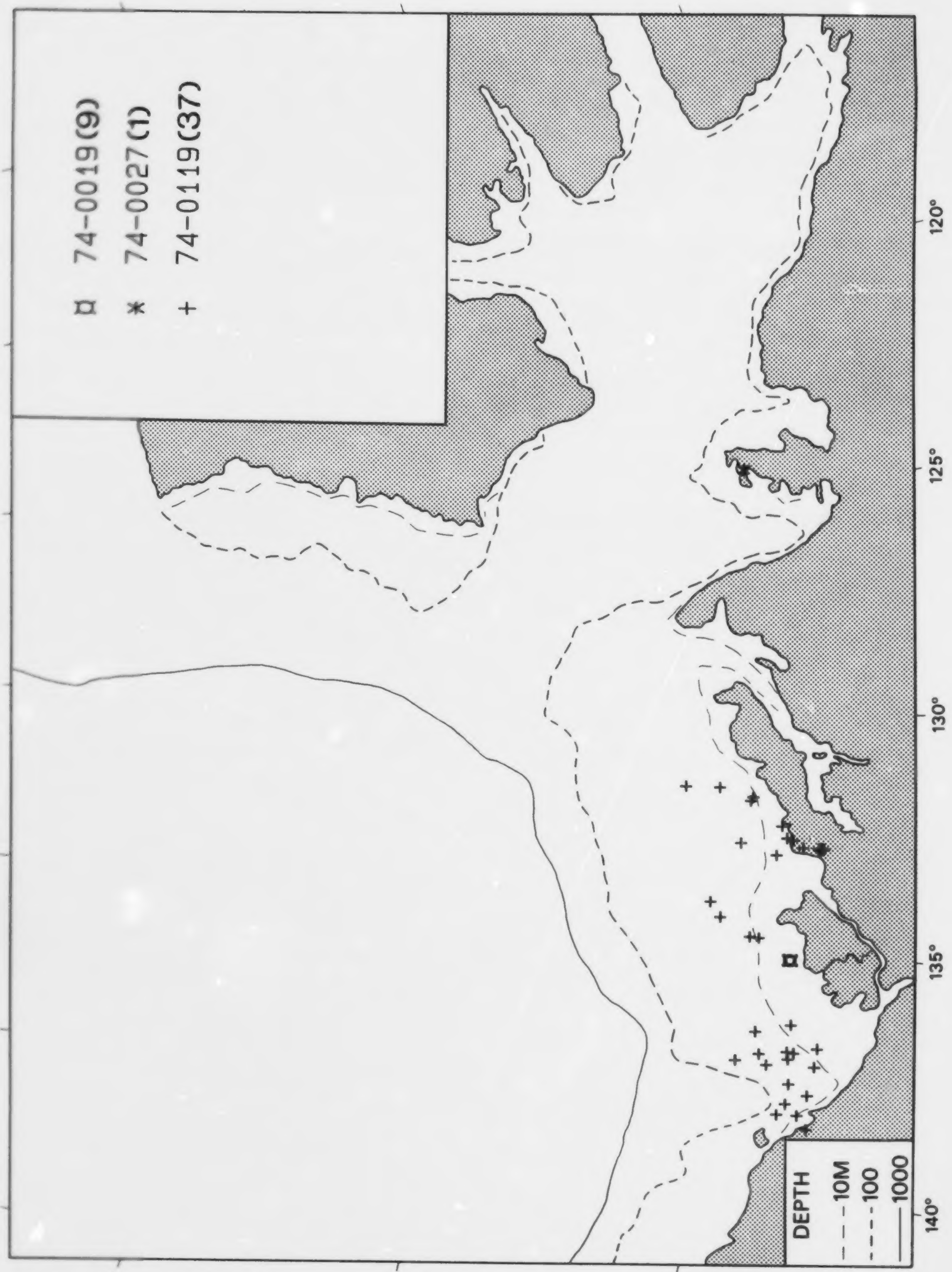
¤ 74-0019(9)
✱ 74-0027(1)
+ 74-0119(37)
DEPTH
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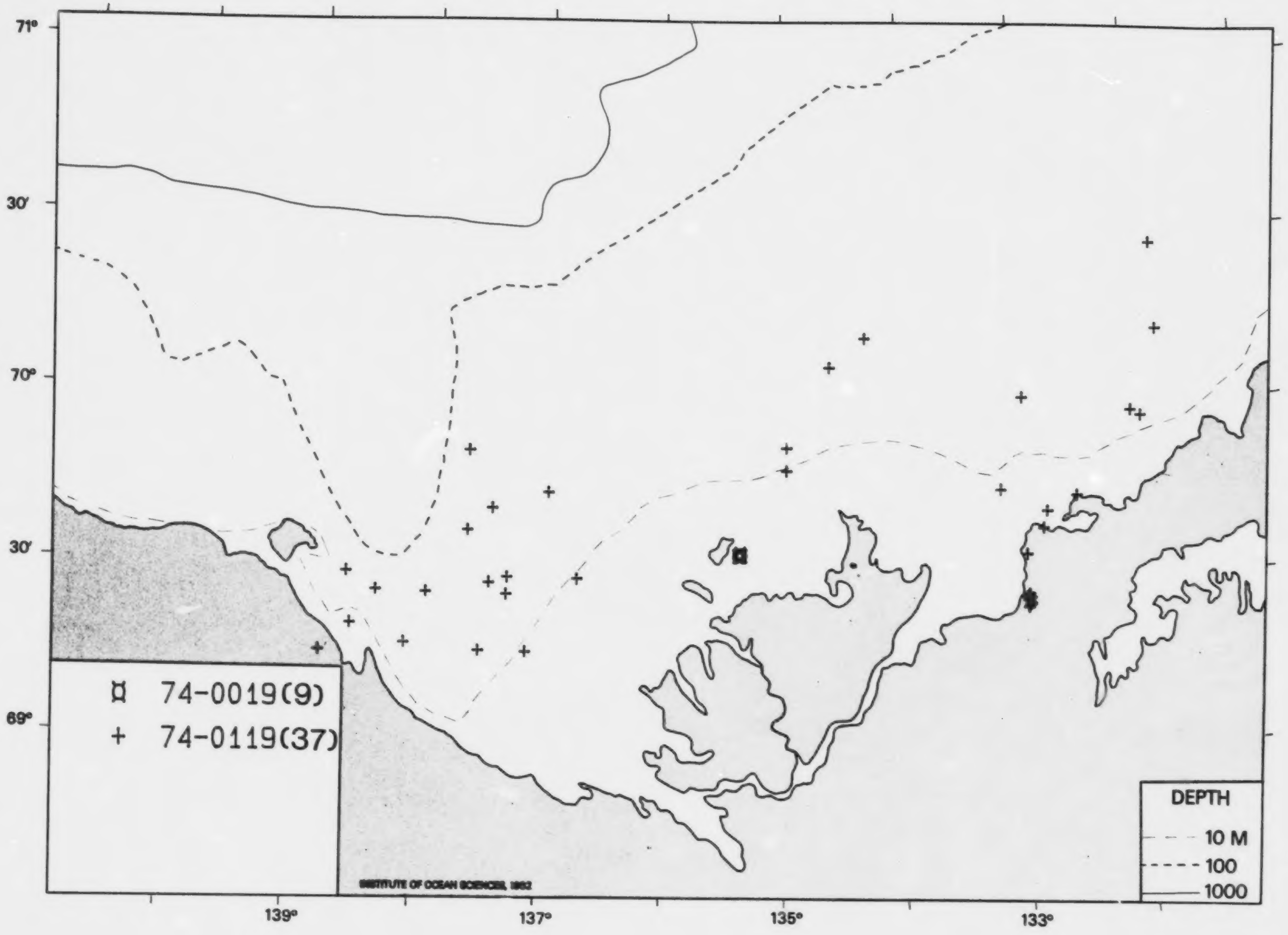
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¤ 74-0019(9)
+ 74-0119(37)
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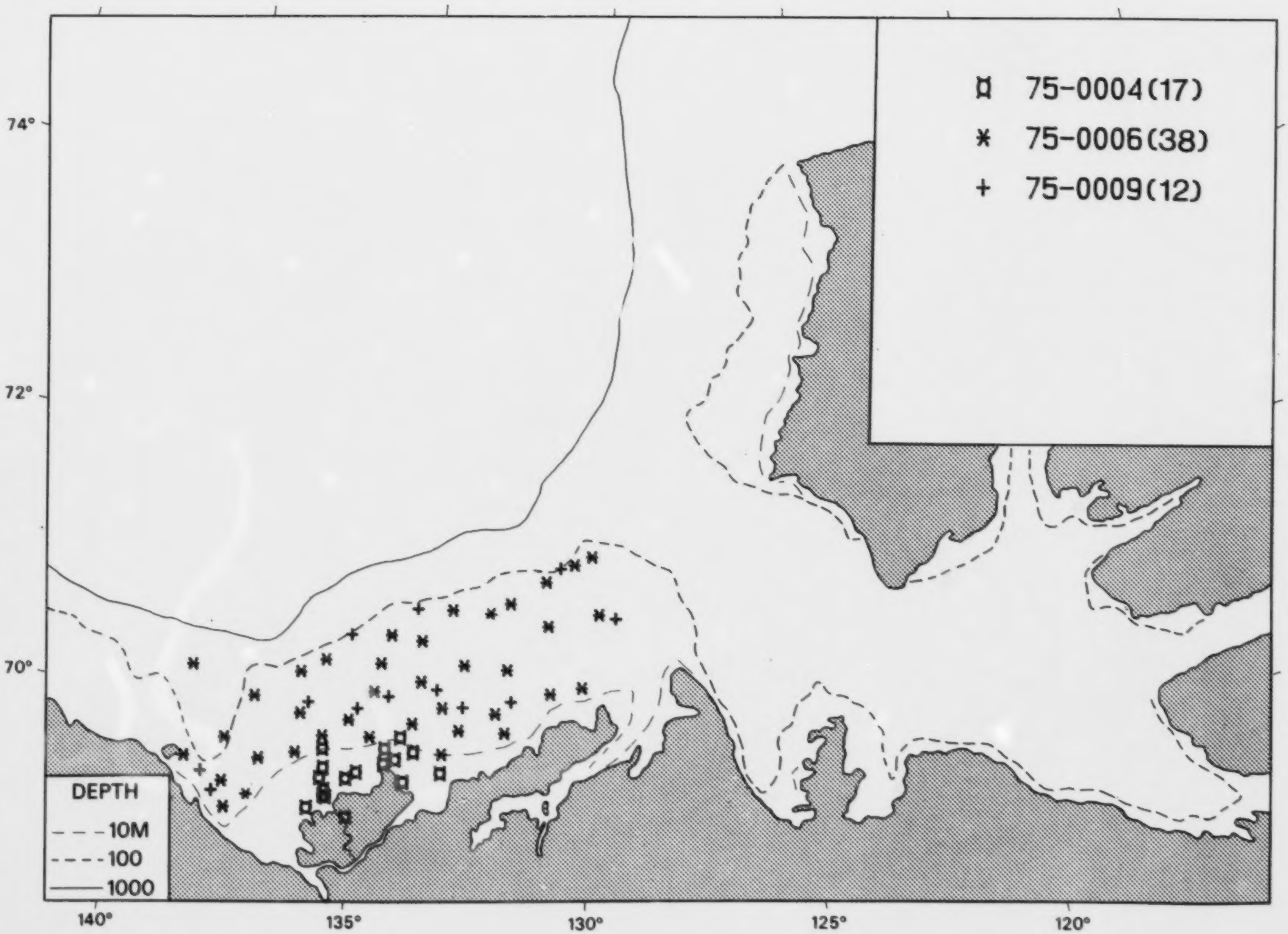
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 75-0004(17)
✱ 75-0006(38)
+ 75-0009(12)
DEPTH
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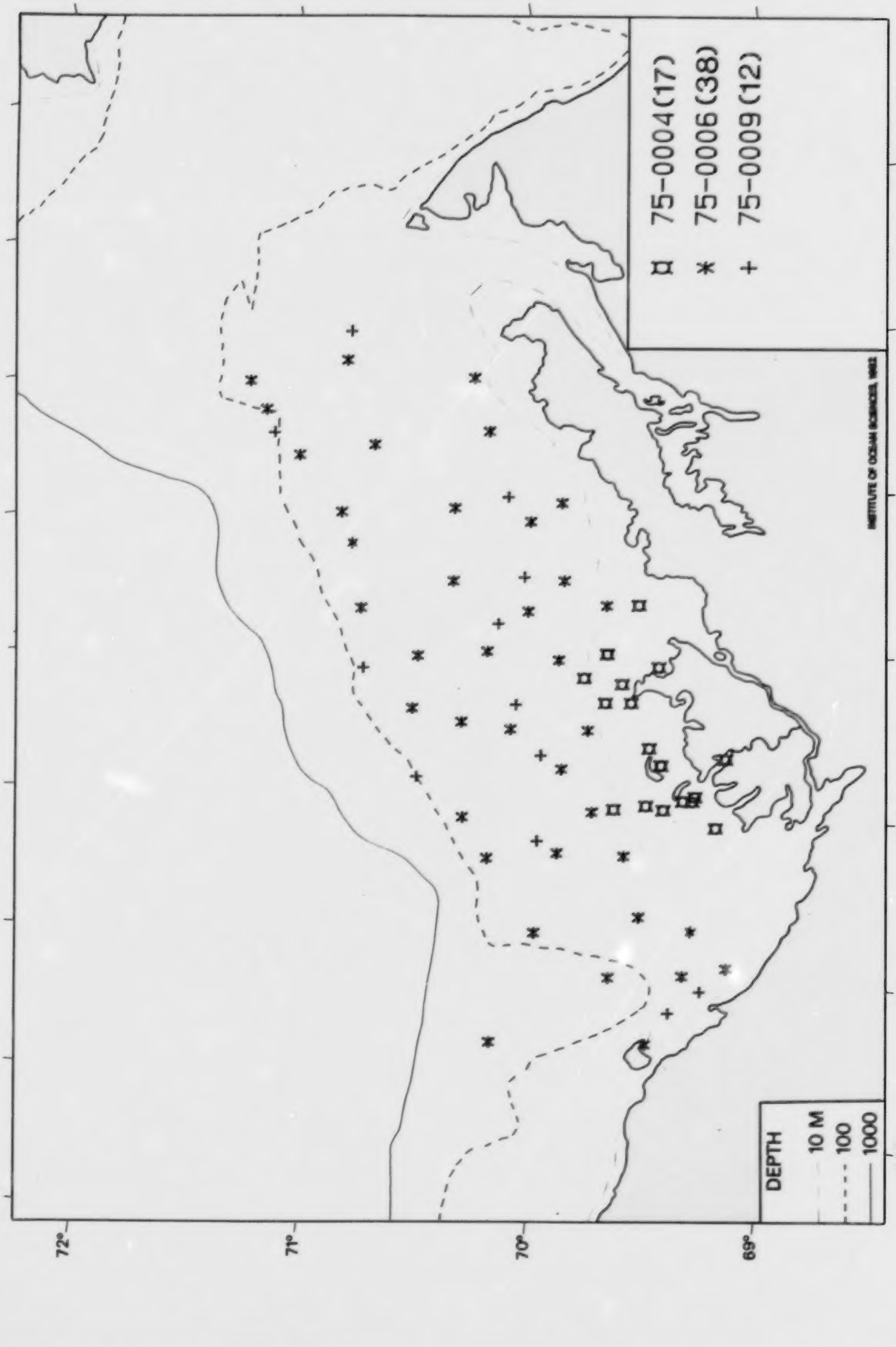
□ 75-0004(17)
* 75-0006(38)
+ 75-0009(12)
DEPTH
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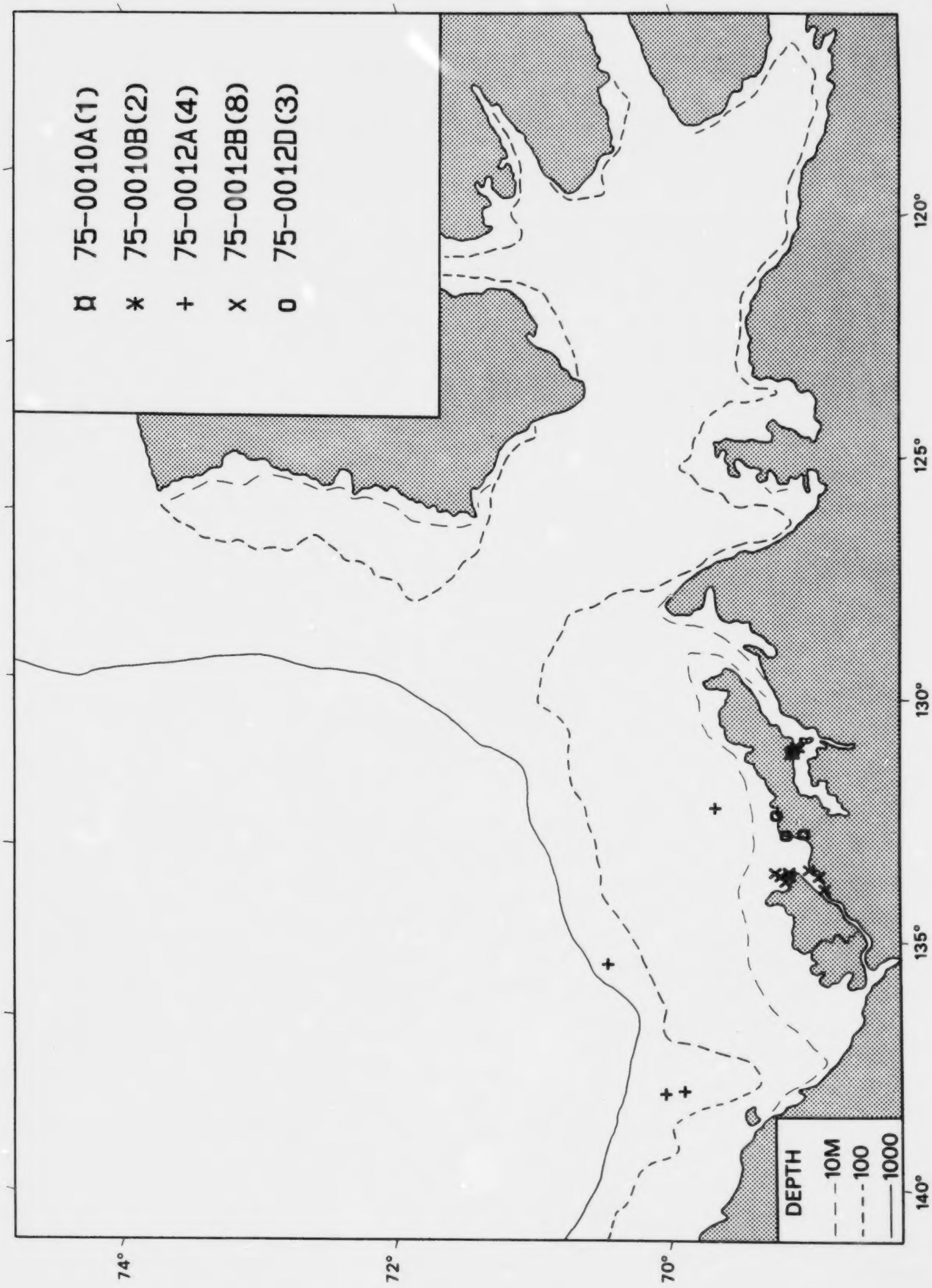
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 75-0010A(1)
* 75-0010B(2)
+ 75-0012A(4)
x 75-0012B(8)
o 75-0012D(3)
DEPTH
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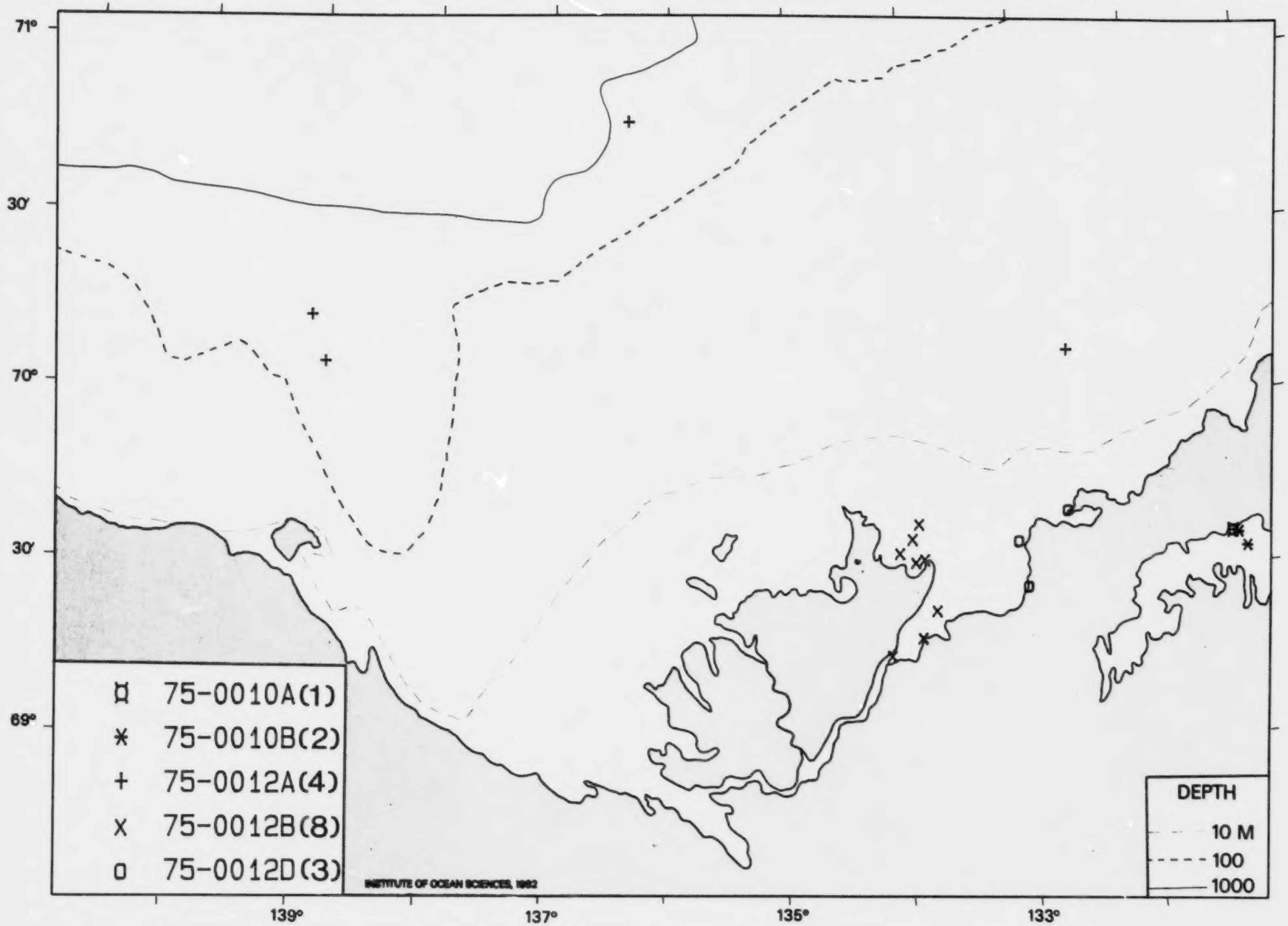
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75-0010A(1)
75-0010B(2)
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75-0012D(3)
DEPTH
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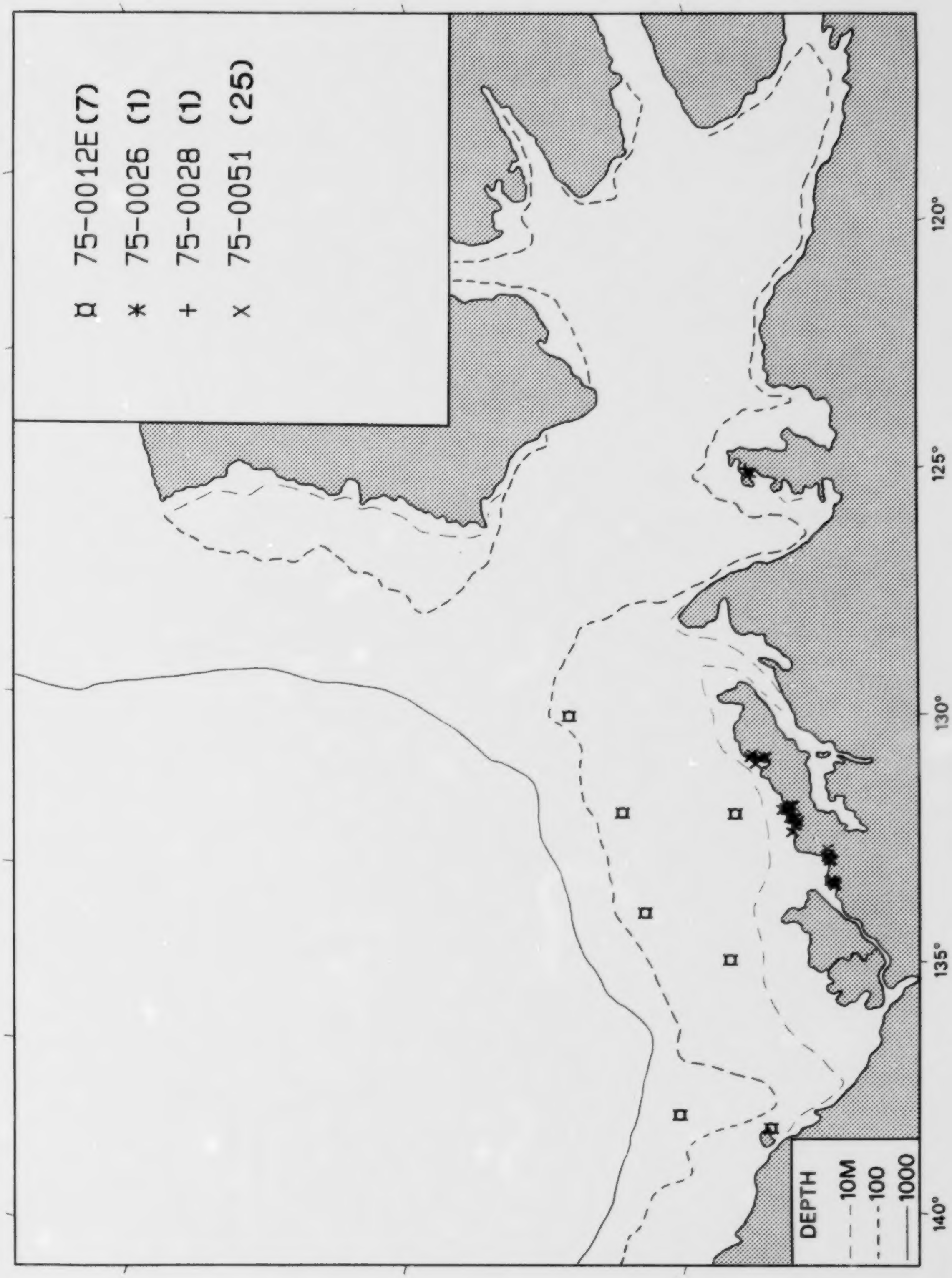
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 75-0012E (7)
* 75-0026 (1)
+ 75-0028 (1)
× 75-0051 (25)
DEPTH
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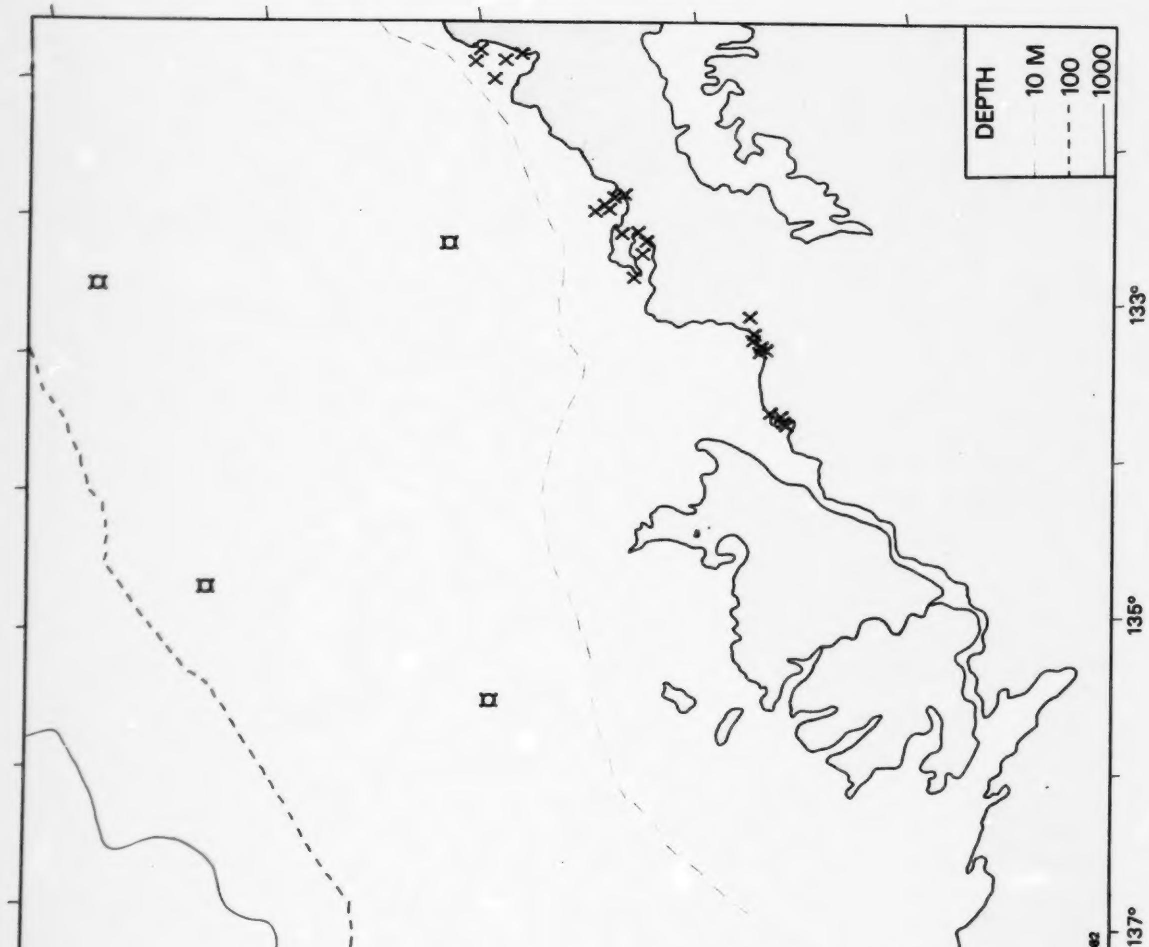
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DEPTH
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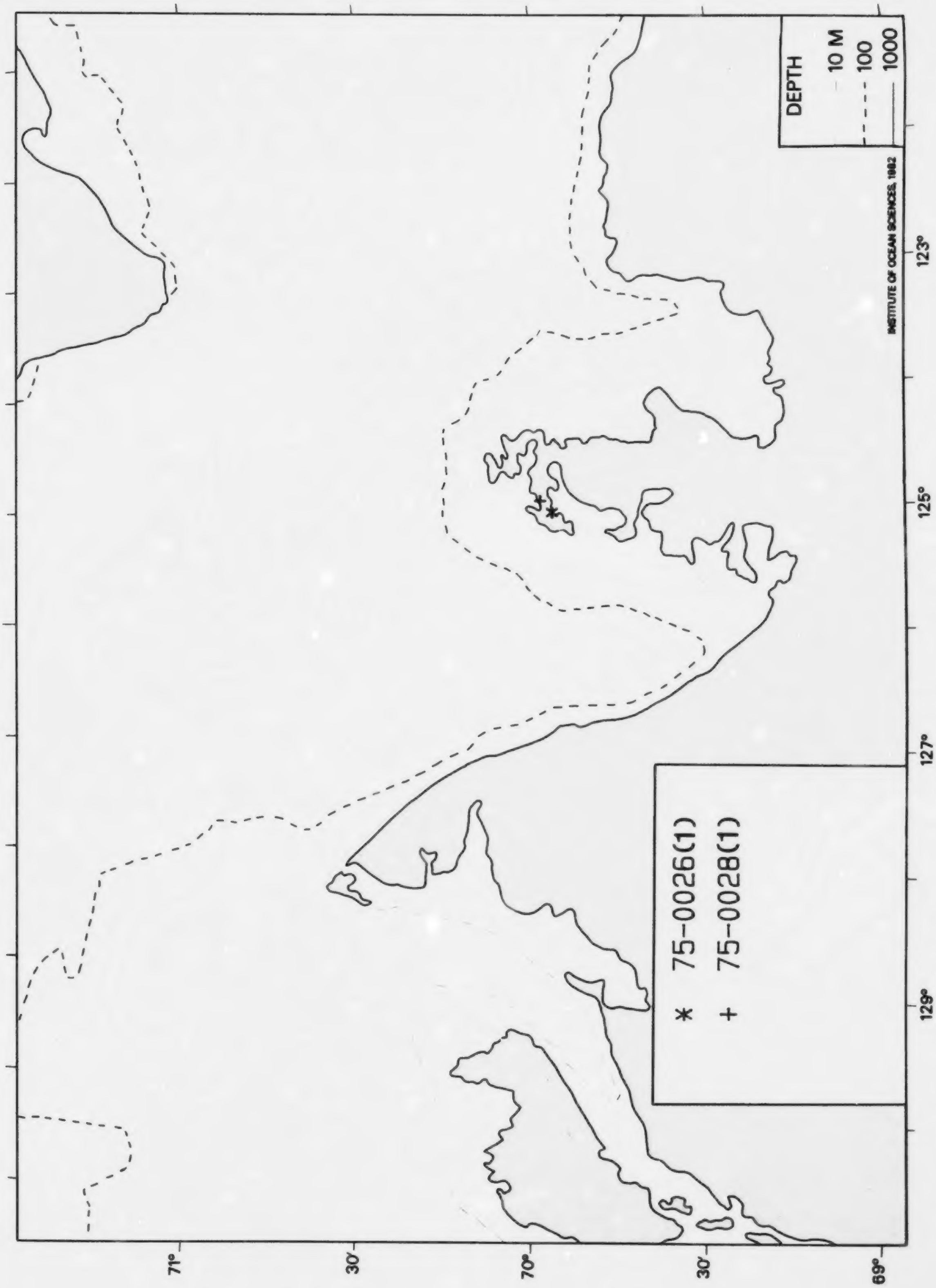
75-0051 (25
INSTITUTE OF OCEAN SCIENCES, 1982


* 75-0026(1)
+ 75-0028(1)
DEPTH
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INSTITUTE OF OCEAN SCIENCES, 1982
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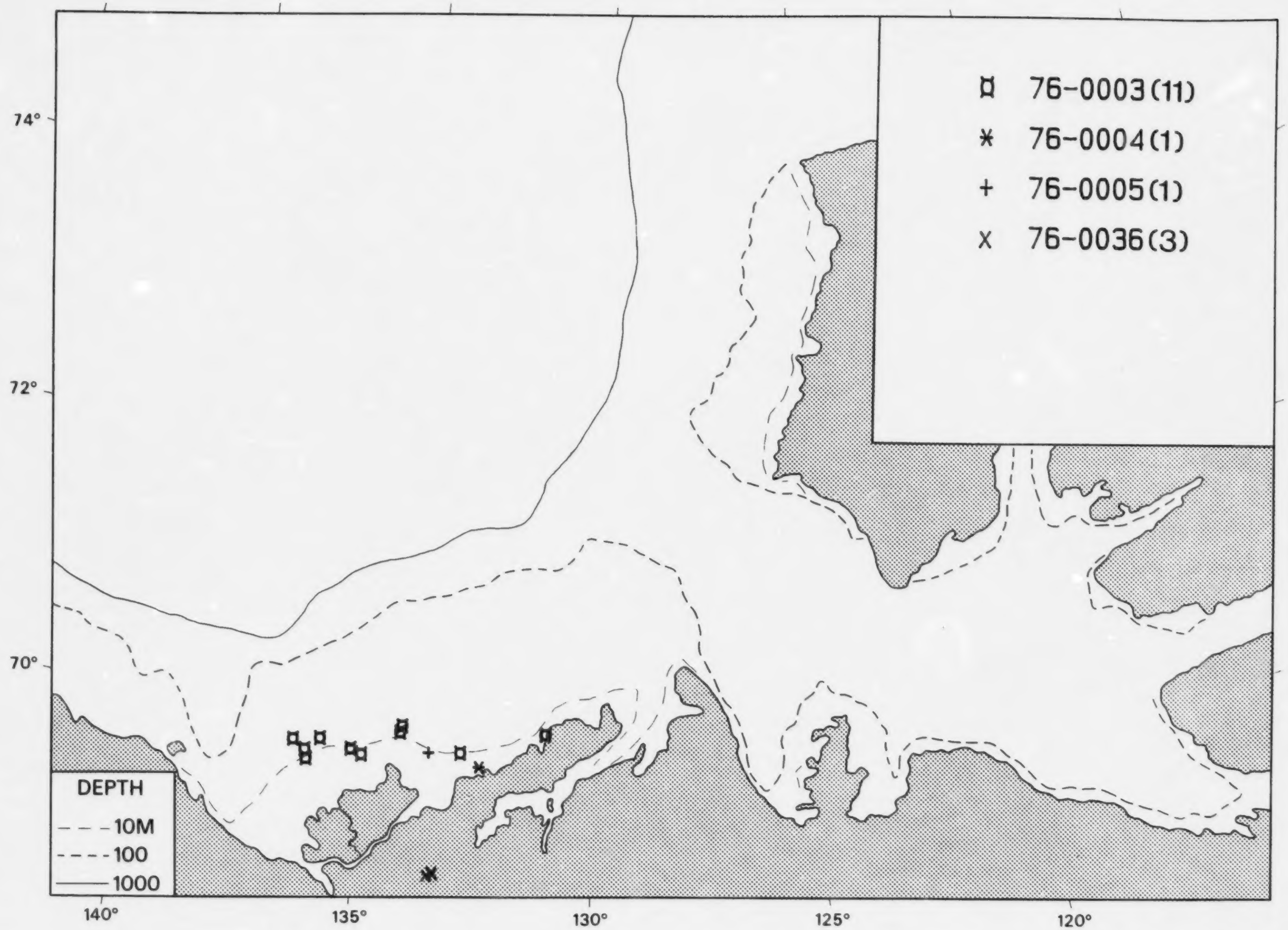
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¤ 76-0003(11)
✱ 76-0004(1)
+ 76-0005(1)
x 76-0036(3)
DEPTH
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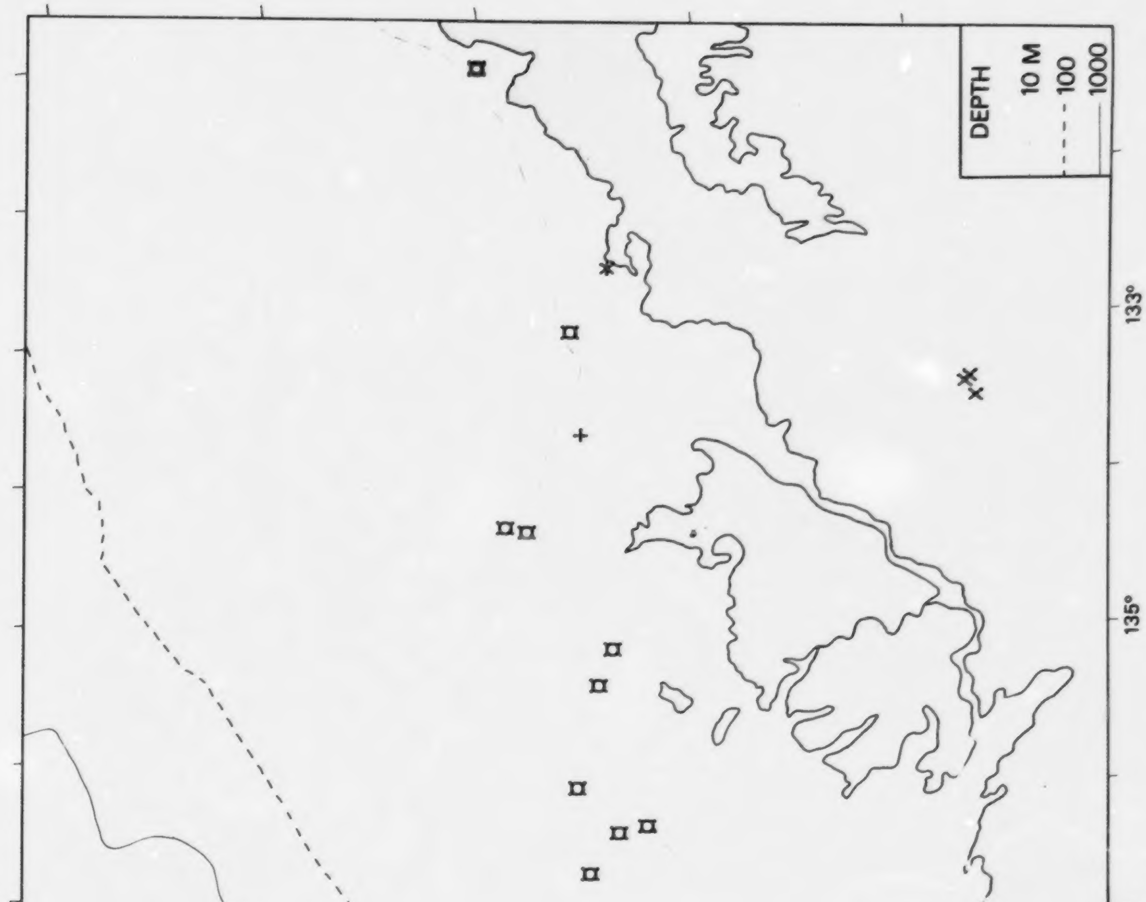
1000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DEPTH
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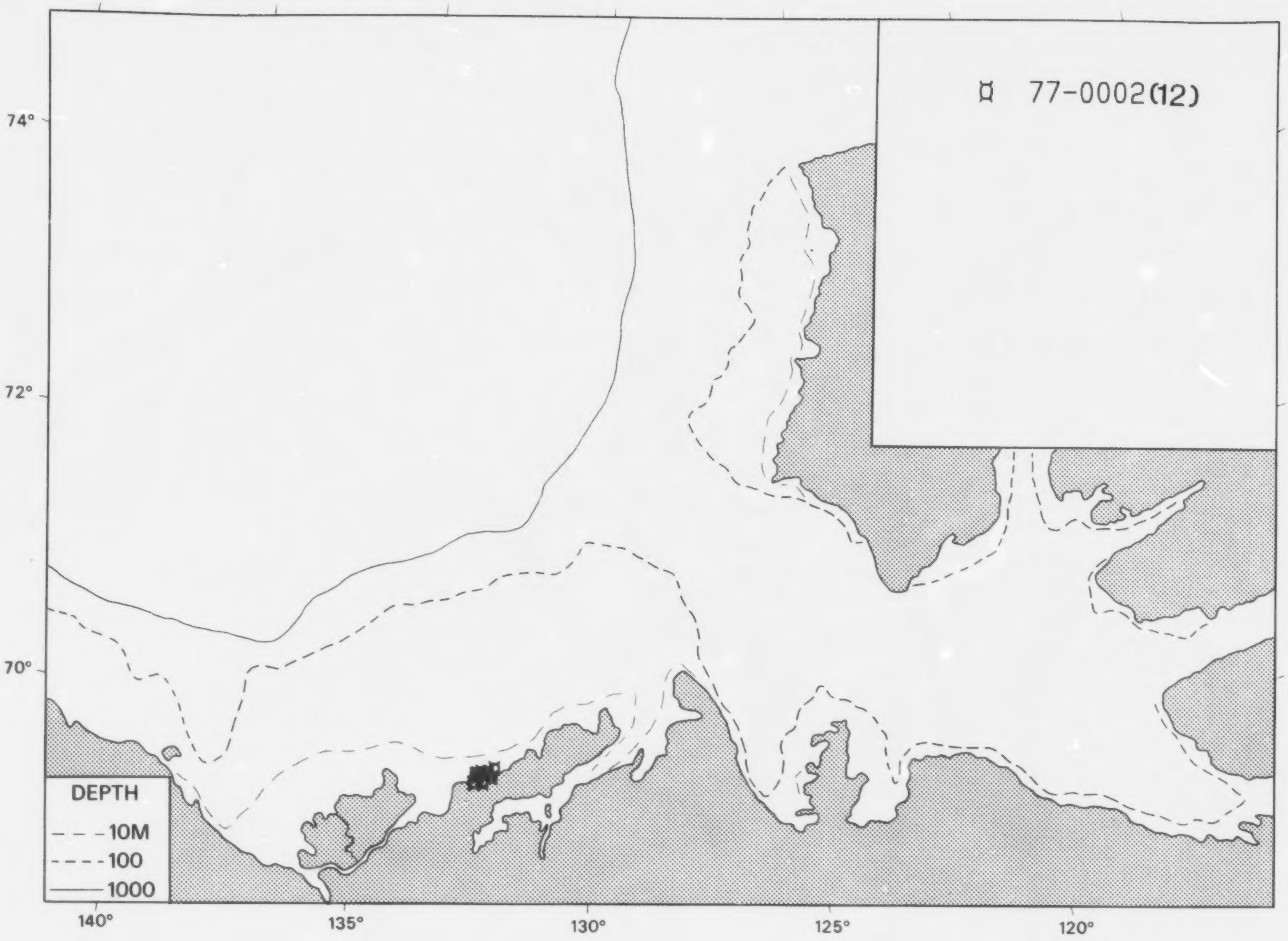
¤ 76-0003(11)
* 76-0004(1)
+ 76-0005(1)
x 76-0036(3)
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 77-0002(12)
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DEPTH
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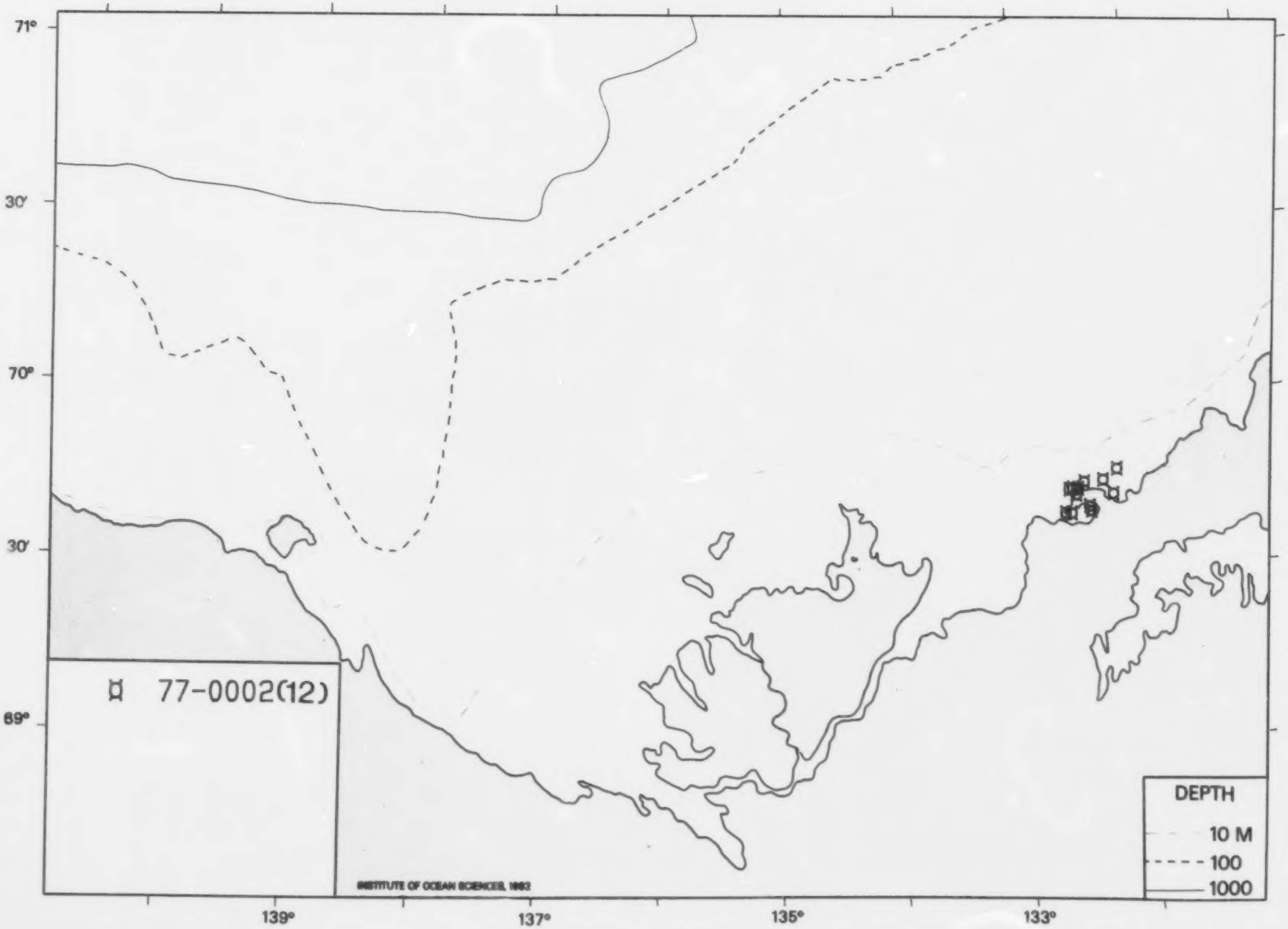
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¤ 77-0002(12)
DEPTH
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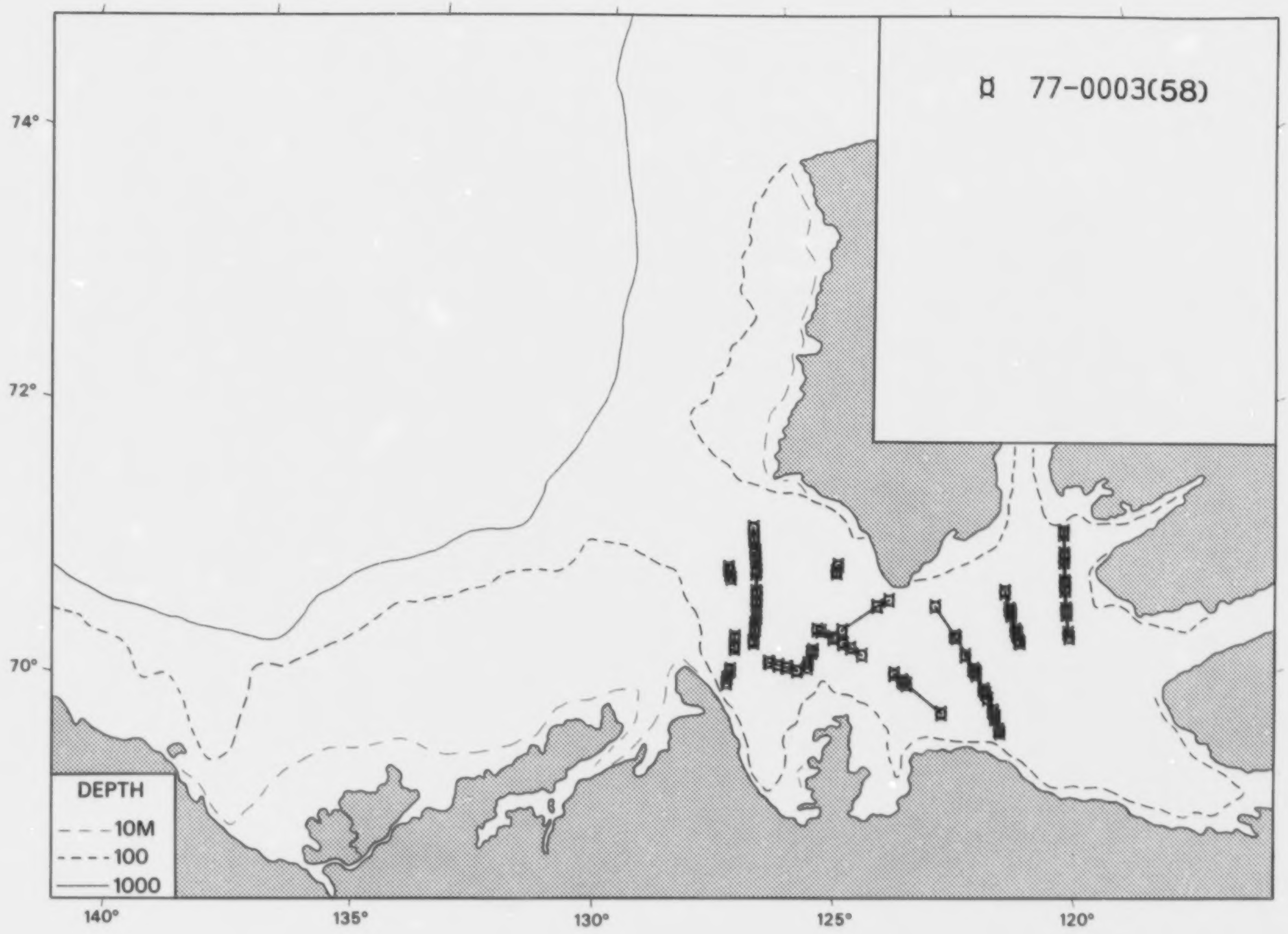
INSTITUTE OF OCEAN SCIENCES, 1982


77-0003(58)
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DEPTH
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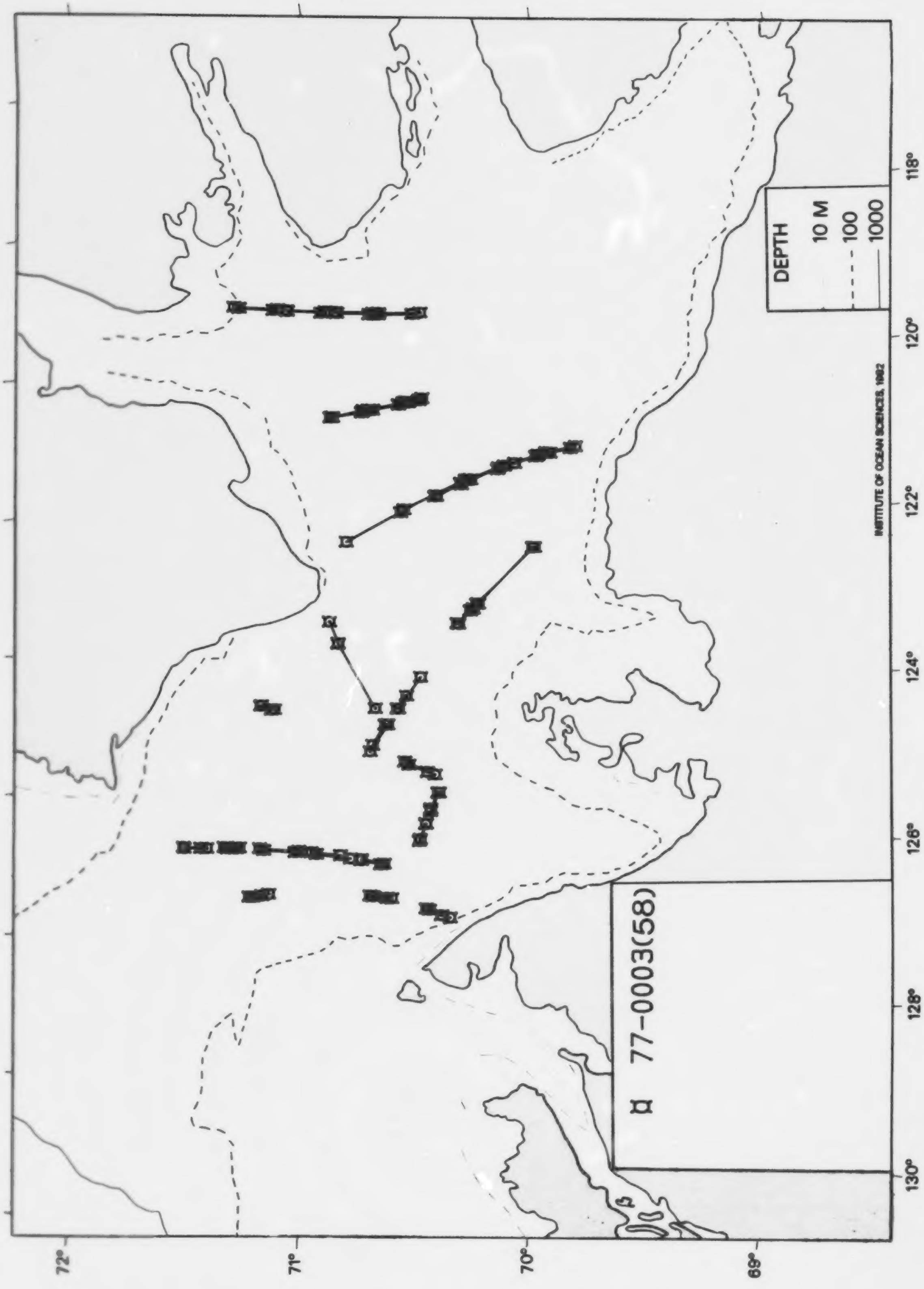
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DEPTH
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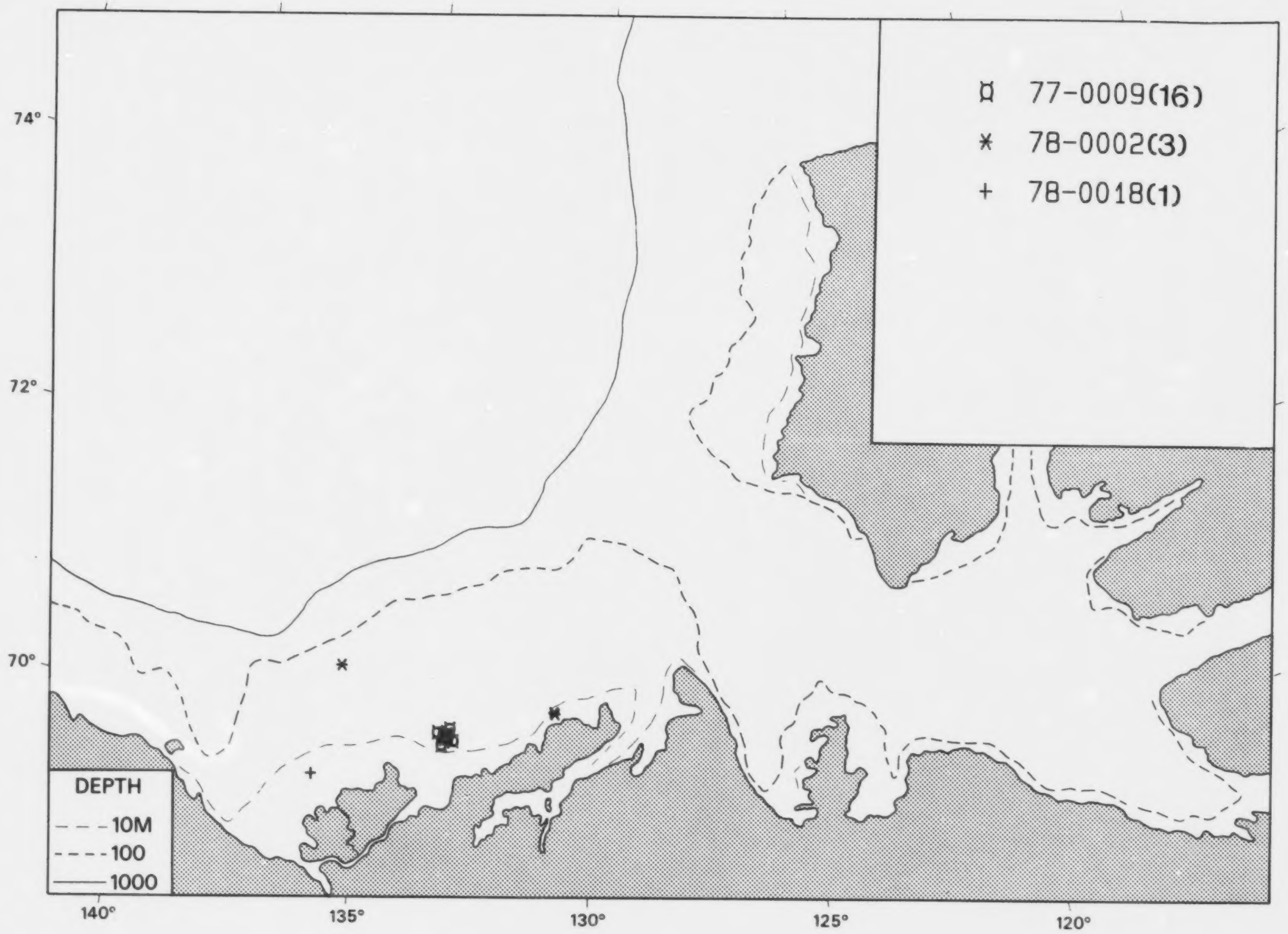
INSTITUTE OF OCEAN SCIENCES, 1982
77-0003(58)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¤ 77-0009(16)
✻ 78-0002(3)
+ 78-0018(1)
DEPTH
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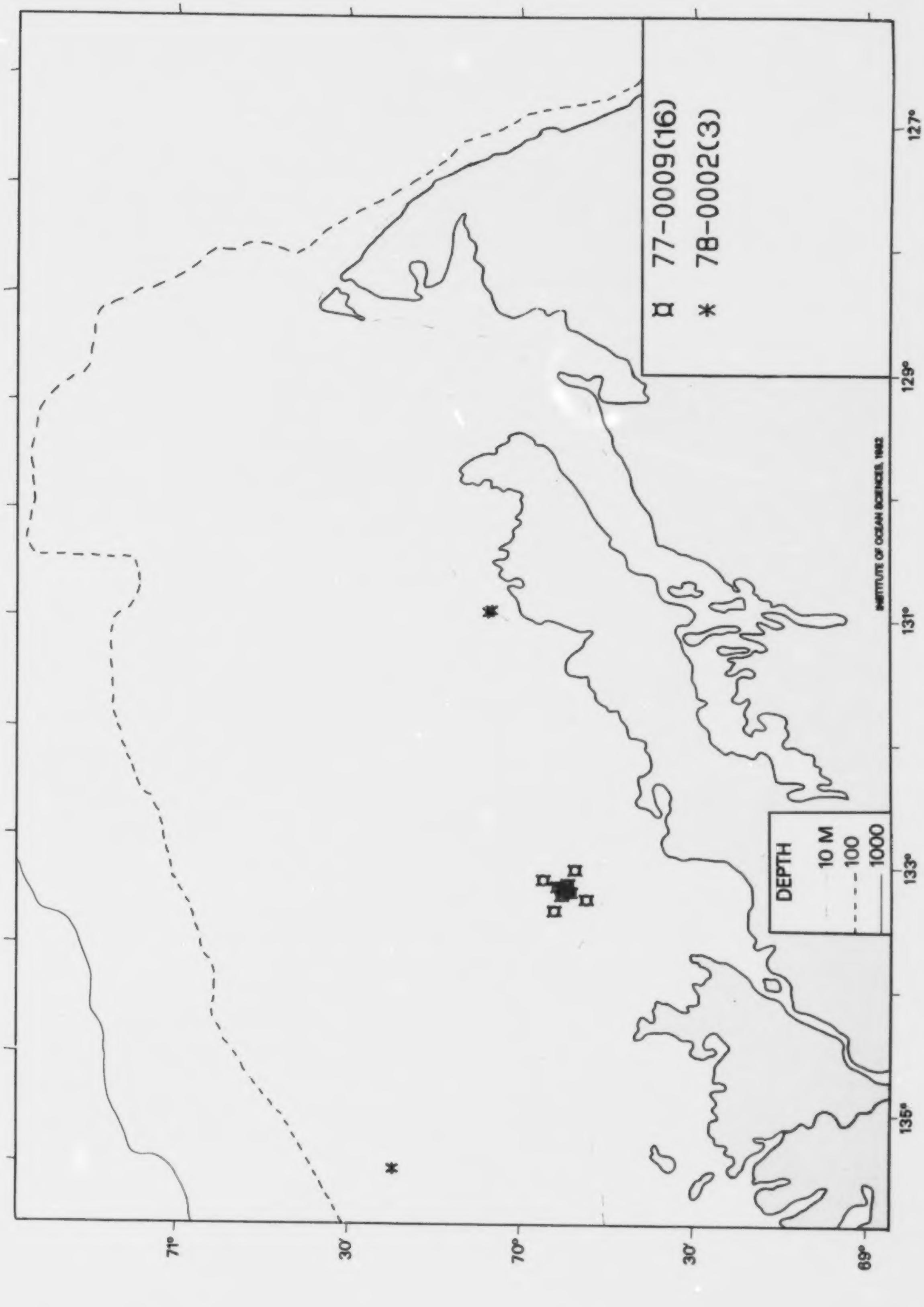
77-0009(16)
78-0002(3)
DEPTH
10 M
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INSTITUTE OF OCEAN SCIENCES, 1982
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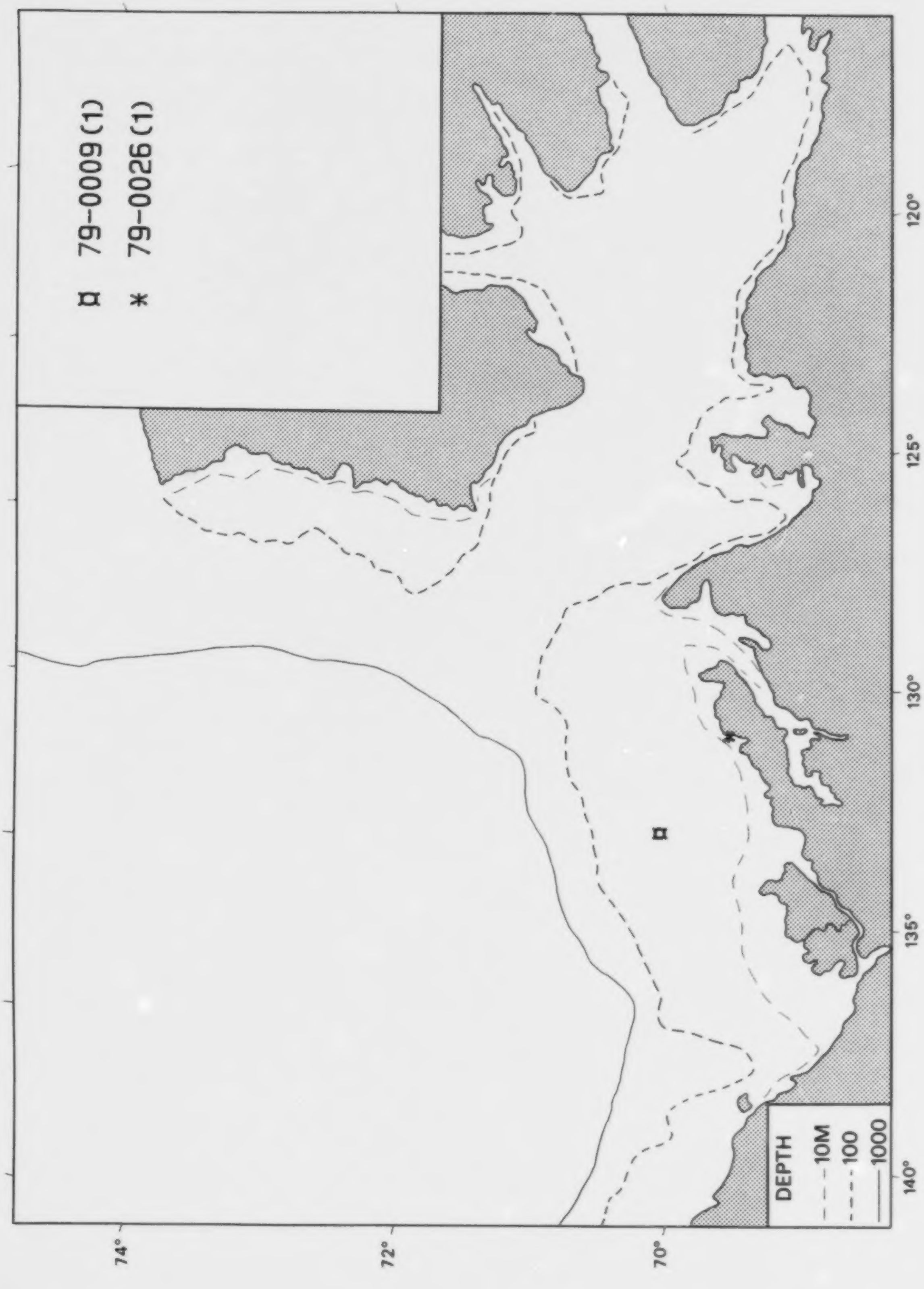
79-0009 (1)
79-0026 (1)
DEPTH
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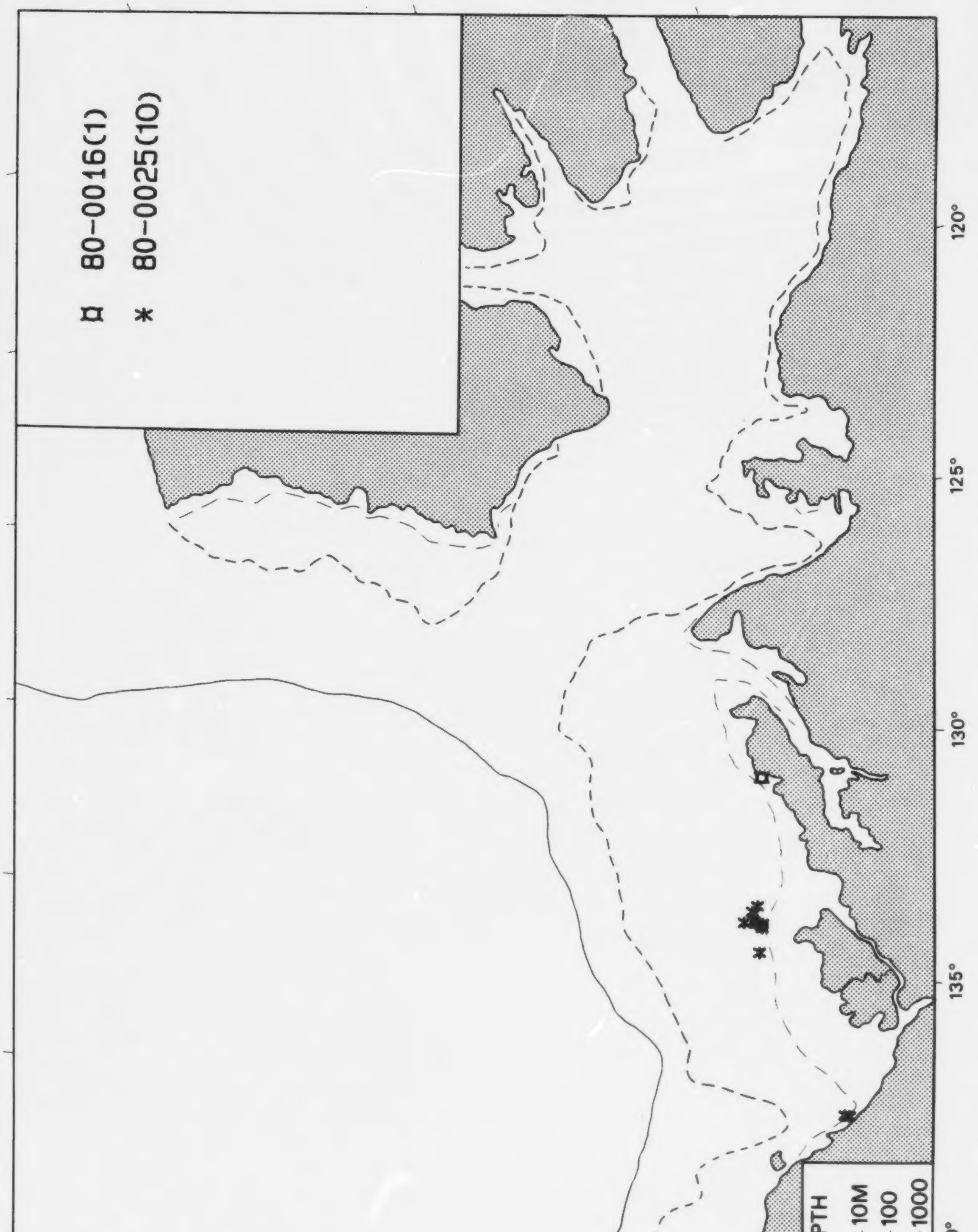
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80-0025(10)
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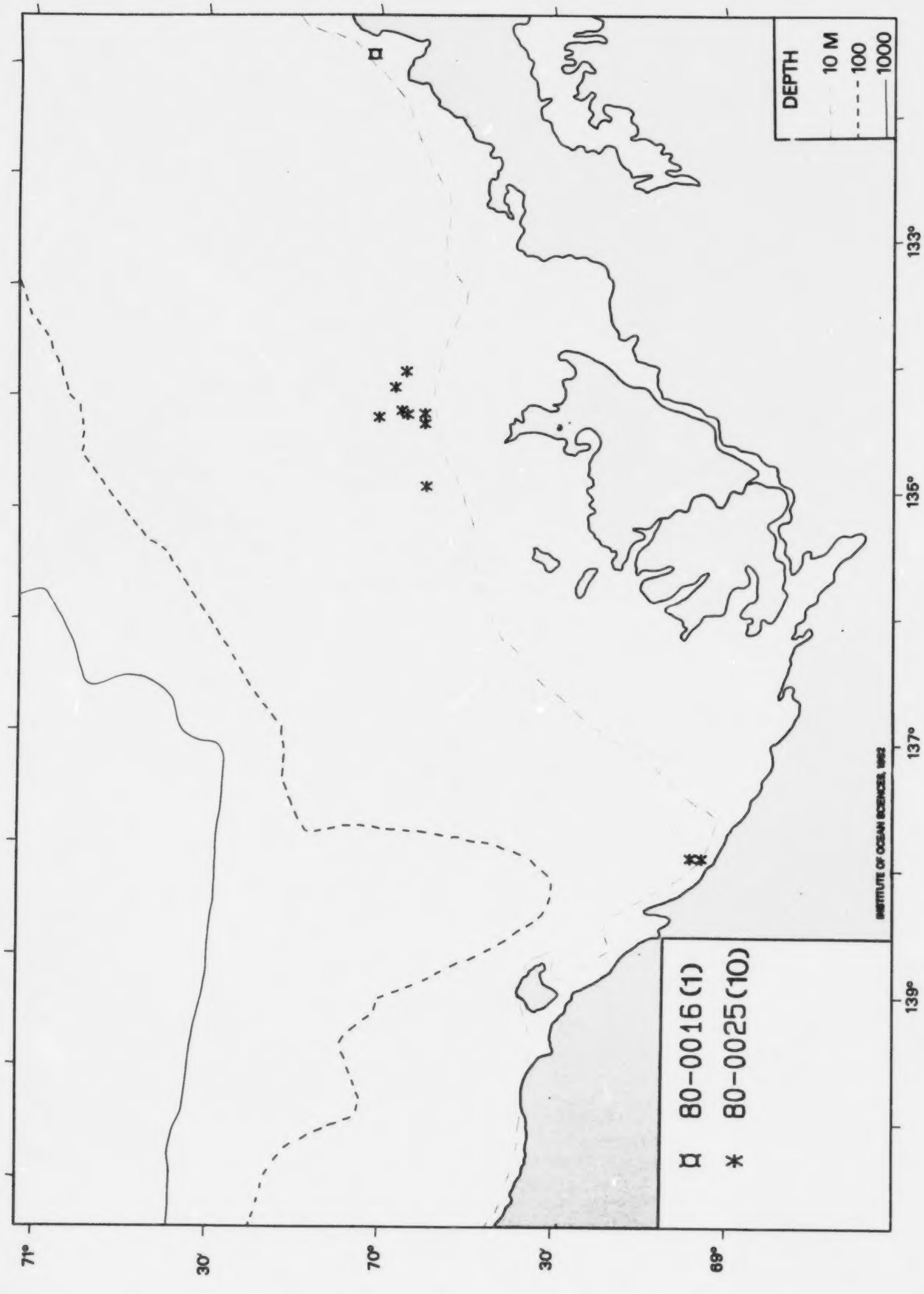
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DEPTH
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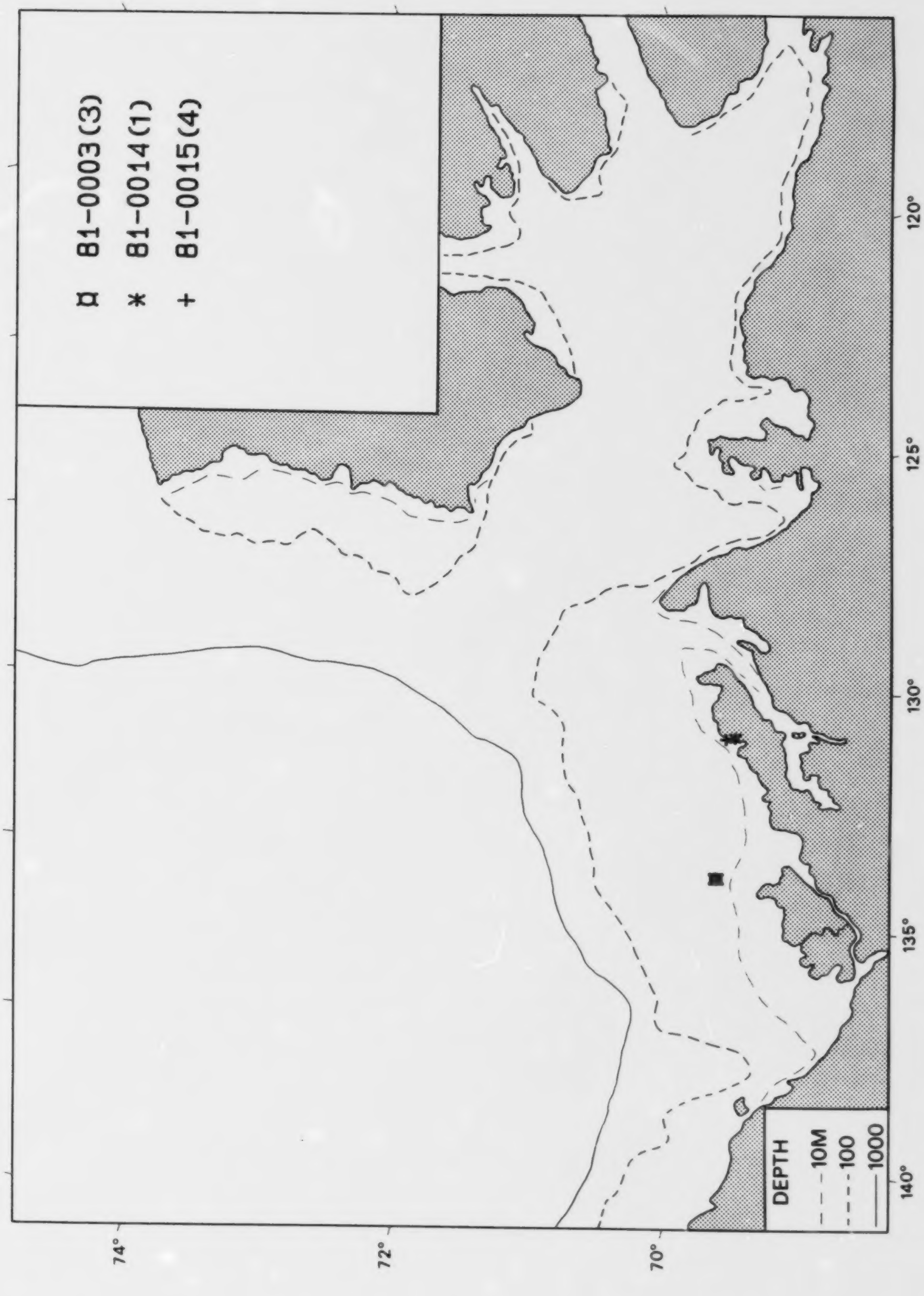
¤ 80-0016 (1)
* 80-0025 (10)
INSTITUTE OF OCEAN SCIENCES, 1982


¤ 81-0003(3)
* 81-0014(1)
+ 81-0015(4)
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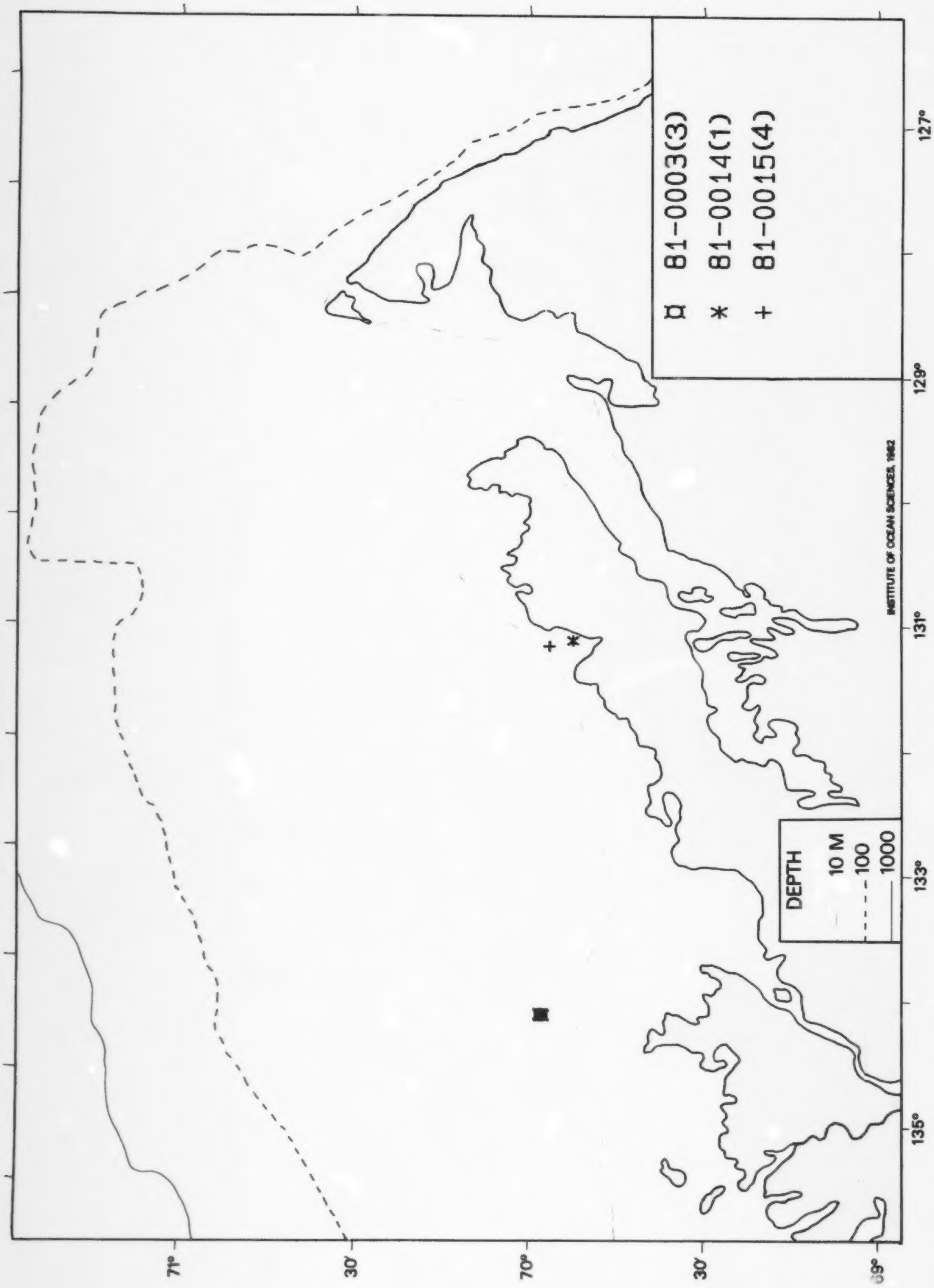
¤ 81-0003(3)
* 81-0014(1)
+ 81-0015(4)
DEPTH
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INSTITUTE OF OCEAN SCIENCES, 1982
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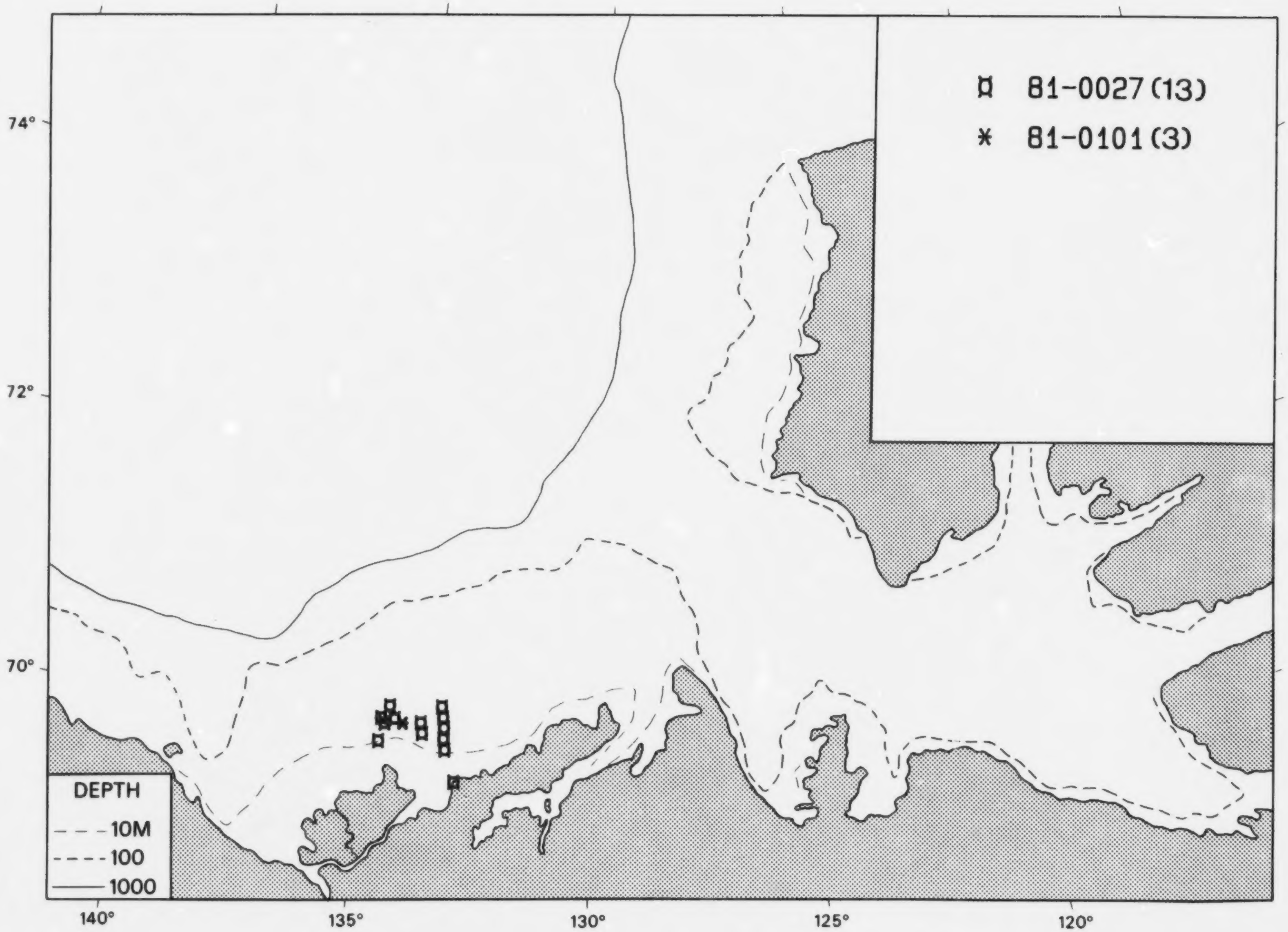
¤ 81-0027 (13)
* 81-0101 (3)
DEPTH
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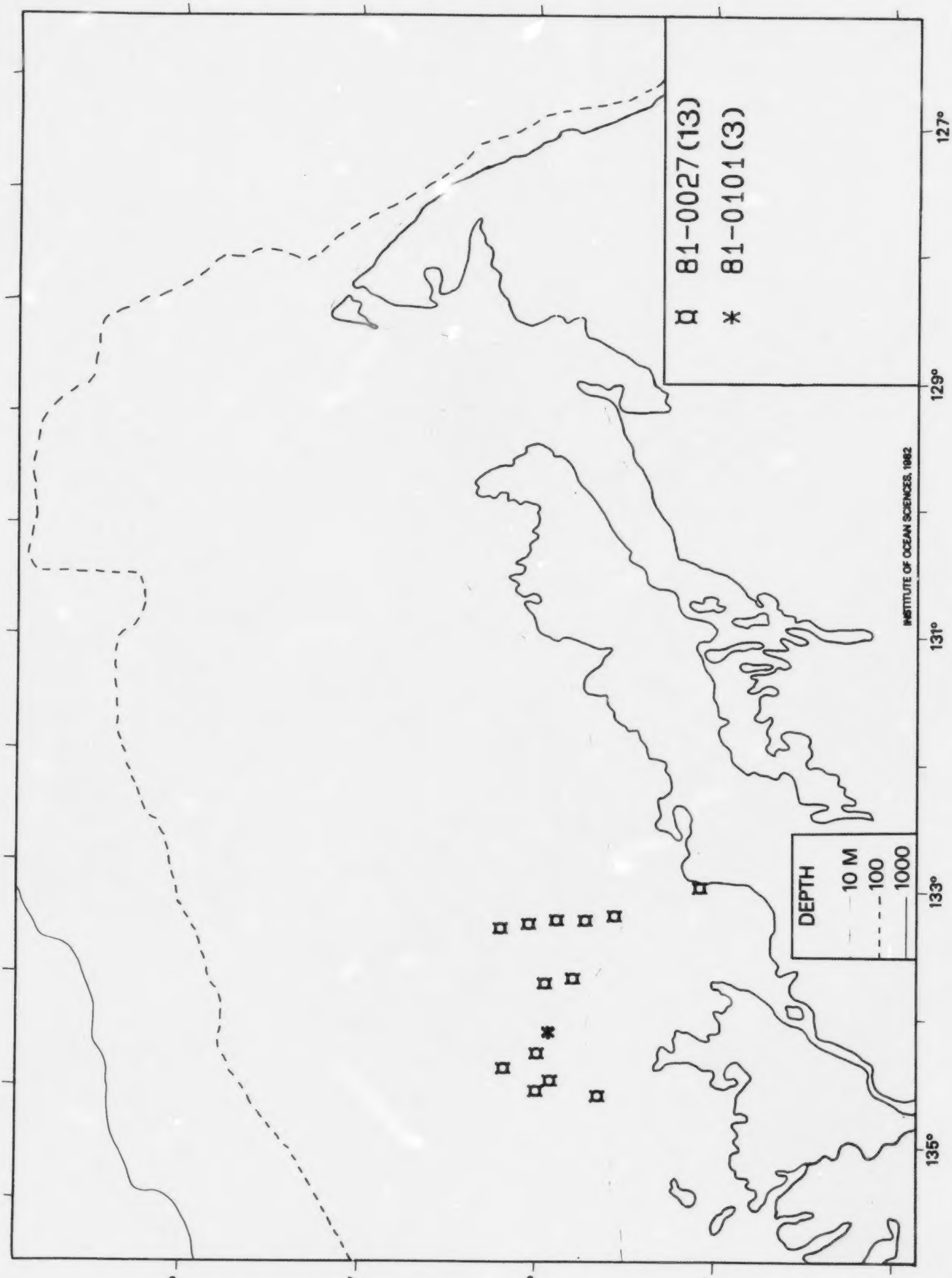

¤ 81-0027 (13)
* 81-0101 (3)
DEPTH
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INSTITUTE OF OCEAN SCIENCES, 1982
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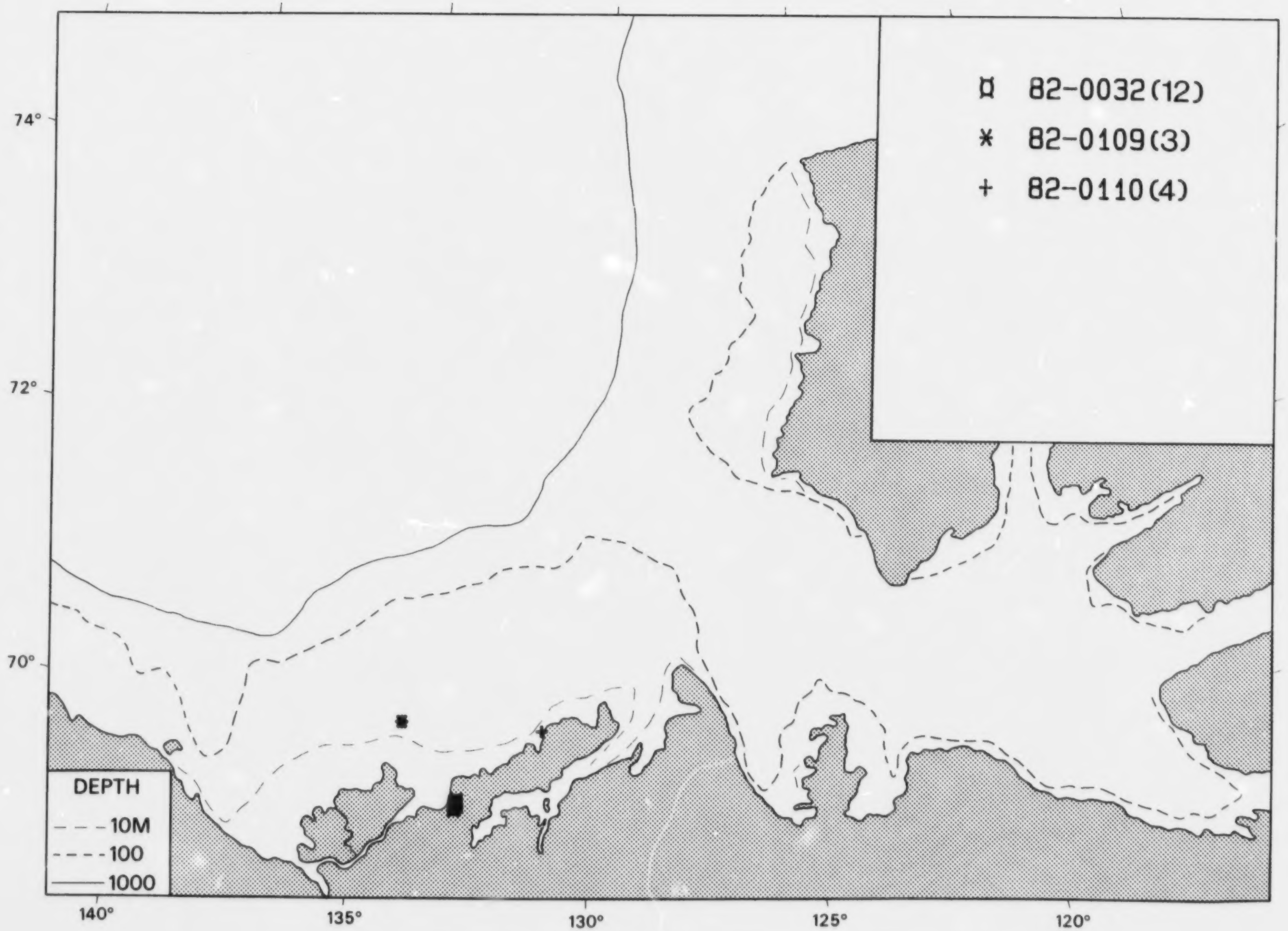
¤ 82-0032(12)
* 82-0109(3)
+ 82-0110(4)
DEPTH
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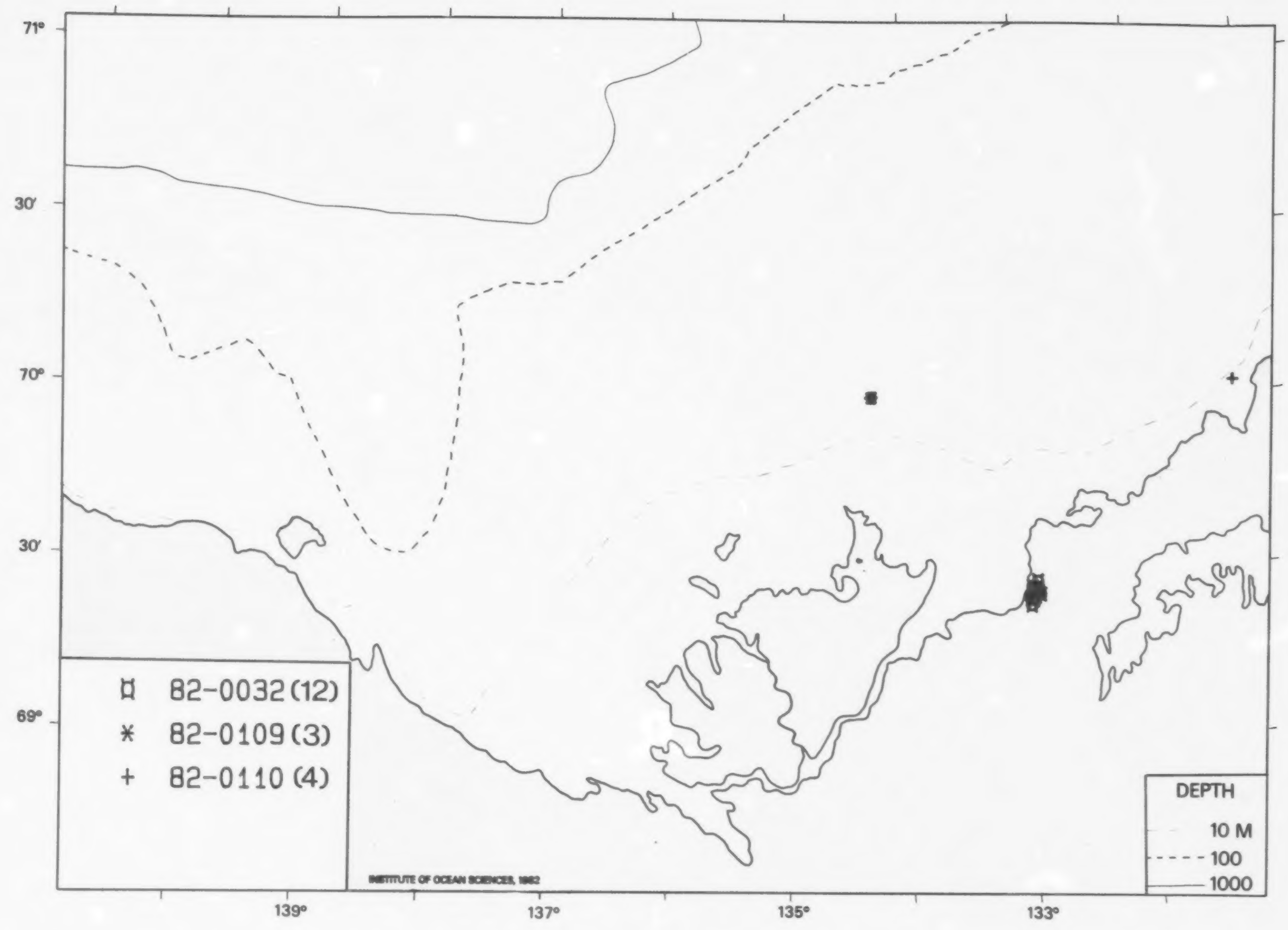
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¤ 82-0032 (12)
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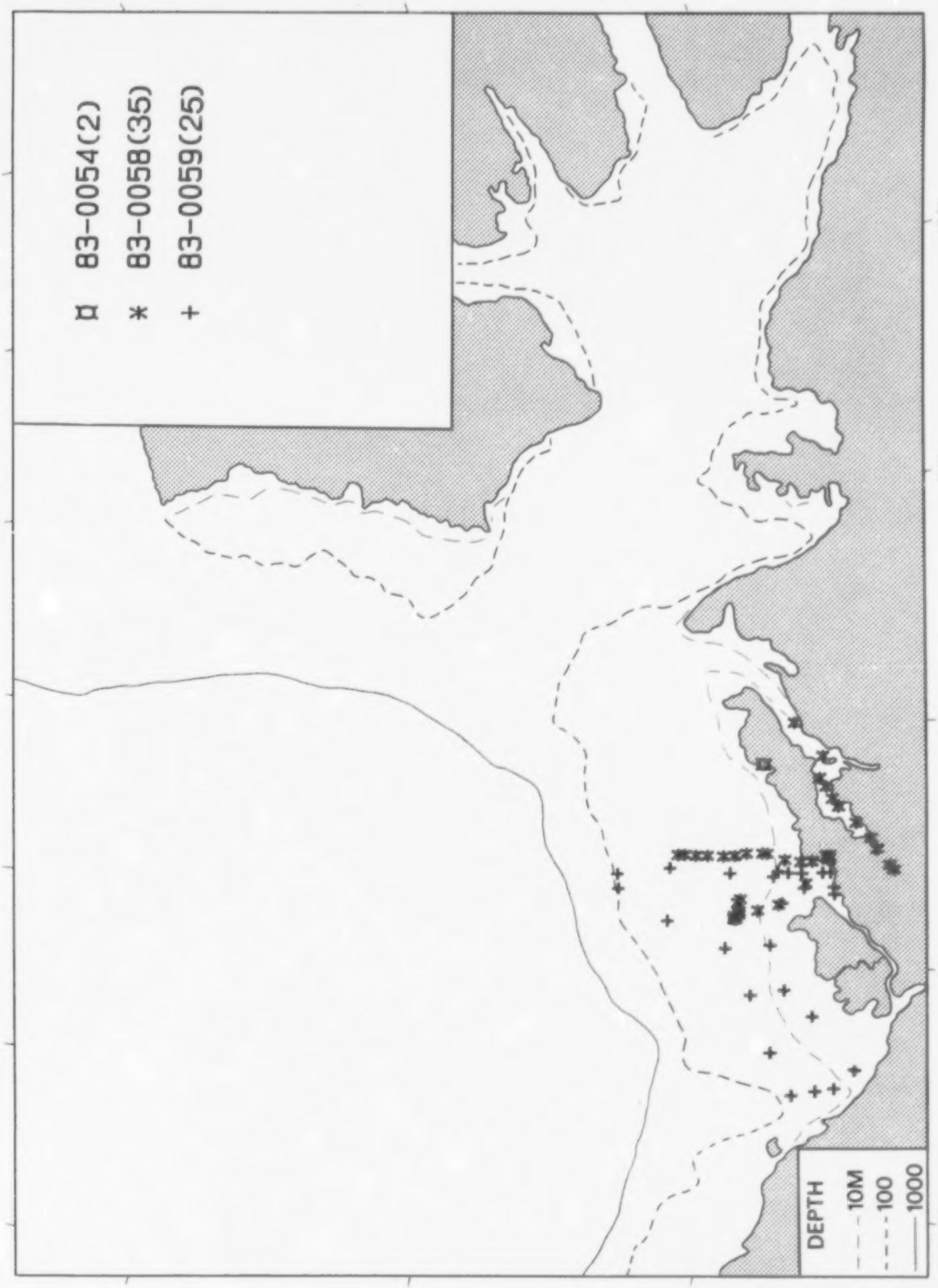
INSTITUTE OF OCEAN SCIENCES, 1982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74°
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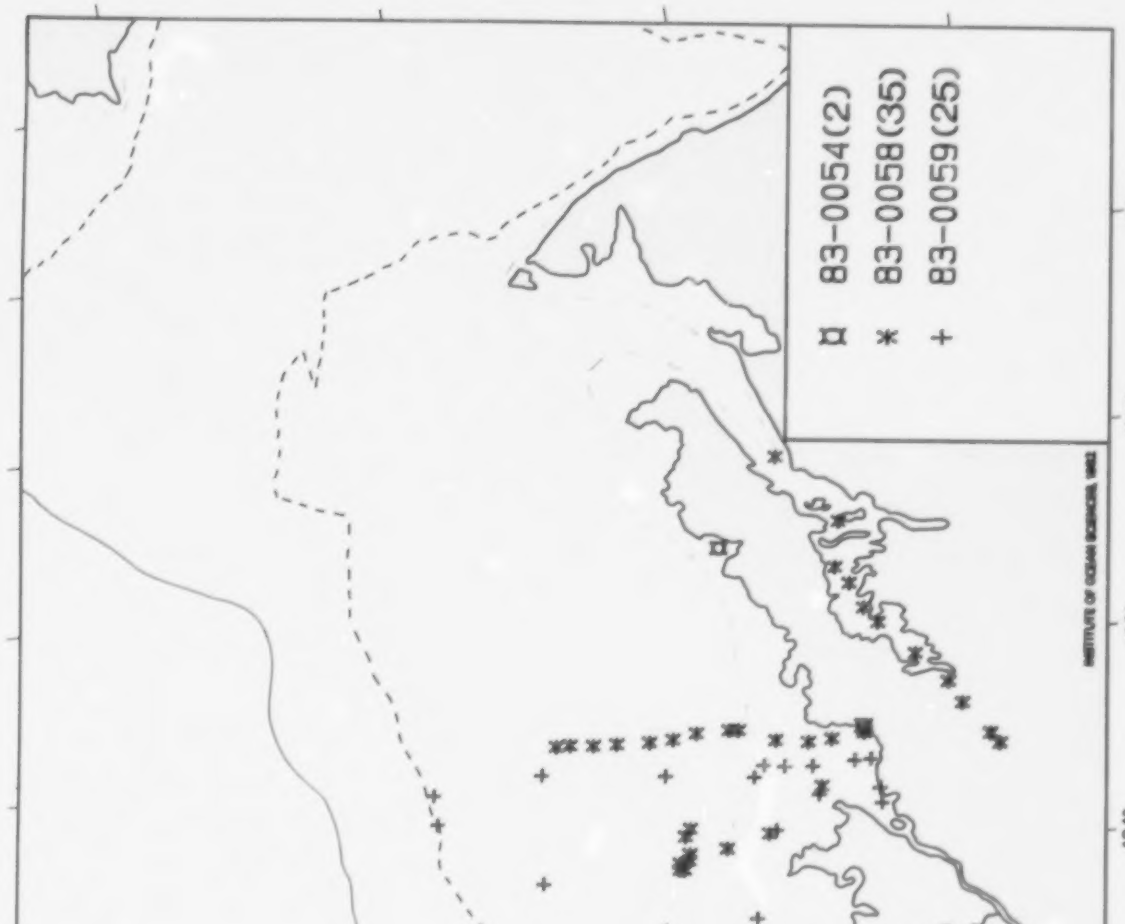
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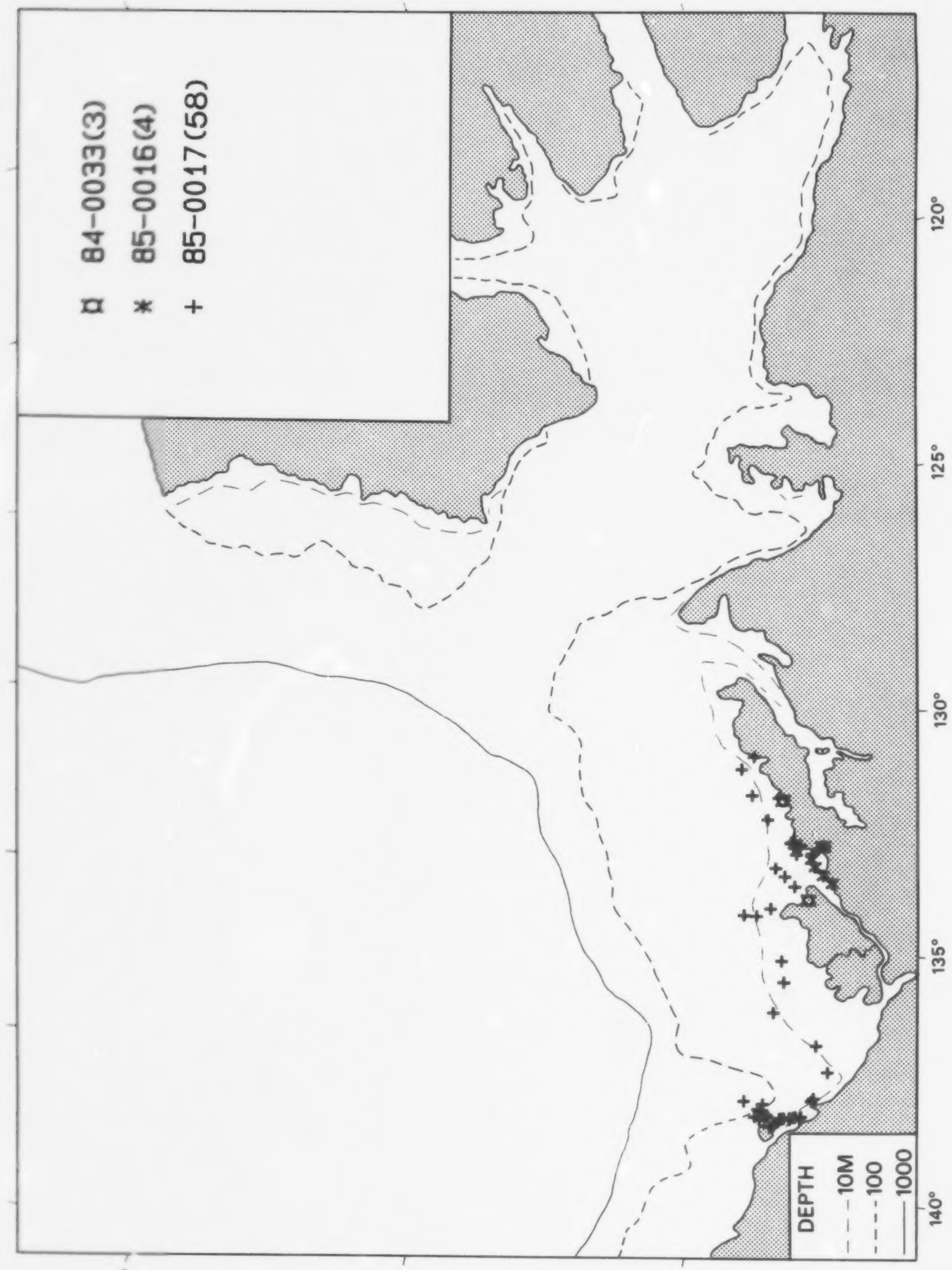

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* 85-0016(4)
+ 85-0017(58)
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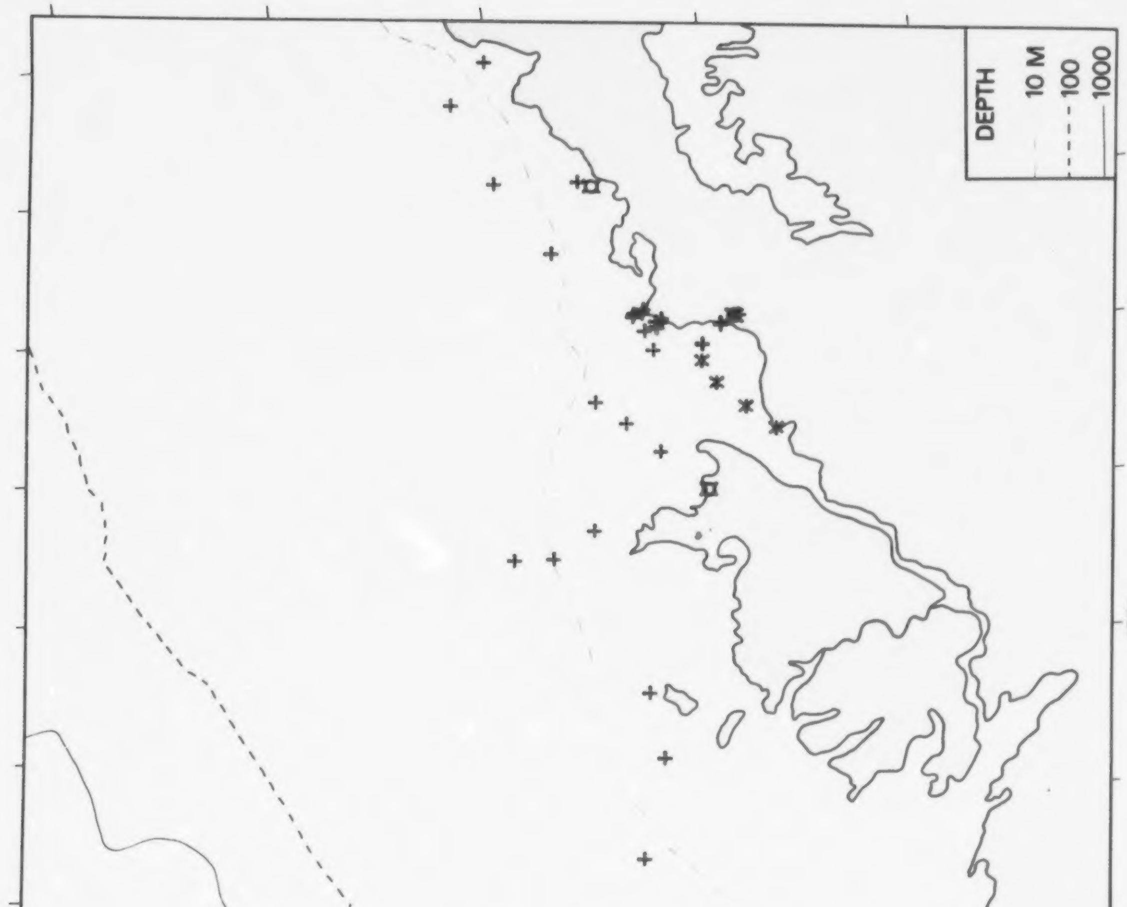
72°
70°
120°
125°
130°
135°
140°



DEPTH
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100
1000
84-0033(3)
85-0016(4)
85-0017(58)
INSTITUTE OF OCEAN SCIENCES, 1982
133°
135°
137°
139°
71°
30'
70°
30'
69°

10. INDICES

10.1 GEOGRAPHIC INDEX
10.2 TAXONOMIC INDEX
10.3 MEASUREMENT INDEX
10.4 DATA REFERENCE/LOCATION INDEX

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70-0002
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75-0006
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71-0001
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81-0003
81-0014
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51-0001
51-0002
52-0002
54-0003

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53-0001
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75-0004
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80-0025
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61-0001 75-0012A
62-0001 77-0003
73-0002 80-0025
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71-0001 75-0012B
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51-0002 73-0003A
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51-0002 73-0002
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61-0001 74-0001
62-0001 74-0007A
69-0066 74-0007C
71-0001 74-0010A
71-0004 74-0011
72-0004 74-0027

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75-0010A 77-0009
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50-0001 75-0006
51-0001 75-0009
51-0002 75-0010A
60-0003 75-0012A
60-0067 75-0012B
61-0001 76-0003
62-0001 76-0004
70-0067 76-0005
71-0001 76-0036
71-0004 77-0002
72-0004 77-0003
73-0002 77-0009
73-0003A 78-0002
73-0003B 78-0018
74-0003 80-0025
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74-0007C 81-0027
74-0010A 82-0110
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51-0002 73-0003B
61-0001 74-0003
62-0001 74-0007A
70-0067 74-0007C
71-0001 74-0010A
71-0004 75-0004
72-0004 75-0010A
73-0002 75-0012A

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76-0036 82-0110
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Cyclopoida
13-0001 74-0007C
50-0001 74-0010A
51-0001 74-0027
51-0002 75-0004
61-0001 75-0010A
62-0001 75-0012A
70-0067 75-0012B
71-0001 76-0003
71-0004 76-0004
72-0004 76-0036
73-0002 77-0003
73-0003A 77-0009
73-0003B 80-0025
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50-0001 74-0007A
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62-0001 76-0003
71-0001 77-0009
71-0004 80-0025
72-0004 81-0027
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Thoracica
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61-0001 74-0007A
62-0001 74-0007C
72-0004 74-0010A
73-0002 75-0012A

Mysidacea
60-0003 75-0004
70-0067 75-0009
71-0001 75-0012B
73-0002 76-0003
73-0003A 78-0002
74-0003 80-0025
74-0007A 81-0015
74-0011 81-0027

Cumacea
71-0001 74-0007C
72-0004 75-0012A
73-0002 80-0025
73-0003A

Isopoda
51-0002 75-0012A
70-0067 80-0025
74-0007A 81-0027
74-0007C 82-0110
74-0011

Amphipoda
50-0001 71-0001
51-0001 72-0004
51-0002 73-0002
60-0003 74-0003
62-0001 74-0007A
70-0067 74-0007C

Amphipoda (cont'd)
74-0011 77-0003
75-0004 77-0009
75-0006 78-0002
75-0009 78-0018
75-0012A 80-0025
75-0012B 81-0015
75-0012E 81-0027
76-0003 82-0110

Euphausiacea
51-0001 76-0003
60-0003 77-0003
73-0002 77-0009
74-0007A 80-0025
74-0007C 81-0027
75-0012A

Decapoda
51-0002 75-0012E
60-0003 77-0003
61-0001 77-0009
62-0001 78-0002
73-0002 80-0025
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62-0001 75-0012B
73-0002 77-0009

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74-0003 76-0036
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Chaetognatha
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60-0003 76-0003
61-0001 77-0003
62-0001 77-0009
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51-0002 75-0012A
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62-0001 76-0003
71-0001 77-0003

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78-0002A 82-0110
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73-0001 75-0012A
73-0003A 75-0012B
73-0003B 75-0028
74-0001 76-0003
74-0003 76-0036
74-0007A 77-0009
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Osteichthyes
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PCHR
(Pacific herring)
82-0032

RNSM
(rainbow smelt)
82-0032

ARCD
(arctic cod)
82-0032

SFCD
(saffron cod)
82-0032

Stichaeidae
81-0015

LUMP
(prickleback)
82-0032

FHSC
(fourhorn sculpin)
82-0032

SC
(sculpin)
75-0012B

STFL
(starry flounder)
82-0032

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62-0001 75-0012E
71-0001 77-0003
72-0004 77-0009
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72-0004 74-0010A
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82-0110

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81-0015

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81-0015 82-0110

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55-0009 79-0037
58-0001 81-0014
61-0079 81-0031
63-0001 83-0054
64-0001 83-0058
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74-0008

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#/Craib core
85-0016

#/Phleger core
74-0010C

#/Petersen grab
75-0010B 75-0012A

#/KB
77-0002

#/NB
79-0009 82-0110
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74-0007C 82-0109
75-0012A

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75-0028 76-0004
76-0003 76-0005

#/PVC
74-0003 75-0004

#/VD
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74-0007C 75-0028
74-0027 76-0036
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75-0012D 85-0016

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84-0033

#/IKT
75-0009

#/NS CBH
51-0002

#/NS N
77-0003

#/NS PN
60-0003

#/NS PNH
51-0002 69-0066

#/NS PNV
51-0002 69-0066

#/NS SNV
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#/C-23 PNV
#/F-23 PNH
13-0001

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#/NS-100 STNH
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#/S-30 PNV
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#/12-20 PNH
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#/50 NNV
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#/72-50 PNH
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#/73?-NS MNH
75-0006

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#/73-30 PNO
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#/79-30 PNV
51-0002

#/80-12.7 BRNH
#/80-12.7 BRNV
76-0036

#/100-30 WNV
74-0019

#/150-50 PNV
77-0009

#/156-10 MNH
76-0004 76-0005

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#/156-50 WNV
76-0003

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81-0015

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#/202-S PNH
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#/202-S PNV
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# tests/202-S PNV
70-0002

#/202-10 MNH or 726-66 PN
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---

* Description of gear type codes (e.g., #/NS) follows this index.

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#/282-30 PNV
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#/505-60 FNH
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width, internal pores
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number, internal pores
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13-0001, 50-0001, 51-0001

shape, head
shape, dorsal antennae
shape, lateral antennae
shape, foot
shape, A1
shape, A2
shape, Mxp2
shape, P1
shape, P2
shape, P4
shape, P5
13-0001

appearance, surface
70-0002

coiling
70-0002, 72-0001

colour
13-0001

age
13-0001, 50-0001, 51-0001, 51-0002, 60-0003, 60-0067, 61-0001, 62-0001, 69-0066, 70-0002, 70-0067, 71-0001, 71-0004, 72-0001, 72-0004, 72-0007, 73-0001, 73-0002, 73-0003A, 73-0003B

10.3 MEASUREMENT INDEX (cont'd)

age (cont'd)

74-0001
74-0003
74-0007A
74-0007C
74-0010A
75-0009
75-0010A
75-0012A
75-0012B
75-0012E
75-0028
76-0036
77-0003
77-0009
78-0002
78-0018
80-0025
81-0015
81-0027
82-0032
82-0110

sex

13-0001
50-0001
51-0002
60-0003
60-0067
61-0001
62-0001
71-0001
71-0004
72-0004
73-0002
73-0003A
74-0007A
74-0007C
74-0010A
75-0009
75-0010A
75-0012A
75-0012B

spermatophores

13-0001

egg number

62-0001

incomplete

52-0002
53-0001
54-0003
55-0009
58-0001
61-0079
63-0001
64-0001
74-0008
74-0119
75-0051
77-0020
79-0037
81-0014
81-0031
83-0054
83-0058
85-0007
85-0017

Description of Gear Type Codes

BN - Bongo Net
BRN - BiRge closing Net
C-23 PN - Coarse-silk conical Plankton Net
CB - Clarke Bumpus net
F-23PN - Finer silk conical Plankton Net
FN - Faber Net
IKT - Isaacs-Kidd midwater Trawl
KB - Kemmerer Bottle
MN - Miller Net
N - Neuston net
NB - Niskin Bottle
NBS - Niskin Bag-Sampler
NN - Nansen-type Net
NS - Not Specified
PB - Plastic Bottle
PN - conical Plankton Net
PP - PhytoPlankton net
PVC - PVC sampler
S corer - Sipre ice corer
S-30 PN - Silk conical Plankton Net
SN - Scor Net
STN - STramin Net
Subsampler - nuclear submarine sampler
VD - Van Dorn sampler
WN - Wisconsin Net

Note: In general, numerical prefixes to the codes indicate mesh size (u) and mouth diameter (cm) of the plankton net. The suffixes H, O and V indicate Horizontal, Oblique and Vertical tows, respectively. Further details are provided in Appendix I.

10.4 DATA REFERENCE/LOCATION INDEX (BY DATA SET NUMBER)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
13-0001	Willey, A. 1919. Marine copepoda. Rep. Can. Arctic Exped., 1913-18, 7(K): 46 p.	NA	
	Bigelow, H.B. 1920. Medusae and ctenophora. Rep. Can. Arctic Exped., 1913-18, 8(H): 23 p.	NA	
	Cushman, J.A. 1920. Foraminifera. Rep. Can. Arctic Exped., 1913-18, 9(M): 13 p.	NA	
	Harring, H.K. 1921. Rotatoria. Rep. Can. Arctic Exped., 1913-18, 8(E): 23 p.	NA	
	Mann, A. 1925. Marine diatoms. Rep. Can. Arctic Exped., 1913-18, 4(F): 31 p.	NA	
	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
50-0001	U.S. Navy Hydrographic Office. 1954. Oceanographic observations, arctic waters, U.S.S. Burton Island, 1950-53. H.O. Pub. 618-C, U.S. Navy Hydrographic Office, Washington, D.C. 309 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Johnson, M.W. 1956. The plankton of the Beaufort and Chukchi Sea areas of the arctic and its relation to the hydrography. Arctic Institute of North America Tech. Paper No. 1: 32 p.	NA	
	Hand, C. and L.B. Kan. 1961. The medusae of the Chukchi and Beaufort Seas of the Arctic Ocean including the description of a new species of Eucodonium (Hydrozoa: Anthomedusae). Arctic Institute of North America Tech. Paper No. 6: 23 p.	NA	
	Johnson, M.W. 1963. Arctic Ocean plankton. p. 173-183. In: Proceedings of the Arctic Basin Symposium, October 1962. Arctic Institute of North America and the Office of Naval Research, U.S. Navy.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
	-	Additional information	Pers. comm.: M. Knight
51-0001	U.S. Navy Hydrographic Office. 1954. Oceanographic observations, arctic waters, U.S.S. Burton Island, 1950-53. H.O. Pub. 618-C, U.S. Navy Hydrographic Office, Washington, D.C. 309 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Johnson, M.W. 1956. The plankton of the Beaufort and Chukchi Sea areas of the arctic and its relation to hydrography. Arctic Institute of North America Tech., Paper No. 1: 32 p.	NA	
	Hand, C. and L.B. Kan. 1961. The medusae of the Chukchi and Beaufort Seas of the Arctic Ocean including the description of a new species of Eucodonium (Hydrozoa: Anthomedusae). Arctic Institute of North America Tech. Paper No. 6: 23 p.	NA	
	Johnson, M.W. 1963. Arctic Ocean plankton. p. 173-183. In: Proceedings of the Arctic Basin Symposium, October 1962. Arctic Institute of North America and the Office of Naval Research, U.S. Navy.	NA	
	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Frost, B.W. 1974. *Calanus marshallae*, a new species of calanoid copepod closely allied to the sibling species *C. finmarchicus* and *C. glacialis*. Mar. Biol. 26:77-99.	NA	
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. *In*: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
	-	Additional information	Pers. comm.: M. Knight
51-0002	Hattersley-Smith, G.F. 1952. Beaufort Sea expedition, 1951. Arctic Circular V(2):13-17.	NA	
	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 519: 54 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
52-0002	U.S. Navy Hydrographic Office. 1954. Oceanographic observations, arctic waters, U.S.S. *Burton Island*, 1950-53. H.O. Pub. 618-C, U.S. Navy Hydrographic Office, Washington, D.C. 309 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
53-0001	U.S. Navy Hydrographic Office. 1954. Oceanographic observations, arctic waters, U.S.S. Burton Island, 1950-53. H.O. Pub. 618-C, U.S. Navy Hydrographic Office, Washington, D.C. 309 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Frost, B.W. 1974. Calanus marshallae, a new species of calanoid copepod closely allied to the sibling species C. finmarchicus and C. glacialis. Mar. Biol. 26:77-99.	NA	
	-	Field notes	Scripps Institution of Oceanography Attn.: M. Knight
	-	Additional information	Pers. comm.: B.W. Frost, M. Knight
54-0003	Maloney, W.L. 1954. Report of the U.S.C.G.C. Northwind (WAGB-282) on the joint Canadian-United States Beaufort Sea expedition, summer 1954. p. 1-35. In: T.D. Jacobs. Joint Canadian-United States Beaufort Sea expedition. Summer 1954. Commander Service Squadron One, Service Force, U.S. Pacific Fleet.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Neave, F. 1954. Western arctic. p. 165-166. In: Annual report of the Pacific Biological Station for 1954. Nanaimo, B.C.	Unpubl. manus.	Pacific Biological Station Attn.: Library
	U.S. Navy Hydrographic Office. 1956. Oceanographic survey results, Beaufort Sea area, summer 1954. U.S.S. Burton Island (AGB-1) and U.S.C.G.C. Northwind (WAGB-282). H.O. 15791. U.S. Navy Hydrographic Office, Washington, D.C. 40 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
55-0009	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
58-0001	Marine Surveys Division, Oceanographic Branch. 1963. Oceanographic data report, arctic, 1958. Informal Manus. Rep. No. 0-26-63. U.S. Naval Oceanographic Office, Marine Sciences Dept., Washington, D.C. 200 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
60-0003	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
60-0067	Grice, G.D. 1962. Copepods collected by the nuclear submarine Seadragon on a cruise to and from the North Pole, with remarks on their geographic distribution. J. Mar. Res. 20(1):97-109.	NA	
	-	Additional information	Pers. comm.: G. Grice
61-0001	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 519: 54 p.	NA	
	LGL Ltd. 1981. An inventory of fisheries information for the waters of the southern Beaufort Sea, 1955-1981. Rep. prep. for Institute of Ocean Sciences, Sidney, B.C. 59 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
61-0079	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
62-0001	Grainger, E.H. 1965. Zooplankton from the Arctic Ocean and adjacent Canadian waters. J. Fish. Res. Bd. Can. 22(2):543-564.	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 519: 54 p.	NA	
	LGL Ltd. 1981. An inventory of fisheries information for the waters of the southern Beaufort Sea, 1955-1981. Rep. prep. for Institute of Ocean Sciences, Sidney, B.C. 59 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
63-0001	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
64-0001	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
69-0066	Burrell, D.C., P.J. Kinney, R.S. Hadley and M.E. Arhelger. 1970. Beaufort Sea environmental data: U.S.C.G.C. *Northwind*, 1968; U.S.C.G.C. *Staten Island*, 1968 and 1969. Univ. Alaska, Inst. Mar. Sci. Rep. No. R70-20: 274 p.	Unpubl. manus.	Univ. of Calgary Attn.: Arctic Institute of North America Library
	-	Additional information	Pers. comm.: R. Horner
70-0002	Vilks, G. 1973. A study of *Globorotalia pachyderma* (Ehrenberg) = *Globigerina pachyderma* (Ehrenberg) in the Canadian arctic. Ph.D. Thesis, Dept. of Oceanography, Dalhousie Univ., Halifax, N.S. 256 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Library
	-	Additional information	Pers. comm.: G. Vilks
70-0067	Sutherland, I. 1982. A collection of zooplankton from Tuktoyaktuk Harbour, Northwest Territories. Can. J. Zool. 60:477-480.	NA	
	-	Additional information	National Museum of Natural Sciences Attn.: D. Faber, L. Marhue
71-0001	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
71-0004	Evans, M.S. and E.H. Grainger. 1980. Zooplankton in a Canadian arctic estuary. p. 199-210. *In:* V.S. Kennedy (ed.). Estuarine perspectives. Academic Press, New York.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
72-0001	Vilks, G. 1973. A study of *Globorotalia pachyderma* (Ehrenberg) = *Globigerina pachyderma* (Ehrinberg) in the Canadian arctic. Ph.D. Thesis, Dept. of Oceanography, Dalhousie Univ., Halifax, N.S. 256 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Library
	-	Additional information	Pers. comm.: G. Vilks
72-0004	Hsiao, S.I.C. 1979. Sea ice microalgal data from the Eskimo Lakes, 1972 to 1974. Fish. Mar. Serv. Data Rep. No. 146: 43 p.	NA	
	Evans, M.S. and E.H. Grainger. 1980. Zooplankton in a Canadian arctic estuary. p. 199-210. *In:* V.S. Kennedy (ed.). Estuarine perspectives. Academic Press, New York.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Hsiao, S.I.C. 1980. Quantitative composition, distribution, community structure and standing stock of sea ice microalgae in the Canadian arctic. Arctic 33(4):768-793.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
72-0007	Slaney, F.F. and Co. Ltd. 1973. 1972 Environmental field program, Taglu-Richards Island, Mackenzie Delta. Interim report. Part 4: Aquatic systems. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 18 p + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Slaney, F.F. and Co. Ltd. 1973a. Environmental impact assessment, Immerk artificial island construction, Mackenzie Bay, N.W.T. Vol. II: Environmental studies. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 59 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Slaney, F.F. and Co. Ltd. 1974. 1972-1974 Environmental program, Mackenzie Delta, N.W.T. Vol. 6: Aquatic resources. Rep. prep. for Imperial Oil Ltd., Gulf Oil Canada Ltd., Shell Canada Ltd. and Canadian Arctic Gas Study Ltd., Calgary, Alta. 111 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	-	Additional information	Pers. comm.: T. Slaney
73-0001	Slaney, F.F. and Co. Ltd. 1974a. Winter benthic and oceanographic surveys, offshore Mackenzie Delta, N.W.T. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 25 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
73-0002	Bunch, J.N. 1974. Distribution and abundance of heterotrophic bacteria in the south Beaufort Sea. Part IIIb. p. 175-195. In: A serien of four reports on the marine ecology of the Mackenzie Delta and Tuktoyaktuk Peninsula region. Environmental-Social Committee, Northern Pipelines, Task Force on Northern Oil Development Rep. No. 74-22.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Bunch, J.N. and R.C. Harland. 1974. Degradation of petroleum by a psychrotolerant marine flora from the Eskimo Lakes. Part I. p. 1-30. In: A series of four reports on the marine ecology of the Mackenzie Delta and Tuktoyaktuk Peninsula region. Environmental-Social Committee, Northern Pipelines, Task Force on Northern Oil Development Rep. No. 74-22.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Grainger, E.H. 1974. Baseline study of the marine environment in the area of the Mackenzie Delta. Part IIIa. p. 101-173. In: A series of four reports on the marine ecology of the Mackenzie Delta and Tuktoyaktuk Peninsula region. Environmental-Social Committee, Northern Pipelines, Task Force on Northern Oil Development Rep. No. 74-22.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p.	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	Bunch, J.N. and R.C. Harland. 1976. Biodegradation of crude petroleum by the indigenous microbial flora of the Beaufort Sea. Beaufort Sea Tech. Rep. No. 10: 52 p.	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution, and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger, S.I.C. Hsiao
73-0003A	Bunch, J.N. and R.C. Harland. 1974. Degradation of petroleum by a psychrotolerant marine flora from the Eskimo Lakes. Part I. p. 1-30. In: A series of four reports on the marine ecology of the Mackenzie Delta and Tuktoyaktuk Peninsula region. Environmental-Social Committee, Northern Pipelines, Task Force on Northern Oil Development Rep. No. 74-22.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p.	NA	
	Hsiao, S.I.C. 1979. Sea ice microalgal data from the Eskimo Lakes, 1972 to 1974. Fish. Mar. Serv. Data Rep. No. 146: 43 p.	NA	
	Hsiao, S.I.C. 1980. Quantitative composition, distribution, community structure and standing stock of sea ice microalgae in the Canadian arctic. Arctic 33(4):768-793.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: J.N. Bunch, E.H. Grainger
73-0003B	Evans, M.S. and E.H. Grainger. 1980. Zooplankton in a Canadian arctic estuary. p. 199-210. In: V.S. Kennedy (ed.), Estuarine perspectives. Academic Press, New York.	NA	
74-0001	Slaney, F.F. and Co. Ltd. 1974a. Winter benthic and oceanographic surveys, offshore Mackenzie Delta, N.W.T. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 25 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
74-0003	Slaney, F.F. and Co. Ltd. 1975. Summer environmental program, Mackenzie River estuary. Vol. 1: Aquatic studies. Rep. prep. for Imperial Oil Ltd. and Sun Oil Co. Ltd., Calgary, Alta. 156 p. + appendix.	Unpubl. manus.	Esso Resourcs Canada Ltd. Attn.: Manager, Frontier Planning
	Slaney, F.F. and Co. Ltd. 1976. Summer environmental program, Mackenzie River estuary. Vol. 1: Aquatic studies. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 152 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
74-0007A	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p.	NA	

10.4 **DATA REFERENCE/LOCATION INDEX** (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Bunch, J.N. and R.C. Harland. 1976. Biodegradation of crude petroleum by the indigenous flora of the Beaufort Sea. Beaufort Sea Tech. Rep. No. 10: 52 p.	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	Hsiao, S.I.C. 1978. Effects of crude oils on the growth of arctic marine phytoplankton. Envir. Pollut. 17:93-107.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	-	Additional information	Pers. comm.: J.N. Bunch, E.H. E.H. Grainger, S.I.C. Hsiao, R. Knowles
74-0007C	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p.	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Percy, J.A. and T.C. Mullin. 1975. Effects of crude oils on arctic marine invertebrates. Beaufort Sea Tech. Rep. No. 11: 169 p.		
	Bunch, J.N. and R.C. Harland. 1976. Biodegradation of crude petroleum by the indigenous flora of the Beaufort Sea. Beaufort Sea Tech. Rep. No. 10: 52 p.	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	Hsiao, S.I.C. 1978. Effects of crude oils on the growth of arctic marine phytoplankton. Envir. Pollut. 17:93-107.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: J.N. Bunch, E.H. Grainger, S.I.C. Hsiao, R. Knowles, J.A. Percy
74-0008	Wong, C.S., W.J. Cretney, R.W. Macdonald and P. Christensen. 1976. Hydrocarbon levels in the marine environment of the southern Beaufort Sea. Beaufort Sea Tech. Rep. No. 38: 113 p.	NA	
	Wong, C.S., R.W. Macdonald, R.D. Bellegay and P. Erickson. 1981. Baseline data on chemical oceanography in the southern Beaufort Sea, 1974-5. Beaufort Sea Tech. Rep. No. 14: 51 p.	NA	
	-	Additional information	Pers. comm.: W.J. Cretney
74-0010A	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	Hsiao, S.I.C. 1979. Sea ice microalgal data from the Eskimo Lakes, 1972 to 1974. Fish. Mar. Serv. Data Rep. No. 146: 43 p.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Hsiao, S.I.C. 1980. Quantitative composition, distribution, community structure and standing stock of sea ice microalgae in the Canadian arctic. Arctic 33(4):768-793.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger, R. Knowles
74-0010C	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	-	Additional information	Pers. comm.: R. Knowles
74-0011	Griffiths, W., P. Craig, G. Walder and G. Mann. 1975. Fisheries investigations in a coastal region of the Beaufort Sea (Nunaluk Lagoon, Yukon Territory). Chapt. II. 219 p. *In*: P.C. Craig (ed.). Fisheries investigations in a coastal region of the Beaufort Sea. Biological Rep. Series. Vol. 34. Rep. prep. by Aquatic Environments Ltd. for Canadian Arctic Gas Study Ltd. and Alaskan Arctic Gas Study Co.	Unpubl. manus.	University of Victoria Attn.: Library
74-0019	Slaney, F.F. and Co. Ltd. 1974b. Winter environmental program: Unark, Pelly and Gary artificial island sites, East Mackenzie Bay, N.W.T. Rep. prep. for Sunoco E & P, Ltd., Calgary, Alta. 34 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
74-0027	NORCOR Engineering and Research Ltd. 1975. The interaction of crude oil with arctic sea ice. Beaufort Sea Tech. Rep. No. 27: 145 p. + appendix.	NA	
	Dickins Assoc. Ltd., D.F., Arctic Laboratories Ltd. and J. Hellebust. 1981. Return to Balaena Bay. Long term effects of a large scale crude oil spill under arctic sea ice, 1975 to 1981. Rep. prep. for Gulf Canada Resources Inc., Calgary, Alta. 127 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
74-0119	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
75-0004	Slaney, F.F. and Co. Ltd. 1976. Summer environmental program, Mackenzie River estuary. Vol. 1: Aquatic studies. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 152 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
75-0006	Wong, C.S., W.J. Cretney, R.W. Macdonald and P. Christensen. 1976. Hydrocarbon levels in the marine environment of the southern Beaufort Sea. Beaufort Sea Tech. Rep. No. 38: 113 p.	NA	
	Wong, C.S., R.W. Macdonald, R.D. Bellegay and P. Erickson. 1981. Baseline data on chemical oceanography in the southern Beaufort Sea, 1974-5. Beaufort Sea Tech. Rep. No. 14: 51 p.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
75-0009	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
75-0010A	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
75-0010B	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	Hsiao, S.I.C., D.W. Kittle and M.G. Foy. 1978. Effects of crude oils and the oil dispersant Corexit on primary production of arctic marine phytoplankton and seaweed. Envir. Pollut. 15:209-221.	NA	
	-	Additional information	Pers. comm.: S.I.C. Hsiao, R. Knowles

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
75-0012A	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	Knowles, R. 1975. Nitrogen fixation in arctic marine sediments. Beaufort Sea Tech. Rep. No. 9: 44 p.	NA	
	Bunch, J.N. and R.C. Harland. 1976. Biodegradation of crude petroleum by the indigenous microbial flora of the Beaufort Sea. Beaufort Sea Tech. Rep. No. 10: 52 p.	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	
	Knowles, R. and C. Wishart. 1977. Nitrogen fixation in arctic marine sediments: effects of oil and hydrocarbon fractions. Envir. Pollut. 13:133-149.	NA	
	Hsiao, S.I.C. 1978. Effects of crude oils on the growth of arctic marine phytoplankton. Envir. Pollut. 17:93-107.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Hsiao, S.I.C., D.W. Kittle and M.G. Foy. 1978. Effects of crude oils and the oil dispersant Corexit on primary production of arctic marine phytoplankton and seaweed. Envir. Pollut. 15:209-221.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger, S.I.C. Hsiao, R. Knowles
75-0012B	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger
	-	Additional information	Pers. comm.: E.H. Grainger
75-0012D	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
75-0012E	Grainger, E.H. 1975. Biological productivity of the southern Beaufort Sea: the physical-chemical environment and the plankton. Beaufort Sea Tech. Rep. No. 12a: 82 p	NA	
	Grainger, E.H. and C. Grohe. 1975. Zooplankton data from the Beaufort Sea, 1951 to 1975. Fish. Mar. Serv. Tech. Rep. No. 591: 54 p.	NA	
	Bunch, J.N. and R.C. Harland. 1976. Biodegradation of crude petroleum by the indigenous microbial flora of the Beaufort Sea. Beaufort Sea Tech. Rep. No. 10: 52 p.	NA	
	Foy, M.G. and S.I.C. Hsiao. 1976. Phytoplankton data from the Beaufort Sea, 1973 to 1975. Fish. Mar. Serv. Tech. Rep. No. 617: 44 p.	NA	
	Hsiao, S.I.C. 1976. Biological productivity of the southern Beaufort Sea: phytoplankton and seaweed studies. Beaufort Sea Tech. Rep. No. 12c: 99 p.	NA	
	Hsiao, S.I.C., M.G. Foy and D.W. Kittle. 1977. Standing stock, community structure, species composition, distribution and primary production of natural populations of phytoplankton in the southern Beaufort Sea. Can. J. Bot. 55(6):685-694.	NA	
	Hsiao, S.I.C. 1978. Effects of crude oils on the growth of arctic marine phytoplankton. Envir. Pollut. 17:93-107.	NA	
	Hsiao, S.I.C., D.W. Kittle and M.G. Foy. 1978. Effects of crude oils and the oil dispersant Corexit on primary production of arctic marine phytoplankton and seaweed. Envir. Pollut. 15:209-221.	NA	
	-	Lab. data sheets	Arctic Biological Station Attn.: E.H. Grainger

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	-	Additional information	Pers. comm.: E.H. Grainger, S.I.C. Hsiao
75-0026	NORCOR Engineering and Research Ltd. 1975. The interaction of crude oil with arctic sea ice. Beaufort Sea Tech. Rep. No. 27: 145 p. + appendix.	NA	
	Dickins Assoc. Ltd., D.F., Arctic Laboratories Ltd. and J. Hellebust. 1981. Return to Balaena Bay. Long term effects of a large scale crude oil spill under arctic sea ice, 1975 to 1981. Rep. prep. for Gulf Canada Resources Inc., Calgary, Alta. 127 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
75-0028	Adams, W.A. 1975. Light intensity and primary productivity under sea ice containing oil. Beaufort Sea Tech. Rep. No. 29: 156 p.	NA	
	-	Additional information	Pers. comm.: W.A. Adams
75-0051	Hunter, J.G. and S.T. Leach. 1983. Station lists of fisheries investigations carried out by the Arctic Biological Station during the years 1947 to 1979. Can. Data Rep. Fish. Aquat. Sci. 413: 220 p.	NA	
76-0003	Slaney, F.F. and Co. Ltd. 1977. Biophysical data appendix. Artificial island sites. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 80 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
	Slaney, F.F. and Co. Ltd. 1977a. 1976 Summer aquatic studies: Arnak L-30 artificial island site and Tuft Point borrow site. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 138 p. + appendix.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Library

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
76-0004	Duval, W.S. 1977a. Planktonic communities. Part 3.2. p. 18-25. In: F.F. Slaney and Co. Ltd. 1976 Summer aquatic studies: Arnak L-30 artificial island site and Tuft Point borrow site. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Library
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.
76-0005	Duval, W.S. 1977. Planktonic communities. Part 2.2. p. 62-86. In: F.F. Slaney and Co. Ltd. 1976 Summer aquatic studies: Arnak L-30 artificial island site and Tuft Point borrow site. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Library
	Duval, W.S. 1977b. Environmental assessment of construction and construction support activities on planktonic communities of the Mackenzie Estuary related to the proposed ten year Beaufort Sea offshore exploration program. Part 4. 78 p. In: F.F. Slaney and Co. Ltd. Environmental assessment of construction and construction support activities related to the proposed ten year Beaufort Sea offshore exploration program. Vol. 2. Rep. prep. for Imperial Oil Ltd., Calgary, Alta.	Unpubl. manus.	Esso Resources Canada Ltd.

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
76-0036	Martin, L.C., V.A. Poulin and G. Bradley. 1976. Lake and stream catalogue, Hans Bay region, N.W.T. Rep. prep. by F.F. Slaney and Co. Ltd. for Gulf Oil Canada Ltd., Calgary, Alta. 6 p. + catalogue sheets.	Unpubl. manus.	Environmental Sciences Ltd. Attn.: L.C. Martin
	Poulin, V.A. and L.C. Martin. 1976. Biological studies, Hans Bay region, N.W.T., 1976. Part 3. p. 15-35 + tables. In: F.F. Slaney and Co. Ltd. Environmental studies, Hans Bay region, 1976. Vol. 1: Aquatic studies. Rep. prep. for Gulf Oil Canada Ltd.	Unpubl. manus.	Environmental Sciences Ltd. Attn.: L.C. Martin
	Duval, W.S. 1977c. Planktonic communities of the Parsons Lake region. Part 4. p. 53-94. In: F.F. Slaney and Co. Ltd. Investigations of the aquatic resources of the Parsons Lake region and environmental impact assessment of the Parsons Lake gas development. Rep. prep. for Gulf Oil Canada Ltd., Calgary, Alta.	Unpubl. manus.	Environmental Sciences Ltd. Attn.: W.S. Duval
77-0002	Jones, M.L. and J. Den Beste. 1977. Tuft Point and adjacent coastal areas fisheries project. Rep. prep. by Aquatic Environments Ltd. for Imperial Oil Ltd., Calgary, Alta. 152 p.	Unpubl. manus.	University of Calgary Attn.: Arctic Institute of North America Library
77-0003	Macdonald, R.W., M.E. McFarland, S.J. De Mora, D.M. Macdonald and W.K. Johnson. 1978. Oceanographic data report, Amundsen Gulf, August -September 1977. Pac. Mar. Sci. Rep. 78-10: 92 p.	NA	
77-0009	Envirocon Ltd. 1977. Isserk artificial island environmental baseline and monitoring study, 1977. Rep. prep. for Imperial Oil Ltd., Calgary, Alta. 125 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
77-0020	Hillaby, B. 1977. Field notes of fisheries investigations at King Point Lagoon, Yukon Territory. Rep. prep. for Department of Fisheries and Oceans, Vancouver, B.C.	Unpubl. manus.	Freshwater Institute Attn.: W. Bond

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
78-0002	Thomas, D.J. 1978. Tingmiark K-91 and Kopanoar D-14: a chemical study one year after the occurrence of water flow, July 1978. Rep. prep. by Seakem Oceanography Ltd. for Canadian Marine Drilling Ltd., Calgary, Alta. 126 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
78-0018	Thomas, D.J. 1978a. A baseline chemical survey at Tarsuit A-25, July 1978. Rep. prep. by Seakem Oceanography Ltd. for Canadian Marine Drilling Ltd., Calgary, Alta. 35 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
79-0009	Thomas, D.J. 1979. The effect of discharged drilling fluid waste on primary productivity at Nerlerk M98. Rep. prep. by Arctic Laboratories Ltd. for Canadian Marine Drilling Ltd., Calgary, Alta. 31 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
79-0026	Acreman, J., G. Borstad and B. Humphrey. 1980. Dome Petroleum experimental oil spill at McKinley Bay, N.W.T. Examination of ice biota. Rep. prep. by Arctic Laboratories Ltd. for Dome Petroleum Ltd., Calgary, Alta. 16 p.	Unpubl. manus.	Dome Petroleum Ltd.
	Dome Petroleum Ltd. 1981. Oil and gas under sea ice study. Vol. 1. 286 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division
79-0037	-	Information	Pers. comm.: M. Lawrence
80-0016	Acreman, J., G. Borstad and B. Humphrey. 1980. Dome Petroleum experimental oil spill at McKinley Bay, N.W.T. Examination of ice biota. Rep. prep. by Arctic Laboratories Ltd. for Dome Petroleum Ltd., Calgary, Alta. 16 p.	Unpubl. manus.	Dome Petroleum Ltd.
	Dome Petroleum Ltd. 1981. Oil and gas under sea ice study. Vol. 1. 286 p.	Unpubl. manus.	Institute of Ocean Sciences Attn.: Scientific Information Division

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
80-0025	Griffiths, W.B. 1981. Characteristics of bowhead feeding areas. p. 197-264. In: W.J. Richardson (ed.). Behavior, disturbance responses and feeding of bowhead whales in the Beaufort Sea, 1980. Rep. prep. by LGL Ecological Research Associates Inc. for U.S. Bureau of Land Management, Washington.	Unpubl. manus.	LGL Ltd. Attn.: W.J. Richardson
	Griffiths, W.B. and R.A. Buchanan. 1982. Characteristics of bowhead feeding areas. p. 347-455. In: W.J. Richardson (ed.). Behavior, disturbance responses and feeding of bowhead whales Balaena mysticetus in the Beaufort Sea, 1980-81. Rep. prep. by LGL Ecological Research Associates Inc. for U.S. Bureau of Land Management, Washington.	Unpubl. manus.	LGL Ltd. Attn.: W.J. Richardson
81-0003	Bunch, J.N., F. Dugre and T. Cartier. 1983. Issungnak oceanographic survey. Part C: Microbiology. Rep. prep. for Esso Canada Resources Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd., Calgary, Alta. 39 p.	Unpubl. manus.	Arctic Laboratories Ltd. Attn.: P. Erickson
81-0014	-	Field notes	Seakem Oceanography Ltd. Attn.: P. Wainwright
	-	Unanalysed samples	Environmental Protection Service, Yellowknife, N.W.T.
81-0015	Pett, R.J., G.P. Vickers, J.C. Acreman and A.G. Ethier. 1983. A seasonal study of the epontic and planktonic communities near the Issungnak artificial island and McKinley Bay in the Canadian Beaufort Sea. Rep. prep. by Arctic Laboratories Ltd. for Fisheries and Oceans Canada. 89 p. + appendix.	Unpubl. manus.	Arctic Biological Station Attn.: Library
81-0027	Griffiths, W.B. and R.A. Buchanan. 1982. Characteristics of bowhead feeding areas. p. 347-455. In: W.J. Richardson (ed.). Behavior, disturbance responses and feeding of bowhead whales Balaena mysticetus in the Beaufort Sea, 1980-81. Rep. prep. by LGL Ecological Research Associates Inc. for U.S. Bureau of Land Management, Washington.	Unpubl. manus.	LGL Ltd. Attn.: W.J. Richardson

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
81-0031	-	Information	Pers. comm.: M. Lawrence
81-0101	Bunch, J.N., F. Dugre and T. Cartier. 1983. Issungnak oceanographic survey. Part C: Microbiology. Rep. prep. for Esso Canada Resources Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd., Calgary, Alta. 39 p.	Unpubl. manus.	Arctic Laboratories Ltd. Attn.: P. Erickson
82-0032	Ratynski, R.A. 1983. Midsummer ichthyoplankton populations of Tuktoyaktuk Harbour, N.W.T. Can. Tech. Rep. Fish. Aquat. Sci. 1218: 21 p.	NA	
82-0109	Bunch, J.N., F. Dugre and T. Cartier. 1983. Issungnak oceanographic survey. Part C: Microbiology. Rep. prep. for Esso Canada Resources Ltd., Gulf Canada Resources Inc. and Dome Petroleum Ltd., Calgary, Alta. 39 p.	Unpubl. manus.	Arctic Laboratories Ltd. Attn.: P. Erickson
82-0110	Pett, R.J., G.P. Vickers, J.C. Acreman and A.G. Ethier. 1983. A seasonal study of the epontic and planktonic communities near the Issungnak artificial island and McKinley Bay in the Canadian Beaufort Sea. Rep. prep. by Arctic Laboratories Ltd. for Fisheries and Oceans Canada. 89 p. + appendix.	Unpubl. manus.	Arctic Biological Station Attn.: Library
83-0054	-	Field notes	Seakem Oceanography Ltd. Attn.: P. Wainwright
	-	Unanalysed samples	Environmental Protection Service, Yellowknife, N.W.T.
83-0058	Borstad, G.A. 1985. Water colour and temperature in the southern Beaufort Sea: remote sensing in support of ecological studies of the bowhead whale. Can. Tech. Rep. Fish. Aquat. Sci. No. 1350: 68 p.	NA	
	-	Field notes	G.A. Borstad Ltd. Attn.: G.A. Borstad

10.4 DATA REFERENCE/LOCATION INDEX (cont'd)

DATA SET I.D.	REFERENCE	STATUS	AVAILABILITY
83-0059	Albright, L.J. 1986. Bacterial dynamics within the lower Mackenzie River, its estuary and the southern Beaufort Sea. Can. J. Microbiol.: in press.	In press	Simon Fraser University Attn.: L.J. Albright
84-0033	Nalewajko, C. 1985. Availability of cadmium in sediments and its effects on phytoplankton. Rep. prep. for Environmental Protection Service, Yellowknife, N.W.T. 23 p. + figures.	Unpubl. manus.	Environmental Protection Service Attn.: P. Mudroch
	-	Additional information	Pers. comm.: P. Mudroch, C. Nalewajko
85-0007	(Data analysis in progress at time this inventory prepared.)	Field notes	LGL Ltd. Attn.: D. Thomson
85-0016	Albright, L.J. 1986. Bacterial dynamics within the lower Mackenzie River, its estuary and the southern Beaufort Sea. Can. J. Microbiol.: in press.	In press	Simon Fraser University Attn.: L.J. Albright
85-0017	(Data analysis in progress at time this inventory prepared.)	Field data notes	Freshwater Institute Attn.: G. Hopky

APPENDIX 1

DESCRIPTION OF GEAR TYPES

DESCRIPTION OF GEAR TYPES

BACTERIA (other than water samplers)

NS - sampler not specified
Craib corer - no description given
Phleger corer - no description given
Petersen grab - no description given

BACTERIA/PLANKTON (water samplers)

KB - Kemmerer bottle
NB - Niskin bottle
NBS - Niskin ss 1.5 sterile bag-sampler
PB - plastic bottle
PVC - sampler constructed from PVC
VD - Van Dorn sampler
Bucket - no description given

PLANKTON (nets)

In general, these codes are interpreted as follows:

> 'mesh size (u) - mouth diameter (cm) - type of net and whether horizontal/oblique/vertical tow'
>
> (e.g., 60-25 PNH = 60 u mesh - 25 cm mouth diameter - conical plankton net and horizontal tow)

Further details are provided when the code deviates from the above. In all cases, however, H or O or V at the end of the code indicates a horizontal or oblique or vertical tow.

IKT - Isaacs-Kidd midwater trawl
NS CBH - Clarke Bumpus net, mesh not specified
NS N - neuston net, mesh not specified
NS PN - plankton net, no description given
NS PNH - plankton net, no description given
NS PNV - plankton net, no description given
NS PPH - phytoplankton net, no description given
NS SNV - Scor net
Subsampler - specially-designed sampler for nuclear submarine
C-23 PNH - conical plankton net, coarse silk, 23 cm mouth diameter, 50 cm long, closed at one end, "Net #3"
C-23 PNV - conical plankton net, description as above
F-23 PNH - conical plankton net, description as above but finer silk, "Net #4"
NS-12 WNV - Wisconsin net, mesh not specified, 12 cm mouth diameter

NS-50 PN - conical plankton net, mesh not specified, 50 cm mouth diameter
NS-50 PNV - conical plankton net, mesh not specified, 50 cm mouth diameter
NS-100 STNH - Stramin net, mesh not specified, 100 cm mouth diameter
S-30 PNV - silk plankton net, mesh not specified, 30 cm mouth diameter, 7.5 cm cod end diameter, 105 cm long, upper half canvas, lower half silk, "Net #5"
12-20 PNH - conical plankton net
50 NNV - Nansen-type plankton net, upper half No. 2 bolting silk, lower half No. 10 bolting silk, 50 cm mouth diameter
60-25 PNH - conical plankton net
60-25 PNV - conical plankton net
72-50 PNH - conical plankton net, 2.5 m long, 5:1 conical taper
73?-NS MNH - Miller net, 73u(?) mesh ("No. 20")
73?-12 WNV - Wisconsin net, 73u(?) mesh ("No. 20")
73?-17.5 PNV - conical plankton net, 73u(?) mesh ("No. 20")
73-30 PNH - conical plankton net, ("No. 20"), 3:1 conical taper
73-30 PNO - conical plankton net, ("No. 20"), 3:1 conical taper
73-30 PNV - conical plankton net, ("No. 20"), 3:1 conical taper
75-C PNV - plankton net, collapsible umbrella frame, 75 u mesh, approximately 2.5 m long
75-NS MNH - Miller net, 75 u mesh
76-50 NNV - Nansen-type plankton net
79-30 PNH - conical plankton net, 79 u mesh ("No. 18")
79-30 PNV - conical plankton net, 79 u mesh ("No. 18")
80-12.7 BRNH - Birge closing plankton net
80-12.7 BRNV - Birge closing plankton net
83-50 BNO - Bongo net
83-50 BNV - Bongo net
100-30 WNV - Wisconsin plankton net
150-50 PNV - conical plankton net
156-10 MNH - Miller net
156-50 WNH - Wisconsin net
156-50 WNV - Wisconsin net
158-22.5 PNO - conical plankton net
158-22.5 PNV - conical plankton net
202-C PNV - conical plankton net, collapsible umbrella frame, 202 u mesh, approximately 2.5 m long
202-S PNH - conical plankton net, square mouth (50 cm on the side), 202 u mesh, approximately 2.5 m long
202-S PNV - conical plankton net, description as above
202-10 MNH - Miller net
233-30 PNV - conical plankton net
233-50 PNH - conical plankton net, 233 u mesh ("No. 6")
233-50 PNV - conical plankton net, 233 u mesh ("No. 6")
240-50 PNH - conical plankton net
240-50 PNV - conical plankton net
243-NS MNH - Miller net, 243 u mesh
253-75 PNV - conical plankton net
282-30 PNH - conical plankton net, 282 u mesh ("No. 5")
282-30 PNV - concial plankton net, 282 u mesh ("No. 5")
392-60 FNH - Faber net, two circular steel hoops around net (upper 60 cm, lower 51 cm diameter), approximately 3 m long

500-NS MNH - Miller net
500-NS N - neuston net, 500 u mesh
500-50 BNH - Bongo net
500-50 BNO - Bongo net
500-61 PNV - conical plankton net
505-60 FNH - Faber net, two circular steel hoops (outer 60 cm, lower 51 cm diameter), approximately 3 m long
505-60 PNH - conical plankton net
569-50 PNO - conical plankton net
571-50 PNH - conical plankton net, 571 u mesh ("No. 0")
571-50 PNV - conical plankton net, 571 u mesh ("No. 0")
571-100 PNH - conical plankton net, 571 u mesh ("No. 0")
726-43 PN - conical plankton net, approximately 3 m long
726-66 PN - conical plankton net, approximately 3 m long
750-50 WNH - Wisconsin net
1000-100 PNH - conical plankton net

EPONTIC COMMUNITY

NS corer - ice corer, not specified
S corer - Sipre ice corer
chainsaw (block) - block cut by chainsaw from a dive hole
ice scraper (scraping) - device made from plastic funnel (25 cm mouth diameter) and plankton net attached to hinged aluminum rod, a piece of styrofoam attached to scraper provides positive buoyancy to force apparatus against under-ice surface

APPENDIX 2

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